

Pathophysiology of GvHD and HSCT related complications

Ernst Holler^{1*}, Sakhila Ghimire¹, Anne M. Dickinson², Daniela Weber¹, Emily Marvine²

¹Department of Haematology/Oncology, University Medical Centre, Germany,

²Hematological Sciences, Institute of Cellular Medicine, Newcastle University, United Kingdom

Submitted to Journal:
Frontiers in Immunology

Specialty Section:
Alloimmunity and Transplantation

ISSN:
1664-3224

Article type:
Review Article

Received on:
06 Aug 2016

Accepted on:
17 Jan 2017

Provisional PDF published on:
17 Jan 2017

Frontiers website link:
www.frontiersin.org

Citation:

Holler E, Ghimire S, Dickinson AM, Weber D and Marvine E(2017) Pathophysiology of GvHD and HSCT related complications. *Front. Immunol.* 8:79. doi:10.3389/fimmu.2017.00079

Copyright statement:

© 2017 Holler, Ghimire, Dickinson, Weber and Marvine. This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution and reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

This Provisional PDF corresponds to the article as it appeared upon acceptance, after peer-review. Fully formatted PDF and full text (HTML) versions will be made available soon.

1 ***Pathophysiology of GvHD and other HSCT related major***
2 ***complications***

3
4 Sakhila Ghimire¹, Daniela Weber¹, Emily Mavin² Anne Dickinson² Ernst Holler¹

5
6 Affiliations

7 ¹Department of Internal Medicine III, University Medical Centre, Regensburg, Germany

8 ²Hematological Sciences, Institute of Cellular Medicine, Newcastle University, United Kingdom

9
10 **Correspondence:**

11 Ernst Holler, Department of Hematology and Oncology, Internal Medicine III, University Medical
12 Center, Franz-Josef-Strauß-Allee 11, 93053 Regensburg, Germany; phone: 0049-941-944-5542,
13 fax: 0049-941-944-5543

14 e-mail: ernst.holler@ukr.de

15 **Running title:**

16 Pathophysiology of GvHD and other HSCT related major complications

17 **Keywords:**

18 Hematopoietic stem cell transplantation, Graft vs. Host Disease, prophylaxis, T cells

19
20 Total Number of Words: 4799

21 Total Number of Figures/Tables: 3

29

30

31 **Abstract**

32 For over 60 years, hematopoietic stem cell transplantation (HSCT) has been the major curative
33 therapy for several hematological and genetic disorders, but its efficacy is limited by the secondary
34 disease called graft versus host disease (GvHD). Huge advances have been made in successful
35 transplantation in order to improve patient quality of life and yet, complete success is hard to
36 achieve. This review assimilates recent updates on pathophysiology of GvHD, prophylaxis and
37 treatment of GvHD related complications and advances in the potential treatment of GvHD.

38 **1. Introduction to Graft versus Host Disease (GvHD)**

39 GvHD is the most recognized complication post HSCT and was first observed in 1956 in a murine
40 model. Barnes et al demonstrated that when irradiated mice were infused with allogenic marrow
41 and spleen cells, mice recovered from radiation injury and aplasia but they developed diarrhea,
42 weight loss, skin changes and liver abnormalities, and subsequently died due to “secondary
43 disease”[1]. This phenomenon was recognized as GvHD. A decade later, in 1966, Billingham
44 postulated three crucial requirements for the development of GvHD:

- 45 i) the transplanted graft must contain immunologically competent cells,
- 46 ii) the recipient must be incapable of rejecting or eliminating transplanted cells,
- 47 iii) the recipient must express tissue antigens that are not present in the transplant donor, thus
48 the recipient antigens are recognized as foreign by donor cells [2].

49 Today, we know that the immunocompetent cells are T lymphocytes that are present in the stem
50 cell inoculum and are required to mount an effective immune response [3]. A normal immune
51 system is able to reject T cells from a foreign donor. However, when recipient’s immune system
52 is compromised through the use of various immune-ablative agents (chemotherapy and/or
53 radiotherapy), the recipient is incapable of rejecting the transplanted cells. We now know that the
54 tissue antigens that differ in donor and recipient are major and minor human leukocyte antigens
55 (HLA), and their expression on cell surfaces are crucial for the activation of allogenic T cells and
56 initiation of GvHD (4). Previously it was believed that acute GvHD occurs within day 100 after
57 transplantation and chronic GvHD occurs beyond day 100 and that the most affected organs at the
58 onset of GvHD is skin (81%), gastrointestinal tract (54%) and liver (50%) [4]. Now it is clear that
59 acute GvHD can occur after day 100 as late acute GvHD (e.g. after cessation of
60 immunosuppression or after donor lymphocyte infusion (DLI)) or cause overlap syndrome of both
61 acute and chronic GvHD [5].

62 **2. Pathophysiology of acute GvHD: a three-step model explaining the current strategies of 63 prophylaxis and treatment**

64 Acute GvHD has been attributed to three stages. Initially there is tissue damage due to conditioning
65 which in turn activates the host antigen presenting cells (APCs). Secondly, APCs activate donor T

66 cells, also known as an afferent phase. Finally, in efferent phase, cellular and inflammatory factors
67 work together to damage the target organs.

68 *a. conditioning mediated tissue damage:* conditioning is crucial to eradicate underlying disease
69 and to support engraftment of donor cells without rejection by recipient[6]. Prior to donor cell
70 infusion, patient's tissues have been profoundly damaged due to underlying disease itself,
71 treatment for the disease, infections, and the conditioning regimen [7, 8]. As a consequence,
72 damaged host tissue release danger signals which include pro-inflammatory cytokines such as
73 tumor necrosis factor (TNF) and interleukin-1 (IL-1) [9], that activate host APCs, ultimately
74 activating donor T cells present in the stem cell inoculum [10, 11]. Conditioning mediated
75 damage to the GI tract remains the main concern as GI tract allows systemic translocation of
76 microbial products like lipopolysaccharide (LPS) and other pathogen associated molecular
77 patterns (PAMPs) that greatly amplify host APC activation [8], leading to amplified T cell
78 activation. Conditioning related damage also explains why the concept of reduced intensity or
79 even non-myeloablative conditioning has contributed to less toxicity, less severe GvHD and
80 reduced treatment related mortality. Some studies showed that delaying the transfer of donor
81 cells after conditioning decreased the risk of GvHD [9, 12].

82 *b. donor T cell activation (the afferent phase):* GvHD occurs when donor T cells activate and
83 respond to HLA differences on recipient's tissue [13]. Experimental models have proved that
84 the host APCs are necessary and sufficient to activate donor T cells and initiate GvHD [11,
85 14]. Donor T cells can recognize alloantigen either on host APC, known as direct antigen
86 presentation [15] or on donor APCs, known as indirect presentation [16]. T cell responses
87 depend on the disparity between the donor and the recipient with regard to HLA [13]. CD4+
88 T cells respond to the variations in MHC class II molecule (HLA-DR, DQ, DP) and CD8+ T
89 cells respond to the variations in MHC class I molecule (HLA-A, B, C) [17]. Transplants
90 carried out in the HLA matched sibling or identical twin setting can still give rise to GvHD
91 due to differences in minor HLA [18]. The first to be described were HA-1 [19] and HA2 [20]
92 and the subsequent clinical impact of minor histocompatibility antigens including H-Y
93 antigens [21, 22] of female to male transplants has recently been reviewed [23, 24]. Minor
94 HLAs are T cell epitopes which are originally derived from polymorphic or normal tissue
95 proteins. These antigenic peptides can be presented on HLA Class I or Class II molecules and
96 to date over 50 minor HLA antigens have been identified [24]. Minor HLA antigens have been
97 associated with GvHD and graft versus leukemia (GvL) effects due to their tissue distribution.
98 Minor HLA antigens restricted to the hematopoietic system may be able to enhance GvL
99 responses while more broadly expressed minor HLA antigens contribute to both GvHD and
100 GvL [25]. As well as cytotoxic T cell responses allogeneic H-Y antibodies have shown to
101 predict chronic GvHD and non-relapse mortality [26, 27].

