

115)

Institute of Pharmacy, University, Regensburg

Colour reaction of cardiotonic cardenolid glycosides with sodium nitroprusside

A. F. VOGT and W. WIEGREBE

Dedicated Prof. Dr. F. Eiden, München, with warm regards on the occasion of his 65th birthday

Digitoxin (**1**) and digitoxigenin (**2**) react with sodium nitroprusside [disodium-pentakis(cyano-C)-nitrosyl-ferrate(II)] under weakly basic conditions forming the corresponding complexes **3** and **4**, which were characterized by elementary analysis and IR-spectra. **3** and **4** are cleaved by cyanide or carbon dioxide/dil. acetic acid to the C-21-oximes **11** and **12**, respectively.

Farbreaktionen von herzaktiven Cardenoliden mit Nitroprussidnatrium

Digitoxin (**1**) und Digitoxigenin (**2**) reagieren mit Nitroprussidnatrium [Dinatrium-pentakis(cyano-C)-nitrosyl-ferrat(II)] unter schwach basischen Bedingungen unter Bildung der entsprechenden Komplexe **3** und **4**, welche durch Elementaranalysen und IR-Spektren charakterisiert wurden. **3** und **4** wurden durch Cyanid bzw. Kohlendioxid/verd. Essigsäure zu den analogen C-21-Oximen **11** und **12** abgebaut.

1. Introduction

There are various colour reactions of cardiotonic glycosides with a cardenolid ring resulting from an interaction of the C-H-acidic $O=C-C=C-CH_2-O-$ increment with electron deficient aromatic compounds under basic conditions, e.g. Baljet reaction (picric acid) and Raymond reaction (m-dinitrobenzene) [1]. According to Kovar [2] Zimmermann-compounds are formed via intermediate Meisenheimer complexes.

Roth et al. [3] have found that C-H-acidic compounds are nitrosylated by disodium-pentakis(cyano-C)-nitrosyl-ferrate(II) (sodium nitroprusside, SNP) under the conditions of the Legal reaction [4]. We have characterized the blue complex formed from acetaldehyde, sec. amines and SNP [5] (Simon-Awe reaction), and Hardegger [6] reports upon a colour reaction of cardenolides with SNP. The analogous cardanolides and isocardenolides, however, do not form coloured complexes [7, 8]. These findings point towards a reaction at the C-H-acidic methylene increment of the unsaturated lactone ring at C-17.

2. Investigations, results and discussion

We have isolated the SNP-complexes of digitoxin (**1**) and of its aglycone digitoxigenin (**2**) and examined them by elementary analysis and degradation reactions.

Concerning the SNP-1 complex **3** the glycosidically attached digitoxoses were partially split off under the hydrolytic reaction conditions (cf. Experimental). Therefore, the elementary analysis data for C and H are lower, those for N are higher than the figures calculated for **3**. TLC using digitoxose for comparison shows a weak spot which gets coloured with anisaldehyde/sulphuric acid [9].

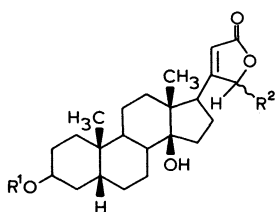
The IR-spectrum of complex **3** shows a weak C=O-absorption (lactone) at 1735–1745 cm⁻¹ besides the dominant C≡N-band (5 CN-groups) at 2100 cm⁻¹.

The well known rearrangement of cardenolides to isocardenolides is caused by bases [10, 11]. Therefore, we examined the stability of **1** under our conditions (cf. Experimental): **1** is rearranged in methanol/sodium methoxide slowly, the isocardenolide is detectable by TLC only after 40 min. So this rearrangement can be neglected because our SNP-1 complex **3** is precipitated after 10 min at the latest.

As stated in the Introduction cardanolides do not react with SNP [7, 8]. In order to correlate this observation with the C–H acidities of the unsaturated/hydrogenated lactones of cardenolides and cardanolides, respectively, we performed H-D exchange experiments [12]. Efforts to hydrogenate **2** to the corresponding cardanolide **12** according to [13–15] (atmospheric pressure, room temperature) failed, addition of acetic acid to the ethanol solution and 10 bar H₂, however, led to 20,22-dihydrodigitoxigenin (**15**).