102 T cell activation is in the focus of current immunosuppressive strategies used for prophylaxis
103 and treatment. Calcineurin inhibitors, mycophenolate and mTOR inhibitors interfere with
104 different signals of T cell activation [28, 29]. The broader strategy is T cell depletion, which is
105 currently applied by in vivo approaches such as the use of anti-thymocyte globuline
106 pretransplant [30]. Cytotoxic approaches more or less selectively eliminates activated T cells
107 if applied posttransplant; the old approach of methotrexate prophylaxis but also the more recent
108 approach of using posttransplant cyclophosphamide engage this principle [31].

109

110 *c. target cell apoptosis (the efferent phase):* in this phase, both innate and adaptive immune
111 cells work synergistically to exacerbate the T cell induced inflammation. Cellular mediators
112 such as cytotoxic T lymphocytes (CTLs) and natural killer (NK) cells utilize the Fas/Fas ligand
113 (FasL) pathway and perforin/granzyme pathway to lyse the target cells [32, 33]. Furthermore,
114 inflammatory cytokines synergize with CTLs, resulting in further tissue injury and possible
115 target organ dysfunction [13]. In addition, microbial products like LPS, released during
116 conditioning, leak through a damaged intestinal mucosa and skin, and stimulate mononuclear
117 cells (monocytes/macrophages) to secrete inflammatory cytokines leading to amplification and
118 propagation of a cytokine storm [13]. This leads to destruction of epithelial cells, mostly in the
119 GI tract.

120 The broad activity of corticosteroids including induction of T cell apoptosis, suppression of
121 macrophage activation and cytokine release explains why these old drugs are still the treatment
122 of choice for first line treatment of both acute and chronic GvHD. Cytokine inhibitors like TNF
123 blocking agents were thought to be more specific but did not result in increased response rates
124 [34]. For almost all second line strategies in steroid-refractory acute GvHD low response rates
125 associated with high treatment related mortality have been reported indicated the urgent need
126 for further improvement [35].

127 In the last 10 years, the concept of GvHD pathophysiology has been largely extended and a more
128 differentiated view has been adapted.

129 Firstly, the mechanism of conditioning related damage has further been specified. It is now clear
130 that the tissue damage results in release of several danger signals such as uric acid and the
131 metabolites of ATP pathway and its receptor have been shown to be involved in activation of
132 GvHD [36].

133 Secondly, the concept of LPS triggered inflammation has been substituted by multiple microbiota
134 derived signals and differential activation of Toll-like receptors (TLRs) and Nod-like receptors
135 (NLRs). NOD2/CARD15 has been shown to be involved in triggering the inflammation both in
136 mice [37, 38] and men [39]. More recently it became clear that the microbiota of epithelial tissues
137 is a the major player influencing epithelial integrity and local immune tolerance by commensal
138 bacteria and millions of metabolites are produced to maintain epithelial homeostasis [40, 41].

139 Finally, and in context with the concept of microbiota as important players, the importance of
140 regulatory immune cells which balance immune reactions is recognized. Regulatory T cells (Tregs)
141 expressing the transcription factor Foxp3 occur as natural, thymus derived T cells and are able to
142 prevent alloreaction [42]. On epithelial surfaces, induced peripheral Tregs try to dampen acute
143 inflammation [43]. Foxp3 positive T cells act in cooperation with numerous newly identified
144 regulatory populations such as invariant natural killer T (iNKT) cells [44], myeloid derived
145 suppressor cells (MDSCs) and a whole new set of innate immune cells such as innate lymphoid
146 cells (ILCs) [45]. **Figure 1** represents the GvHD initiation phase. **Figure 2** summarizes the
147 complete pathophysiology of aGvHD.

148 **3. In vitro Modelling of GvHD to give insight into the pathophysiology**

149 The skin explant model has long been established as a tool for studying the immunobiology of
150 GvHD and more recently has been used to investigate the specificity of anti-viral T cells in graft
151 versus host reactions (34, 35, 36) the role of Tregs and mechanisms of apoptosis [46, 47].

152 The skin explant model has also been used to assess the safety of *ex vivo* expanded Treg cells as
153 well as their capacity to prevent graft versus host (GvH) reactions [48]. Activated and expanded
154 polyclonal Treg, at any cell concentration, did not induce any significant GvH reactions.

155 Over recent years significant advances in the understanding of the benefits of Tregs in
156 hematopoietic stem cells transplantation have resulted in the completion of early stage clinical
157 trials as well as the initiation of trials in solid organ transplantation [49-52]. These early stage
158 HSCT trials have provided promising results showing a reduction the incidence of GvHD without
159 adversely effecting relapse, transplant related mortality and engraftment. Using the skin explant
160 model it has been possible to investigate the cellular and molecular mechanisms by which Treg
161 are likely to be preventing GvHD following HSCT.

162 We have shown that for Treg to suppress GvH reactions they need to be present during the priming
163 of allo-reactive T cells [48]. Polyclonal Treg cells were expanded *ex vivo* and added into the skin
164 explant model at either the priming or the effector stage. The later addition of Treg, during the
165 effector phase, impaired their suppressive capacity. This suggests that Treg may be more effective
166 when given early, as prophylaxis, rather than as a treatment. This study also demonstrated that in
167 humans an effector to Treg ratio of 4:1 was sufficient to modulate GvH reactions, whereas previous
168 studies in mice had suggested a 1:1 ratio was necessary. This study has therefore provided
169 preclinical evidence to support the safety and feasibility of *ex vivo* expanded Treg as a novel
170 therapeutic and provides information on the optimal timing and dose of Treg to prevent GvH
171 reactions.

172 Further work using the skin explant model has been able to elucidate some of the mechanisms by
173 which Treg are able to prevent GvHR. The presence of Treg during the priming of allo-reactive T
174 cells reduced their cytotoxic capacity [48]. Further investigations showed Treg also impaired the
175 ability of allo-reactive T cells to migrate into the target tissues [53]. The presence of Treg during
176 priming resulted in a reduction in IFN γ production by CD8+ cytotoxic T cells, as well as a reducing
177 expression of skin homing molecules CXCR3 and CLA. This paired with a reduction in levels of
178 the chemokines CXCL10 and CXCL11 in the skin resulted in a significant reduction in the number
179 of cytotoxic T cells present in the skin and decreased the GvH severity. We have since
180 demonstrated that Treg are able to modulate GvH reactions through impairment of dendritic cells
181 at a transcriptional level, arresting them in a semi-mature status and leaving them functionally
182 impaired [54]

183 The skin explant model has also been used to investigate the involvement of epithelial FAS in the
184 pathophysiology of GvHD [55]. Animal models have previously shown the critical role for
185 FAS/FASL in GvHD [56]. Ruffin et al showed that there was a significant increase in FAS
186 expressing cells in GvHR positive experiments and that FAS mediated apoptosis was involved in
187 the induction of GvHR, as blocking FAS mediated apoptosis reduced the severity of GvHR. They
188 also showed that levels of FAS in the serum of patients who received myeloablative conditioning
189 was increased, possibly due to the higher toxicity. This supports the potential use of FAS as a
190 therapeutic target.

191 4. Identification of Biomarkers

192 As well as investigating the safety of cellular therapies and immunology of GvHD, the skin explant
193 model has been used in recent years to identify biomarkers. Within our group we used the skin
194 explant model to validate a number of biomarkers which had been identified in the serum of HSCT
195 patients [57]. BAFF and IL-33 levels were elevated pre-transplant in patients who then went on to
196 develop aGvHD, and therefore could have the potential to act as predictive biomarkers. We also
197 found that CXCL10 and CXCL11 were suitable as diagnostic markers of GvHD. Training and
198 validation cohorts were used to highlight the association of these potential biomarkers to GvHD.
199 Then the skin explant model was used to confirm their association with GvH reactions.
200 Immunohistochemistry was carried out on sections from the skin explant and increased staining
201 for BAFF, IL-33, CXCL10 and CXCL11 was seen in skin explants with a higher grade GVHR.
202 This was further confirmed in clinical biopsies demonstrating increased levels of protein, measured
203 with immunohistochemistry and gene expression for BAFF, CXCL10 and CXCL11. In this study
204 the skin explant proved to be a useful tool in validating a panel of biomarkers which had been
205 identified in patient samples. The skin explant is not exclusive to the human setting. Recently
206 Zinocker et al have described the use of a rat skin explant model for investigating the
207 pathophysiology of GvHD [58] as well as gene expression profiling [59].

208 Harris et al [60] have recently reviewed the use of biomarkers in predicting acute GvHD which
209 include genomic factors as well as plasma proteins. One of the first studies demonstrated that a
210 panel of tumor necrosis factor receptor –type 1 (TNFR1) interleukin -2 receptor alpha (IL-2 α),
211 IL-8 and hepatocyte growth factor (HGF) had prognostic as well as diagnostic value in predicting
212 acute GvHD [61]. Other markers in the skin such as elafin [61] and plasma biomarkers of the lower
213 GI tract and liver acute GvHD have been validated in subsequent studies and the most significant
214 of these was regenerating islet-derived 3 alpha (Reg3a) [62, 63]. These studies led to the use of
215 the biomarkers TNFR1, ILR α , IL-8, HGF, Reg3 α and elafin for measuring responsiveness to
216 GvHD therapy. The panel was able to predict 28 day post therapy non response and day 180-
217 mortality in a cohort of 112 patients [64].

218 In addition an algorithm using concentrations of three biomarkers-TNFR1, soluble IL-33 receptor
219 [ST2] and Reg3 α , Levine and colleagues [65] were able to calculate the probability of nonrelapse
220 mortality caused by non-responsive GVHD and divide the patients into distinct groups to predict
221 response to GVHD therapy. The researchers subsequently developed the Mount Sinai Acute
222 GVHD International Consortium (MAGIC), which consists of a group of 10 transplant centers in
223 the United States and Europe who collaborate on the use of this scoring system to test new
224 treatments for acute GVHD.

225 5. Pathophysiology of chronic GvHD

226 Although the pathophysiology of chronic GvHD (cGvHD) is poorly understood, it remains the
227 major cause of late non-relapse death after HSCT [66]. cGvHD may manifest simultaneously from
228 aGvHD, develop after the treatment of aGvHD or may occur *de novo* [67]. Classical cGvHD
229 occurs 100 days after transplantation but may also overlap with aGvHD [5, 68].

230 Acute GvHD is a major risk factor of cGvHD and strategies aiming at T cell depletion at the time
231 of transplantation to prevent cGvHD demonstrate that early events impact on the development of
232 cGvHD. As immune cells and immune organs such as thymus, bone marrow niche and spleen are

233 the primary targets of acute GvHD, thymus destruction and deficient selection of donor T cells by
234 the thymus are the major factors resulting in allo- and autoimmunity associated with chronic
235 GvHD [69]. Due to early damage of the B cell niche in the bone marrow, B cell development is
236 strongly disturbed resulting in elevated BAFF levels as a predictor of chronic GvHD and
237 insufficient elimination in B cells producing auto- and alloantibodies [70]. A hallmark of cGvHD
238 is development of sclerotic lesions which can occur in almost every organ [68]. While previous
239 data favors a concept of defective wound healing with increase production of sclerotic cytokines
240 such as TGF β and PDGF, recent evidence supports a role of specific TH17 subsets in this sclerotic
241 process [71].

242 **6. Target organ damage during GvHD**

243 Skin is the principal target organ of GvHD and the initial manifestation in the skin is
244 maculopapular rash which has the potential to spread throughout the body [13]. The rash may
245 resemble folliculitis or may resemble sunburn. In extreme cases, skin may blister and ulcerate [13,
246 72]. Acute cutaneous GvHD usually begins with erythematous, rashes on the ears, palms and soles.
247 Martin and coworkers reported results of 740 allogeneic transplantations and 81% of patients with
248 aGvHD had skin involvement [4]. Damage to the skin could be defined by vacuolar degeneration
249 of the basal cell layer, dyskeratotic keratinocytes and mononuclear cell infiltrates [73]. Epithelial
250 damage occurs at the tips of rete ridges and hair follicles, regions where selective targeted apoptotic
251 rete cells are located [74]. A recent study by Paczesny et al reported that elafin could be a potential
252 biomarker for diagnosis and prognosis of skin GvHD [75].

253 Liver is another target organ of GvHD. Hepatic GvHD is manifested by abnormal liver function
254 tests and a rise in the serum level of bilirubin and alkaline phosphatase. Donor lymphocytes attack
255 the bile duct epithelial cells causing endothelialitis, pericholangitis and apoptotic bile duct
256 destruction [76]. While liver GvHD affecting bile ducts and resulting in severe hyperbilirubinemia
257 occurs less frequently, there is an increasing rate of hepatitis like cGvHD as another, but less
258 harmful liver lesion [77].

259 Gastro-intestinal (GI) tract represents the most severely affected organ after conditioning. GI
260 GvHD is characterized by secretory and voluminous diarrhea, severe abdominal pain, vomiting
261 and anorexia [13]. Snover and colleagues used immunohistochemistry to explain histologic
262 features of the GI tract during GvHD [78]. Single cell apoptosis was observed along with patchy
263 ulcerations and apoptotic bodies in the base of crypts with loss of the surface epithelium [13, 78].
264 The base of the intestinal crypts, where epithelial stem cells are located, is the most sensitive target
265 for GvHD as it is the site of epithelium regeneration and Paneth cells. Recently, Levine and
266 colleagues observed loss of the Paneth cells at the onset of GI GvHD [79] suggesting these cells
267 as sensitive targets of GvHD. In addition, as stated earlier, it was proposed that regenerating islet-
268 derived 3-a (reg3a), released from Paneth cells, was a potential plasma biomarker for lower GI
269 GvHD [63], Paneth cell damage contributes to loss of antimicrobial peptides and accelerates the
270 loss of microbial diversity in GvHD, a major risk factor of treatment related mortality [80, 81].

271 **7. Further HSCT related complications**

272 **7.1 Overview**

273 Although GvHD is the main complication of allogeneic SCT, non-relapse related mortality (NRM)
274 can occur independently from the occurrence of GvHD or in patients with minor GvHD. Overall,
275 NRM has decreased in the last 10 years as a result of several improvements such as reduced
276 intensity conditioning; resulting in reduced organ toxicity, improved donor selection and matching,
277 and progress in supportive treatment [82].