According to Haberland et al. [16] all the three protons of the unsaturated lactone in cardenolides, e.g. **2**, are exchanged in dimethylformamide using triethylamine as a base, whilst the steroid skeleton remains unaffected: our conditions (CD₃OD/CD₃ONa) yielded the same result. An analogous experiment with the corresponding cardanolid **13** led to incorporation of maximal three D-atoms (FD-MS: d₀: 54%, d₁: 27%, d₂: 15%, d₃: 4%) but this reaction was accompanied by isomerization, as indicated by spectral data [12]. Lindig et al. [17] have reported that the cardanolid **13** is easily hydrolyzed at the lactone system and that ring closure occurs by acidification with glacial acetic acid. There are, however, no hints given concerning an isomerization.

Digitoxigenin (**2**) when being reacted with SNP in methanol/sodium methoxide formed a violet complex **4**. Elementary analysis established Na₃C₂₈H₃₄FeN₆O₅, the IR-spectrum revealed bands at 3400 (OH), 2920 (CH), 2080 (C≡N), 1740 (C=O, lactone) and 1620 cm⁻¹ (C=N). These data are in accordance with structure **4**. The stereochemistry at C-21 is unknown because **4** cannot be examined by ¹H NMR-spectroscopy: **4** is either poorly soluble in usual (deuterated) dipolar aprotic solvents or quickly decomposes.

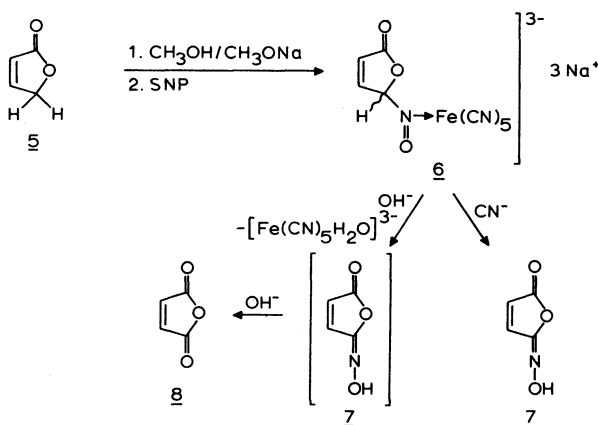


	R ¹	R ²
1	(digitoxose) ₃	H (digitoxin)
2	H	H (digitoxigenin)
3	(digitoxose) ₃	N→Fe(CN) ₅
4	H	N→Fe(CN) ₅

In order to find suitable cleavage conditions for the SNP-cardenolide complexes we studied the reaction of 2(5H)-furanone (**5**) with SNP: a violet precipitate **6** arose with correct analytical data for Na₃C₉H₃FeN₆O₃ and absorption bands in its IR-spectrum analogous to those of complex **4**.

Cleavage of the SNP-2(5H)-furanone complex **6** with potassium cyanide [5] led to 5-hydroxyimino-5(H)-furan-2-one (N-hydroxyisomaleimide **7**) showing data identical with those reported by Narita et al. [18] (Scheme 1). Unfortunately we were not able to reproduce Narita's procedure for the preparation of authentic **7**. So, our method seems to be an easy access to this compound. Smooth hydrolysis of complex **6** by 1.5 mol · l⁻¹ NaOH according to Roth et al. [3] delivered maleic anhydride (**8**) probably via **7** besides some maleic acid, but no fumaric acid. This hydrolysis converted complex **3** to anhydride **9**, the **2** complex **4** gave the corresponding anhydride **10**.

Scheme 1



Decomposition of **3** and **4**, respectively, with potassium cyanide [5] or cleavage with acetic acid/carbon dioxide according to Küster [19] led to the pertinent nitroso compounds which tautomerize to the oximes **11** and **12** as expected [20]. Oxime **11** is identical with the nitrosation product of **1**, synthesized following the general prescription of Tousters [21] (Scheme 2).