278 Major complications include viral and fungal infections, which can occur independently from
279 GvHD due to the immunodeficiencies induced by HSCT. GvHD and its treatment aggravate and
280 prolong the risk of infectious complications, and many patients suffering from severe GvHD die
281 from infectious complications. Beyond the period of acute GvHD, chronic GvHD and long term
282 complications are major causes of NRM and morbidity. Long term complications include organ
283 toxicities, endocrine deficiencies and most important secondary cancers. HSCT patients survivors
284 therefore need a long term follow up in order to allow early detection of complications and several
285 guidelines summarize the current recommendations [83, 84].

286 A detailed presentation of infectious complications, organ toxicities and long term complications
287 is beyond the focus of this review, we therefore focus on the most relevant targets of complications:
288 endothelial cells and pulmonary complications.

289 **7.2 Endothelial complications**

290 Endothelial complications occur clinically throughout the different phases of HSCT. In the early
291 weeks after transplantation, sinusoidal obstruction syndrome (SOS) formally known as veno-
292 occlusive disease (VOD) can result in severe liver damage and eventually multi-organ failure [85,
293 86]. SOS results from conditioning related toxicity in the sinusoids of the liver with subsequent
294 occlusions by thrombosis and fibrosis. In the period of engraftment, cytokine storm mediated
295 capillary leakage syndrome can occur. With the introduction of calcineurin-inhibitors (CNI) for
296 prophylaxis of GvHD, which also give rise to some endothelial toxicity, transplant associated
297 microangiopathy (TAM) has been increasingly observed during acute GvHD [87]. Manifestations
298 of intestinal TAM can mimic severe GvHD and provoke intestinal bleeding but requires a different
299 treatment regimen. Besides CNI associated TAM, it can also occur as atypical hemolytic uremic
300 syndrome (HUS) which results from a failure of cleaving von Willebrand Factor [88, 89]. In long
301 term patients cerebro- and cardiovascular complications are increased.

302 While clinical endothelial complications have been well known for many years, more recently the
303 pathophysiology of endothelial cells in GvHD has been studied. *In vitro* models of endothelial cell
304 cultures reveal that conditioning can induce endothelial apoptosis, which is aggravated by LPS
305 mediated inflammation and followed allogeneic cytotoxic T cell damage [90]. Murine models have
306 demonstrated the role of endothelial neovascularization induced by conditioning leading to GvHD
307 [91-93] and infiltrating donor T cells. Recently, Schmid et al showed for the first time in a murine
308 system, that not only endothelial venules but also arterial vessels suffer direct endothelial damage
309 during GvHD [94]. Detailed studies in patients have shown an association of loss of dermal
310 vessels, with CD8+ T cell infiltrates, demonstrating allogeneic reactions against endothelial cells
311 [95, 96]. More recently, endothelial damage has been shown to contribute to steroid resistance and
312 failure to recover from GvHD. Loss of protective thrombomodulin was observed in biopsies from
313 GvHD patients [97] together with increased serum thrombomodulin [98]. In addition, genetic
314 SNPs within the thrombomodulin gene have been identified as risk factors for GvHD [99].
315 Finally, circulating endothelial factors such as angiotensin levels pre transplant and VEGF levels

316 post-transplant have been identified as risk factors of GvHD [100] which paves the way for
317 infiltrating donor T cells.

318 **7.3 Pulmonary complications**

319 A further central target organ of HSCT related complications is the lung. Early after
320 transplantation, bacterial and fungal pneumonia are common, mainly due to aspergillus
321 predominance. In the post-transplant period of GvHD and immune reconstitution, viral pneumonia
322 caused by CMV, respiratory viruses (Influenza, Parainfluenza, RSV, Metapneumovirus) and
323 adenoviruses predominate as well as fungal pneumonia, especially in patients with severe
324 immunosuppression [101]. In addition, further infectious agents such as toxoplasma gondii and
325 pneumocystis jirovecii causing toxoplasma and pneumocystis pneumonia (PcP) respectively, can
326 cause pneumonia during the period of B cell reconstitution while B cell numbers are absent or low.
327 Pneumonias caused by encapsulated bacteria such as pneumococci are also observed [102, 103].
328 Early after HSCT, peri-engraftment respiratory distress syndrome (PERDS) causes rapid
329 deterioration of respiratory functions during leukocyte recovery, but responds rapidly to high dose
330 corticosteroid treatment. In the initial stages of aGvHD, idiopathic pneumonia syndrome (IPS) is
331 a serious complication resulting from conditioning related toxicity and LPS triggered allogeneic
332 reactions. IPS may or may not be exacerbated by occult or unknown infections [89] and in either
333 case, TNF blocking agents have been shown to be effective in both experimental models and in
334 patients [86-88]. The most frequent complication is bronchiolitis obliterans syndrome (BOS)
335 characterized by inflammation of the small bronchiole with subsequent obstruction and lung
336 destruction [104, 105]. Early monitoring and intervention with topical corticosteroids,
337 azithromycin and possibly systemic immunosuppression is needed to prevent progression to
338 irreversible lung damage which may lead a requirement for lung transplantation [106, 107].
339 Besides BOS, restrictive changes can be observed such as pulmonary fibrosis, BOOP
340 (bronchiolitis obliterans organizing pneumonia) and pulmonary veno-occlusive disease [108].

341 Cellular therapy is one approach increasingly used as a second line treatment. Tregs [109] and
342 MSCs [110-112] are promising cellular products, but phase 3 trials are yet to be conducted.

343 **8. Recent advances and perspectives in GvHD**

344 For over 30 years, immunosuppressive drugs have served as a central strategy to reduce GvHD.
345 Drugs such as sirolimus, tacrolimus and methotrexate are the main-stay in the treatment of GvHD
346 [113]. Complete ex vivo T cell depletion is no longer routinely used in HLA matched
347 transplantation as it also largely abolishes GvL effects. A more recent report from Finke and
348 colleagues suggested ATG as an *in vivo* T cell depletion may be more efficacious in lowering the
349 incidence of severe acute GvHD in matched and mismatched HSCT from unrelated donors while
350 GvL effects seemed less affected [114]. 100 patients were enrolled in the study. Comparable
351 outcomes were obtained for GvHD patients receiving bone marrow or peripheral blood stem cells
352 from matched or one antigen mismatched unrelated donors when ATG was added to the standard
353 prophylaxis (cyclosporine+methotrexate) [114]. The use of ATG may therefore contribute to
354 balance GvH versus GvL effect and enable HLA mismatch donors to be used as well as fully match
355 unrelated donors, with no difference in outcome. As an alternative, elimination of alloreactive T
356 cells by post-transplant cyclophosphamide may become an option, which is already widely used
357 for GvHD prophylaxis following haploidentical transplantation [115]. Whether this approach can

358 be integrated in the HLA identical setting as a potential alternative to calcineurin inhibitors is under
359 current investigation.

360

361 Pathogen recognition receptors (PRRs) like NLRs and TLRs are known to control adaptive
362 immune responses in inflammatory disorders [37] and the research on the role of these receptors
363 has resulted in the description of the interaction of the microbiota and the immune system in the
364 setting of GvHD. Loss of microbiome diversity early after HSCT has been recognized as a new
365 risk factor for GvHD and HSCT related complications [116]. This observation suggests that
366 restoration of a diverse microbiome could be a new approach to induce intestinal and systemic
367 tolerance, and pre-/pro- and post biotic strategies, as well as several approaches of fecal microbiota
368 transplantation (FMT) which are currently being tested in both experimental and clinical settings
369 of HSCT [41].

370 Tregs have been expanded *in vitro* and used for prophylaxis and treatment of GvHD in
371 experimental and small clinical trials [117, 118]. Another option is induction of Tregs in patients
372 e.g. by Interleukin-2 (IL-2) [119]. Induction of Tregs has also been postulated as one mechanism
373 explaining the beneficial action of extracorporeal photopheresis for treatment of acute and chronic
374 GvHD [120, 121]. Besides Tregs, numerous alternative candidates for cellular therapy of GvHD
375 exist such as MDSCs [122]. MSCs are indirect immunoregulatory cells which induce tissue repair
376 and show some promising activity in steroid refractory GvHD [123, 124].

377 Among pharmacological agents, drugs with anti-inflammatory effects of corticosteroids but
378 without numerous side effects are urgently needed. Recently, anti-inflammatory JAK2 inhibitors
379 have shown promising effects both in GvHD and in rheumatology. Proteasome inhibitors and
380 histone deacetylase (HDAC) inhibitors originally developed as anticancer drugs now show some
381 promising activity in dampening T cell responses [125]. In cGvHD, the role of aberrant B cells is
382 increasingly recognized which paves the way for anti-B cell strategies like rituximab or new B cell
383 development inhibitors like the Bruton's tyrosine kinase (BTK)-inhibitor ibrutinib [70]

384 A major issue in the treatment of aGvHD is that most approaches are initiated too late, when major
385 changes have already severely damaged the target tissue. Therefore, biomarkers allowing early
386 identification of patients at high risk are needed. A handful of biomarkers have been discovered
387 which might be used to guide treatment in the future [65].

388 Finally, the practice of stem cell transplantation differs between countries, within the same
389 countries and between transplantation institutes. Approaches aimed at standardization of diagnosis
390 and treatment are urgently needed, some of which have been addressed by several consensus
391 projects [68, 126].

392

393

394

395

396
397
398
399
400
401
402
403
404
405
406
407
408
409
410
411
412
413
414
415
416
417
418
419
420
421
422
423
424
425

Conflict of Interest

All authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Author Contribution

EH and AD designed and revised the review. SG provided the draft, summarized available data, selected the references and wrote the review. DW and EM contributed to the review. All authors approved the final version of the manuscript.

Funding

The project was supported by Marie Curie Initial Training Networks grant; Project Number 315963 “Improving HSCT By Validation of Biomarkers & Development Of Novel Cellular Therapies”.

Acknowledgements

We acknowledge Dr. Saroj Ghimire for critical revision of the manuscript and Mrs. Katie Gray for her outstanding management throughout the Celleurope Project.

Provisional

427 **References**

- 428 1. Barnes, D. and J. Loutit, *Treatment of murine leukaemia with X-rays and homologous*
429 *bone marrow: II*. British journal of haematology, 1957. **3**(3): p. 241-252.
- 430 2. Billingham, R.E., *The biology of graft-versus-host reactions*. Harvey Lect, 1966. **62**: p.
431 21-78.
- 432 3. Kernan, N.A., et al., *Clonable T lymphocytes in T cell-depleted bone marrow transplants*
433 *correlate with development of graft-v-host disease*. Blood, 1986. **68**(3): p. 770-773.
- 434 4. Martin, P.J., et al., *A retrospective analysis of therapy for acute graft-versus-host disease:*
435 *initial treatment*. Blood, 1990. **76**(8): p. 1464-1472.
- 436 5. Filipovich, A.H., et al., *National Institutes of Health consensus development project on*
437 *criteria for clinical trials in chronic graft-versus-host disease: I. Diagnosis and staging*
438 *working group report*. Biology of Blood and Marrow Transplantation, 2005. **11**(12): p.
439 945-956.
- 440 6. Gratwohl, A. and E. Carreras. *Principles of conditioning*. in *Haematopoietic Stem Cell*
441 *Transplantation. The EBMT Handbook, 5th edn. Forum Service Editore: Genoa, Italy.*
442 2008.
- 443 7. Kaitin, K.I., *Graft-versus-host disease*. N. Engl. J. Med, 1991. **325**(5): p. 357-358.
- 444 8. Hill, G.R., et al., *Total body irradiation and acute graft-versus-host disease: the role of*
445 *gastrointestinal damage and inflammatory cytokines*. Blood, 1997. **90**(8): p. 3204-3213.
- 446 9. Xun, C., et al., *Effect of total body irradiation, busulfan-cyclophosphamide, or*
447 *cyclophosphamide conditioning on inflammatory cytokine release and development of*
448 *acute and chronic graft-versus-host disease in H-2-incompatible transplanted SCID*
449 *mice*. Blood, 1994. **83**(8): p. 2360-2367.
- 450 10. Matzinger, P., *The danger model: a renewed sense of self*. Science, 2002. **296**(5566): p.
451 301-305.
- 452 11. Shlomchik, W.D., et al., *Prevention of graft versus host disease by inactivation of host*
453 *antigen-presenting cells*. Science, 1999. **285**(5426): p. 412-415.
- 454 12. Johnson, B.D. and R. Truitt, *Delayed infusion of immunocompetent donor cells after*
455 *bone marrow transplantation breaks graft-host tolerance allows for persistent*
456 *antileukemic reactivity without severe graft-versus-host disease*. Blood, 1995. **85**(11): p.
457 3302-3312.
- 458 13. Ferrara, J.L., et al., *Graft-versus-host disease*. The Lancet, 2009. **373**(9674): p. 1550-
459 1561.
- 460 14. Teshima, T., et al., *Acute graft-versus-host disease does not require alloantigen*
461 *expression on host epithelium*. Nature medicine, 2002. **8**(6): p. 575-581.
- 462 15. Newton-Nash, D.K., *The molecular basis of allorecognition assessment of the*
463 *involvement of peptide*. Human immunology, 1994. **41**(2): p. 105-111.
- 464 16. Markey, K.A., et al., *Conventional dendritic cells are the critical donor APC presenting*
465 *alloantigen after experimental bone marrow transplantation*. Blood, 2009. **113**(22): p.
466 5644-5649.
- 467 17. Sprent, J., et al., *Role of T cell subsets in lethal graft-versus-host disease (GVHD)*
468 *directed to class I versus class II H-2 differences. I. L3T4+ cells can either augment or*
469 *retard GVHD elicited by Lyt-2+ cells in class I different hosts*. The Journal of
470 experimental medicine, 1988. **167**(2): p. 556-569.