3. Experimental

3.1. Devices

M.p. (uncorr.): apparatus according to Dr. Tottoli (Büchi). IR-spectra in KBr: Beckman Acculab III. ¹H NMR and ¹³C NMR spectra: Varian EM 390 (90 MHz), Bruker WM 250 (250 MHz), CDCl₃, 35 °C, TMS as internal standard. MS: FAB-MS (glycerol, Xe): Varian MAT 311 A. All reactions were performed under N₂ and light protection between 0 and 5 °C. TLC on SiO₂ (Merck 5554), PTLC on SiO₂, 2 mm (Merck 5717).

3.2. Digitoxin-SNP complex **3**

The solution of 765 mg (1 mmol) digitoxin (**1**) and 296 mg (1 mmol) sodium nitroprusside (SNP) in 70 ml of abs. CH₃OH was alkalinized by 7 drops of 4 mol · l⁻¹ NaOCH₃, leading to a deeply red coloured solution. After being acidified with CH₃COOH this solution was evaporated in vacuo to 40 ml and a dark red solid precipitated which was washed several times with ice cold C₂H₅OH until no more **1** was found by TLC. Red precipitate of **3**. IR (cm⁻¹): 3400–3250 (OH); 2100 (C≡N); 1735–45 (C=O, lactone); 1610 (C=N).

Na₃C₄₆H₆₄FeN₆O₁₄ (1049.9)

Calcd.: C 52.6 H 6.10 N 8.0

Found: C 50.1 H 5.6 N 9.8

3.3. Digitoxigenin-SNP complex **4**

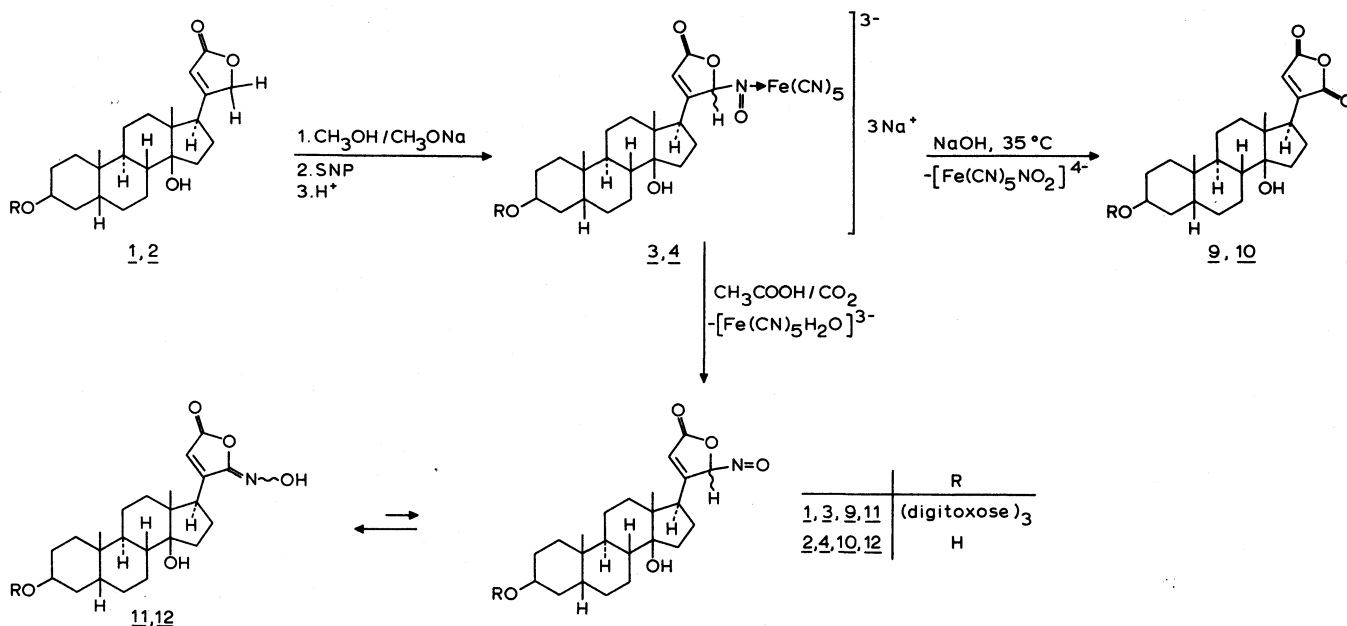
4 is formed from 375 mg (1 mmol) digitoxigenin (**2**) and 296 mg SNP as described for **3** as a red solid. IR (cm⁻¹): 3400 (OH); 2920 (OH); 2080 (C≡N); 1740 (C=O, lactone); 1620 (C=N).