- 471 18. Goulmy, E., et al., *Mismatches of minor histocompatibility antigens between HLA-*
472 *identical donors and recipients and the development of graft-versus-host disease after*
473 *bone marrow transplantation.* New England Journal of Medicine, 1996. **334**(5): p. 281-
474 285.
- 475 19. den Haan, J.M., et al., *The minor histocompatibility antigen HA-I: a diallelic gene with a*
476 *single amino acid polymorphism.* Science, 1998. **279**(5353): p. 1054-7.
- 477 20. den Haan, J.M., et al., *Identification of a graft versus host disease-associated human*
478 *minor histocompatibility antigen.* Science, 1995. **268**(5216): p. 1476-80.
- 479 21. Goulmy, E., et al., *A minor transplantation antigen detected by MHC-restricted cytotoxic*
480 *T lymphocytes during graft-versus-host disease.* Nature, 1983. **302**(5904): p. 159-61.
- 481 22. Goulmy, E., et al., *Y-antigen killing by T cells of women is restricted by HLA.* Nature,
482 1977. **266**(5602): p. 544-5.
- 483 23. Popli, R., et al., *Clinical impact of H-Y alloimmunity.* Immunol Res, 2014. **58**(2-3): p.
484 249-58.
- 485 24. Spierings, E., *Minor histocompatibility antigens: past, present, and future.* Tissue
486 Antigens, 2014. **84**(4): p. 374-60.
- 487 25. de Bueger, M., et al., *Tissue distribution of human minor histocompatibility antigens.*
488 *Ubiquitous versus restricted tissue distribution indicates heterogeneity among human*
489 *cytotoxic T lymphocyte-defined non-MHC antigens.* J Immunol, 1992. **149**(5): p. 1788-
490 94.
- 491 26. Nakasone, H., et al., *Allogeneic HY antibodies detected 3 months after female-to-male*
492 *HCT predict chronic GVHD and nonrelapse mortality in humans.* Blood, 2015. **125**(20):
493 p. 3193-201.
- 494 27. Miklos, D.B., et al., *Antibody responses to H-Y minor histocompatibility antigens*
495 *correlate with chronic graft-versus-host disease and disease remission.* Blood, 2005.
496 **105**(7): p. 2973-8.
- 497 28. Nash, R.A., et al., *Phase 3 study comparing methotrexate and tacrolimus with*
498 *methotrexate and cyclosporine for prophylaxis of acute graft-versus-host disease after*
499 *marrow transplantation from unrelated donors.* Blood, 2000. **96**(6): p. 2062-2068.
- 500 29. Pinana, J.L., et al., *MTX or mycophenolate mofetil with CsA as GVHD prophylaxis after*
501 *reduced-intensity conditioning PBSCT from HLA-identical siblings.* Bone Marrow
502 Transplant, 2010. **45**(9): p. 1449-56.
- 503 30. Bacigalupo, A., et al., *Antithymocyte globulin for graft-versus-host disease prophylaxis in*
504 *transplants from unrelated donors: 2 randomized studies from Gruppo Italiano Trapianti*
505 *Midollo Osseo (GITMO).* Blood, 2001. **98**(10): p. 2942-2947.
- 506 31. Storb, R., et al., *Stable mixed hematopoietic chimerism in DLA-identical littermate dogs*
507 *given sublethal total body irradiation before and pharmacological immunosuppression*
508 *after marrow transplantation.* Blood, 1997. **89**(8): p. 3048-3054.
- 509 32. Kagi, D., et al., *Fas and perforin pathways as major mechanisms of T cell-mediated*
510 *cytotoxicity.* Science, 1994. **265**(5171): p. 528-30.
- 511 33. Lowin, B., et al., *Cytolytic T-cell cytotoxicity is mediated through perforin and Fas lytic*
512 *pathways.* Nature, 1994. **370**(6491): p. 650-652.
- 513 34. Couriel, D., et al., *Tumor necrosis factor-alpha blockade for the treatment of acute*
514 *GVHD.* Blood, 2004. **104**(3): p. 649-54.
- 515 35. Westin, J.R., et al., *Steroid-refractory acute GVHD: predictors and outcomes.* Advances
516 in hematology, 2011. **2011**.

- 517 36. Apostolova, P. and R. Zeiser, *The role of danger signals and ectonucleotidases in acute*
518 *graft-versus-host disease*. Human immunology, 2016.
- 519 37. Penack, O., E. Holler, and M.R. van den Brink, *Graft-versus-host disease: regulation by*
520 *microbe-associated molecules and innate immune receptors*. Blood, 2010. **115**(10): p.
521 1865-1872.
- 522 38. Penack, O., et al., *NOD2 regulates hematopoietic cell function during graft-versus-host*
523 *disease*. The Journal of experimental medicine, 2009. **206**(10): p. 2101-2110.
- 524 39. Wehkamp, J., et al., *NOD2 (CARD15) mutations in Crohn's disease are associated with*
525 *diminished mucosal α -defensin expression*. Gut, 2004. **53**(11): p. 1658-1664.
- 526 40. Mathewson, N.D., et al., *Gut microbiome-derived metabolites modulate intestinal*
527 *epithelial cell damage and mitigate graft-versus-host disease*. Nature immunology, 2016.
- 528 41. Peled, J.U., et al., *Role of gut flora after bone marrow transplantation*. Nature
529 Microbiology, 2016. **1**: p. 16036.
- 530 42. Edinger, M., et al., *CD4+ CD25+ regulatory T cells preserve graft-versus-tumor activity*
531 *while inhibiting graft-versus-host disease after bone marrow transplantation*. Nature
532 medicine, 2003. **9**(9): p. 1144-1150.
- 533 43. Bollrath, J. and F.M. Powrie. *Controlling the frontier: regulatory T-cells and intestinal*
534 *homeostasis*. in *Seminars in immunology*. 2013. Elsevier.
- 535 44. Schneidawind, D., et al., *Third-party CD4+ invariant natural killer T cells protect from*
536 *murine GVHD lethality*. Blood, 2015. **125**(22): p. 3491-3500.
- 537 45. Hanash, A.M., et al., *Interleukin-22 protects intestinal stem cells from immune-mediated*
538 *tissue damage and regulates sensitivity to graft versus host disease*. Immunity, 2012.
539 **37**(2): p. 339-350.
- 540 46. Einsele, H., et al., *Infusion of cytomegalovirus (CMV)-specific T cells for the treatment of*
541 *CMV infection not responding to antiviral chemotherapy*. Blood, 2002. **99**(11): p. 3916-
542 22.
- 543 47. Feuchtinger, T., et al., *Adoptive transfer of pp65-specific T cells for the treatment of*
544 *chemorefractory cytomegalovirus disease or reactivation after haploidentical and*
545 *matched unrelated stem cell transplantation*. Blood, 2010. **116**(20): p. 4360-7.
- 546 48. Wang, X.N., et al., *Regulatory T-cell suppression of CD8+ T-cell-mediated graft-versus-*
547 *host reaction requires their presence during priming*. Transplantation, 2009. **88**(2): p.
548 188-97.
- 549 49. Trzonkowski, P., et al., *First-in-man clinical results of the treatment of patients with graft*
550 *versus host disease with human ex vivo expanded CD4+CD25+CD127- T regulatory*
551 *cells*. Clin Immunol, 2009. **133**(1): p. 22-6.
- 552 50. Brunstein, C.G., et al., *Infusion of ex vivo expanded T regulatory cells in adults*
553 *transplanted with umbilical cord blood: safety profile and detection kinetics*. Blood,
554 2011. **117**(3): p. 1061-70.
- 555 51. Di Ianni, M., et al., *Tregs prevent GVHD and promote immune reconstitution in HLA-*
556 *haploidentical transplantation*. Blood, 2011. **117**(14): p. 3921-8.
- 557 52. Geissler, E.K., *The ONE Study compares cell therapy products in organ transplantation:*
558 *introduction to a review series on suppressive monocyte-derived cells*. Transplantation
559 Research, 2012. **1**: p. 11-11.
- 560 53. Mavin, E., et al., *Regulatory T cells inhibit CD8(+) T-cell tissue invasion in human skin*
561 *graft-versus-host reactions*. Transplantation, 2012. **94**(5): p. 456-64.