Na₃C₂₈H₃₄FeN₆O₅ (659.4)

Calcd.: C 51.0 H 5.16 N 12.7

Found: C 50.6 H 4.94 N 12.6

Scheme 2



3.4. 2(5H)-Furanone-SNP complex 6

15 Dropping 0.2 mol of 4 mol · l⁻¹ NaOCH₃ in CH₃OH to 0.42 g (5 mmol) 2(5H)-furanone (5) (Aldrich chemicals) and 1.49 g (5 mmol) SNP in 15 ml of CH₃OH afforded a violet oil which solidified on repeated washing with ice cold C₂H₅OH. Violet solid after drying in vacuo. IR (cm⁻¹): 3400 (OH); 2100 (C≡N); 1745 (C=O, lactone); 1600 (C=N).

Na₃C₉H₃FeN₆O₃ (367.9)

Calcd.: C 29.3 H 0.82 N 22.8

Found: C 29.1 H 0.77 N 23.0

3.5. 5-Hydroxyimino-5(H)-furan-2-one (N-hydroxyisomaleimide, 7)

70 mg of complex 6 are stirred in 10 ml of 0.5 mol · l⁻¹ KCN for 3 h. Extraction with CH₂Cl₂, drying (Na₂SO₄) and evaporation in vacuo produced 15 mg (69%) of 7. M.p. 145 °C (dec.; lit. [14]: 145 °C (dec.)). IR (cm⁻¹): 1770 (CO-O-CN); 1630 (C=N-O).

C₄H₃NO₃ (113.07)

Calcd.: C 42.5 H 2.67 N 12.4

Found: C 42.4 H 2.59 N 12.5

3.6. Maleic anhydride (8) from complex 6

100 mg of complex 6, dissolved in 10 ml aqueous 1.5 mol · l⁻¹ NaOH, were kept at 35 °C for 30 min. TLC [diisopropylether/HCOOH/H₂O (90:7:3)] with maleic anhydride, maleic and fumaric acid for comparison shows much maleic anhydride (8, R_F 0.5) besides traces of maleic acid (R_F 0.2). Preparative TLC produces 11 mg (41%) maleic anhydride (8). M.p. 52–54 °C (authentic sample 54–56 °C). Its IR-spectrum is congruent with that of authentic material.

3.7. 21-Oxo-digitoxin (9)

0.2 g of complex 3 were dissolved in 10 ml of aqueous 1.5 mol · l⁻¹ NaOH and kept at 35 °C for 30 min. After extraction with (C₂H₅)₂O and usual work-up: 38 mg (26%) 9. Yellowish crystals. M.p. 189 °C. IR (cm⁻¹): 3250 (OH, br); 1790 (CO-O-CO). NI-FAB-MS: m/z 778 (M-H)⁻; 650; 520; 390; the pertinent differences correspond to the loss of one molecule of digitoxose (130 mu) each.

C₄₁H₆₂O₁₄ (778.9)

Calcd.: C 63.2 H 7.97

Found: C 62.6 H 7.08

Various attempts to get more precise data failed.

3.8. 21-Oxo-digitoxigenin (10)

0.2 g of complex 4 were hydrolyzed and worked up as described for 9: 37 mg (31%) of 10. M.p. 137 °C. IR (cm⁻¹): 3300 (OH); 1795 (CO-O-CO). NI-FAB-MS: m/z 387 (M-H)⁻.

C₂₃H₃₂O₅ (388.5)

Calcd.: C 71.1 H 8.25

Found: C 70.9 H 8.17

3.9. 21-Hydroxyiminodigitoxin (11)

3.9.1. 0.5 g of complex 3 were dissolved in 10 ml of H₂O and acidified to pH 5 with 2 drops of CH₃COOH. Complex 3 was decomposed by addition of dry ice (solid CO₂). Extraction with (C₂H₅)₂O, drying (Na₂SO₄) and evaporation in vacuo produced the oxime 11 as an amorphous solid. M.p. 183 °C (dec.). IR

(cm⁻¹): 3300 (OH); 1760 (CO-O); 1620 (C=N-O). ¹H NMR (250 MHz): all the data as known from 1 [22], except δ = 4.92–4.87 ppm (m; 0.4 H, H-21). ¹³C NMR (250 Hz; δ, ppm): all the data as known from 1 [22] except δ = 157.9 (C-21). PI-FAB-MS: m/z 794 (MH⁺), 664 (MH⁺-digitoxose), 534 (MH⁺-2 digitoxoses) 404 (MH⁺-3 digitoxoses), 386 (MH⁺-3 digitoxoses-H₂O). NI-FAB-MS: m/z 792 (M-H)⁻, 662 (M-H-digitoxose)⁻, 531 (M-H-2 digitoxoses)⁻, 513 (M-H-2 digitoxoses-H₂O)⁻, 401 (M-H-3 digitoxoses)⁻, 385 (M-H-3 digitoxoses-H₂O). [α]_D²⁰ (CHCl₃) = +35°.

3.9.2. 0.1 g of complex 3 were stirred in 10 ml of 0.5 mol · l⁻¹ KCN for 3 h. After bleaching the solution became dark blue ("prussian blue"). M.p., elementary analysis and IR-spectrum are identical with those of material obtained in 3.9.1.

3.9.3. To 0.59 g (0.8 mmol) 1 in 50 ml of abs. CH₃OH 0.55 g NaNO₂ (8 mmol) in 25 ml CH₃OH were added under stirring at 20 °C. Dropwise addition of fuming HCl afforded a white precipitate (NaCl) which was filtered off by suction after cooling. After evaporated to 1–2 ml, about 15 ml of H₂O were added and 11 was extracted with (C₂H₅)₂O. Evaporation and recrystallization from CHCl₃ afforded crystals: 104 mg (17%). M.p. 183 °C (dec.).

3.10. 21-Hydroxyiminodigitoxigenin (12)

0.1 g complex 4 were treated as described for compound 11 (procedure 3.9.1.) leading to 35 mg (57%) 12. M.p. 199 °C. IR (cm⁻¹): 1745 (CO-O), 1650 (C=N-O). ¹H NMR (250 MHz; δ, ppm): 5.87 (s; 1 H, H-22), 4.92–4.87 (m; 0.4 H, H-21), the other data as known from lit. [22]. ¹³C NMR (250 Hz; δ, ppm): all the data known for 2 [22] except δ = 157.9 ppm (C-21). PI-FAB-MS: m/z 404 (MH⁺), 389, 307, 263.

C₂₃H₃₃NO₅ (403.5)

3.11. 20,22-Dihydrodigitoxigenin (13)

100 mg of 2 in 25 ml of abs. C₂H₅OH and 3 drops of glacial CH₃COOH were hydrogenated with 40 mg of Pt-black at 10 bar H₂ and room temperature for 26 h. After filtration and evaporation in vacuo the oily residue was crystallized and recrystallized from CHCl₃: white needles, m.p. 224 °C (lit. [13]: 226 °C, CH₃OH). ¹H NMR; δ, ppm): 4.54–4.35 (m, 2 H, H-21), 2.5–1.8 (m, 3 H, H-20 and H-22), all the other data are found as described for 12 except the signals at δ = 5.87 and 4.92–4.87 ppm.