- 562 54. Mavin, E., et al., *Human Regulatory T Cells Mediate Transcriptional Modulation of*
563 *Dendritic Cell Function*. J Immunol, 2017. **198**(1): p. 138-146.
- 564 55. Ruffin, N., et al., *The involvement of epithelial Fas in a human model of graft versus host*
565 *disease*. Transplantation, 2011. **91**(9): p. 946-51.
- 566 56. Mori, T., et al., *Involvement of Fas-mediated apoptosis in the hematopoietic progenitor*
567 *cells of graft-versus-host reaction-associated myelosuppression*. Blood, 1998. **92**(1): p.
568 101-7.
- 569 57. Ahmed, S.S., et al., *Identification and validation of biomarkers associated with acute and*
570 *chronic graft versus host disease*. Bone Marrow Transplant, 2015. **50**(12): p. 1563-71.
- 571 58. Zinöcker, S., et al., *Immune Reconstitution and Graft-Versus-Host Reactions in Rat*
572 *Models of Allogeneic Hematopoietic Cell Transplantation*. Frontiers in Immunology,
573 2012. **3**: p. 355.
- 574 59. Novota, P., et al., *Expression profiling of major histocompatibility and natural killer*
575 *complex genes reveals candidates for controlling risk of graft versus host disease*. PLoS
576 One, 2011. **6**(1): p. e16582.
- 577 60. Harris, A.C., J.L. Ferrara, and J.E. Levine, *Advances in predicting acute GVHD*. Br J
578 Haematol, 2013. **160**(3): p. 288-302.
- 579 61. Paczesny, S., et al., *A biomarker panel for acute graft-versus-host disease*. Blood, 2009.
580 **113**(2): p. 273-8.
- 581 62. Ferrara, J.L., et al., *Regenerating islet-derived 3-alpha is a biomarker of gastrointestinal*
582 *graft-versus-host disease*. Blood, 2011. **118**(25): p. 6702-8.
- 583 63. Harris, A.C., et al., *Plasma biomarkers of lower gastrointestinal and liver acute GVHD*.
584 Blood, 2012. **119**(12): p. 2960-2963.
- 585 64. Levine, J.E., et al., *Acute graft-versus-host disease biomarkers measured during therapy*
586 *can predict treatment outcomes: a Blood and Marrow Transplant Clinical Trials*
587 *Network study*. Blood, 2012. **119**(16): p. 3854-60.
- 588 65. Levine, J.E., et al., *A prognostic score for acute graft-versus-host disease based on*
589 *biomarkers: A multicentre study*. The Lancet Haematology, 2015. **2**(1): p. e21-e29.
- 590 66. Lee, S.J., et al., *Severity of chronic graft-versus-host disease: association with treatment-*
591 *related mortality and relapse*. Blood, 2002. **100**(2): p. 406-414.
- 592 67. Shimabukuro-Vornhagen, A., et al., *The role of B cells in the pathogenesis of graft-*
593 *versus-host disease*. Blood, 2009. **114**(24): p. 4919-4927.
- 594 68. Jagasia, M.H., et al., *National institutes of health consensus development project on*
595 *criteria for clinical trials in chronic graft-versus-host disease: I. The 2014 diagnosis and*
596 *staging working group report*. Biology of Blood and Marrow Transplantation, 2015.
597 **21**(3): p. 389-401. e1.
- 598 69. Socié, G. and J. Ritz, *Current issues in chronic graft-versus-host disease*. Blood, 2014.
599 **124**(3): p. 374-384.
- 600 70. Sarantopoulos, S. and J. Ritz, *Aberrant B-cell homeostasis in chronic GVHD*. Blood,
601 2015. **125**(11): p. 1703-1707.
- 602 71. Varelias, A., et al., *Lung parenchyma-derived IL-6 promotes IL-17A-dependent acute*
603 *lung injury after allogeneic stem cell transplantation*. Blood, 2015. **125**(15): p. 2435-
604 2444.
- 605 72. Griffith, L.M., et al., *Chronic Graft-versus-Host Disease-Implementation of the National*
606 *Institutes of Health Consensus Criteria for Clinical Trials*. Biology of blood and marrow

- 607 transplantation: journal of the American Society for Blood and Marrow Transplantation,
608 2008. **14**(4): p. 379.
- 609 73. Peñas, P.F. and S. Zaman, *Many faces of graft-versus-host disease*. Australasian Journal
610 of Dermatology, 2010. **51**(1): p. 1-10.
- 611 74. Johnson, M.L. and E.R. Farmer, *Graft-versus-host reactions in dermatology*. Journal of
612 the American Academy of Dermatology, 1998. **38**(3): p. 369-392.
- 613 75. Paczesny, S., et al., *Elafin is a biomarker of graft-versus-host disease of the skin*. Science
614 translational medicine, 2010. **2**(13): p. 13ra2-13ra2.
- 615 76. Snover, D.C., et al., *Hepatic graft versus host disease: a study of the predictive value of*
616 *liver biopsy in diagnosis*. Hepatology, 1984. **4**(1): p. 123-130.
- 617 77. McDonald, G.B., *Hepatobiliary complications of hematopoietic cell transplantation, 40*
618 *years on*. Hepatology, 2010. **51**(4): p. 1450-1460.
- 619 78. Snover, D.C., et al., *A histopathologic study of gastric and small intestinal graft-versus-*
620 *host disease following allogeneic bone marrow transplantation*. Human pathology, 1985.
621 **16**(4): p. 387-392.
- 622 79. Levine, J.E., et al., *Low Paneth cell numbers at onset of gastrointestinal graft-versus-host*
623 *disease identify patients at high risk for nonrelapse mortality*. Blood, 2013. **122**(8): p.
624 1505-1509.
- 625 80. Weber, D., et al., *Low urinary indoxyl sulfate levels early after transplantation reflect a*
626 *disrupted microbiome and are associated with poor outcome*. Blood, 2015. **126**(14): p.
627 1723-1728.
- 628 81. Eriguchi, Y., et al., *Graft-versus-host disease disrupts intestinal microbial ecology by*
629 *inhibiting Paneth cell production of α -defensins*. Blood, 2012. **120**(1): p. 223-231.
- 630 82. Gooley, T.A., et al., *Reduced mortality after allogeneic hematopoietic-cell*
631 *transplantation*. New England Journal of Medicine, 2010. **363**(22): p. 2091-2101.
- 632 83. Bhatia, S., *Caring for the long-term survivor after allogeneic stem cell transplantation*.
633 ASH Education Program Book, 2014. **2014**(1): p. 495-503.
- 634 84. Konopacki, J., et al., *Long-term follow up after allogeneic stem cell transplantation in*
635 *patients with severe aplastic anemia after cyclophosphamide plus antithymocyte globulin*
636 *conditioning*. Haematologica, 2012. **97**(5): p. 710-716.
- 637 85. Mohty, M., et al., *Sinusoidal obstruction syndrome/veno-occlusive disease: current*
638 *situation and perspectives—a position statement from the European Society for Blood*
639 *and Marrow Transplantation (EBMT)*. Bone marrow transplantation, 2015. **50**(6): p. 781-
640 789.
- 641 86. Carreras, E., *How I manage sinusoidal obstruction syndrome after haematopoietic cell*
642 *transplantation*. British journal of haematology, 2015. **168**(4): p. 481-491.
- 643 87. Stavrou, E. and H.M. Lazarus, *Thrombotic microangiopathy in haematopoietic cell*
644 *transplantation: an update*. Mediterranean journal of hematology and infectious diseases,
645 2010. **2**(3): p. 2010033.
- 646 88. Jodele, S., et al., *Diagnostic and risk criteria for HSCT-associated thrombotic*
647 *microangiopathy: a study in children and young adults*. Blood, 2014. **124**(4): p. 645-653.
- 648 89. Jodele, S., et al., *A new paradigm: diagnosis and management of HSCT-associated*
649 *thrombotic microangiopathy as multi-system endothelial injury*. Blood reviews, 2015.
650 **29**(3): p. 191-204.
- 651 90. Palomo, M., et al., *The release of soluble factors contributing to endothelial activation*
652 *and damage after hematopoietic stem cell transplantation is not limited to the allogeneic*