3.12. H/D exchange experiments with digitoxigenin (2) and 20,22-dihydrodigitoxigenin (13)

To 100 mg of 2 or 13, respectively, in 1 ml of CD₃OD were added 2 drops of NaOCD₃ and CD₃OD (40% solution, from Na⁰ and CD₃OD). After 5 min the mixture was slightly acidified by DCl (25% solution), Merck reagents) and evaporated at 40 °C in vacuo. OD groups were reexchanged to OH groups by 3 ml of H₂O, then the organic material was extracted with abs. (C₂H₅)₂O. Crystallization from CDCl₃ led to d₃-2. M.p. 254 °C (m.p. of 2: 255–256 °C [23]). ¹H NMR (δ, ppm): the singlet for H-22 at 5.87 and the doublet of C-21-CH₂, J = 18 Hz, in 2 are missing.

References

- 1 Auterhoff, H.; Knabe, J.: Lehrbuch der Pharmazeutischen Chemie, 11. ed., p. 427, Wissenschaftl. Verlagsges. mbH, Stuttgart 1983
- 2 Kovar, K.-A.: Pharm. unserer Zeit 1, 17 (1972)
- 3 Roth, H. J.; Surborg, K. H.: Arch. Pharm. (Weinheim, Ger.) 301, 686 (1968)
- 4 Legal, E.: Jahresber. Fortschr. Chem. Verw. Theile Anderer Wiss. 1883, 1648, cited according to [3]

- 5 Wiegrebe, W.; Vilbig, M.: *Z. Naturforsch.* **37b**, 490 (1982); **36b**, 1297 (1981)
- 6 Hardegger, E.; Heusser, H.; Blank, F.: *Helv. Chim. Acta* **29**, 477 (1946)
- 7 Elderfield, R. C.: *Chem. Rev.* **17**, 187 (1935)
- 8 Shoppe, C. W.: *Ann. Rev. Biochem.* **11**, 103 (1942)
- 9 Jork, H.; Funk, W.; Fischer, W.; Wimmer, H.: *Dünnschicht-Chromatographie*, p. 195, VCH Verlagsges. mbH, Weinheim 1989
- 10 Lindig, C.; Repke, K. R. H.: *Tetrahedron* **28**, 1847 (1972)
- 11 Krasso, A. F.; Binder, M.; Tamm, Ch.: *Helv. Chim. Acta* **55**, 1352 (1972)
- 12 Vogt, A. F.: forthcoming PhD Thesis, Regensburg
- 13 Cardwell, H. M. E.; Smith, S.: *J. Chem. Soc.* **1954**, 2012
- 14 Jacobs, W. A.; Scott, A. B.: *J. Biol. Chem.* **54**, 253 (1922); **78**, 573 (1928); **87**, 601 (1930); **93**, 131 (1931)
- 15 Smith, S.: *J. Chem. Soc.* **1930**, 2478 and **1935**, 1050
- 16 Haberland, G.; Maerten, G.: *Naturwissenschaften* **56**, 516 (1969)
- 17 Lindig, C.; Repke, K. R. H.: *Acta Biol. Med. Germ.* **26**, 501 (1971)
- 18 Narita, M.; Akiyama, M.; Okawara, M.: *Bull. Chem. Soc. Jpn.* **44**, 437 (1971)
- 19 Küster, W.: *Z. Physiol. Chem.* **51**, 157 (1926)
- 20 March, J.: *Advanced Organic Chemistry*, 3rd., p. 69, John Wiley & Sons, New York 1985
- 21 Tousters, O.; in: *Organic Reactions*, Vol. 7, p. 327, Wiley & Sons, New York 1957
- 22 Yamauchi, T.; Abe, F.; Wan, A. S. C.: *Chem. Pharm. Bull.* **35**, 2744 (1987)
- 23 Rheiner, A.; Hunger, A.; Reichstein, T.: *Helv. Chim. Acta* **35**, 687 (1952)

Received December 7, 1989

Prof. Dr. W. Wiegrebe
Universitätsstr. 31
W-8400 Regensburg