- 653 *setting and involves several pathogenic mechanisms. Biology of Blood and Marrow*
654 *Transplantation, 2009. 15(5): p. 537-546.*
- 655 91. Leonhardt, F., et al., *Inflammatory neovascularization during graft-versus-host disease is*
656 *regulated by α integrin and miR-100. Blood, 2013. 121(17): p. 3307-3318.*
- 657 92. Penack, O., et al., *Inhibition of neovascularization to simultaneously ameliorate graft-vs-*
658 *host disease and decrease tumor growth. Journal of the National Cancer Institute, 2010.*
659 **102(12): p. 894-908.**
- 660 93. Penack, O., G. Socié, and M.R. van den Brink, *The importance of neovascularization and*
661 *its inhibition for allogeneic hematopoietic stem cell transplantation. Blood, 2011.*
662 **117(16): p. 4181-4189.**
- 663 94. Schmid, P.M., et al., *Endothelial dysfunction and altered mechanical and structural*
664 *properties of resistance arteries in a murine model of graft-versus-host disease. Biology*
665 *of Blood and Marrow Transplantation, 2014. 20(10): p. 1493-1500.*
- 666 95. Biedermann, B.C., *Vascular endothelium and graft-versus-host disease. Best Practice &*
667 *Research Clinical Haematology, 2008. 21(2): p. 129-138.*
- 668 96. Biedermann, B.C., et al., *Endothelial injury mediated by cytotoxic T lymphocytes and loss*
669 *of microvessels in chronic graft versus host disease. The Lancet, 2002. 359(9323): p.*
670 *2078-2083.*
- 671 97. Andrulis, M., et al., *Loss of endothelial thrombomodulin predicts response to steroid*
672 *therapy and survival in acute intestinal graft-versus-host disease. haematologica, 2012:*
673 *p. haematol. 2011.061051.*
- 674 98. Luft, T., et al., *Steroid-refractory GVHD: T-cell attack within a vulnerable endothelial*
675 *system. Blood, 2011. 118(6): p. 1685-1692.*
- 676 99. Rachakonda, S.P., et al., *Single-nucleotide polymorphisms within the thrombomodulin*
677 *gene (THBD) predict mortality in patients with graft-versus-host disease. Journal of*
678 *Clinical Oncology, 2014. 32(30): p. 3421-3427.*
- 679 100. Holtan, S.G., et al., *Circulating angiogenic factors associated with response and survival*
680 *in patients with acute graft-versus-host disease: results from Blood and Marrow*
681 *Transplant Clinical Trials Network 0302 and 0802. Biology of Blood and Marrow*
682 *Transplantation, 2015. 21(6): p. 1029-1036.*
- 683 101. Tomblyn, M., et al., *Guidelines for preventing infectious complications among*
684 *hematopoietic cell transplantation recipients: a global perspective. Biology of Blood and*
685 *Marrow Transplantation, 2009. 15(10): p. 1143-1238.*
- 686 102. Maschmeyer, G. and J.P. Donnelly, *How to manage lung infiltrates in adults suffering*
687 *from haematological malignancies outside allogeneic haematopoietic stem cell*
688 *transplantation. British journal of haematology, 2016.*
- 689 103. Rieger, C., et al., *Infectious complications after allogeneic stem cell transplantation:*
690 *incidence in matched-related and matched-unrelated transplant settings. Transplant*
691 *Infectious Disease, 2009. 11(3): p. 220-226.*
- 692 104. Hildebrandt, G., et al., *Diagnosis and treatment of pulmonary chronic GVHD: report*
693 *from the consensus conference on clinical practice in chronic GVHD. Bone marrow*
694 *transplantation, 2011. 46(10): p. 1283-1295.*
- 695 105. Wolff, D. and G. Hildebrandt, *Bronchiolitis obliterans—pleading for a pragmatic*
696 *approach. Biology of Blood and Marrow Transplantation, 2016.*

- 697 106. Cheng, G.-S., et al., *Outcomes of lung transplantation after allogeneic hematopoietic*
698 *stem cell transplantation*. *Biology of Blood and Marrow Transplantation*, 2014. **20**(8): p.
699 1169-1175.
- 700 107. Sengsayadeth, S.M., et al., *Time to explore preventive and novel therapies for*
701 *bronchiolitis obliterans syndrome after allogeneic hematopoietic stem cell*
702 *transplantation*. *Biology of Blood and Marrow Transplantation*, 2012. **18**(10): p. 1479-
703 1487.
- 704 108. Bacigalupo, A., et al. *Late pulmonary complications after allogeneic hematopoietic stem*
705 *cell transplantation: diagnosis, monitoring, prevention, and treatment*. in *Seminars in*
706 *hematology*. 2012. Elsevier.
- 707 109. Edinger, M. and P. Hoffmann, *Regulatory T cells in stem cell transplantation: strategies*
708 *and first clinical experiences*. *Current opinion in immunology*, 2011. **23**(5): p. 679-684.
- 709 110. Le Blanc, K., et al., *Mesenchymal stem cells for treatment of steroid-resistant, severe,*
710 *acute graft-versus-host disease: a phase II study*. *The Lancet*, 2008. **371**(9624): p. 1579-
711 1586.
- 712 111. Le Blanc, K., et al., *Treatment of severe acute graft-versus-host disease with third party*
713 *haploidentical mesenchymal stem cells*. *The Lancet*, 2004. **363**(9419): p. 1439-1441.
- 714 112. Moll, G. and K. Le Blanc, *Engineering more efficient multipotent mesenchymal stromal*
715 *(stem) cells for systemic delivery as cellular therapy*. *ISBT Science Series*, 2015. **10**(S1):
716 p. 357-365.
- 717 113. Antin, J.H., et al., *Sirolimus, tacrolimus, and low-dose methotrexate for graft-versus-host*
718 *disease prophylaxis in mismatched related donor or unrelated donor transplantation*.
719 *Blood*, 2003. **102**(5): p. 1601-1605.
- 720 114. Finke, J., et al., *Matched and Mismatched Allogeneic Stem-Cell Transplantation From*
721 *Unrelated Donors Using Combined Graft-Versus-Host Disease Prophylaxis Including*
722 *Rabbit Anti-T Lymphocyte Globulin*. *Journal of clinical oncology*, 2003. **21**(3): p. 506-
723 513.
- 724 115. Luznik, L., et al., *HLA-haploidentical bone marrow transplantation for hematologic*
725 *malignancies using nonmyeloablative conditioning and high-dose, posttransplantation*
726 *cyclophosphamide*. *Biol Blood Marrow Transplant*, 2008. **14**(6): p. 641-50.
- 727 116. Taur, Y., et al., *Role of intestinal microbiota in transplantation outcomes*. *Best Practice*
728 *& Research Clinical Haematology*, 2015. **28**(2): p. 155-161.
- 729 117. Pierini, A., M. Alvarez, and R.S. Negrin, *NK Cell and CD4+ FoxP3+ Regulatory T Cell*
730 *Based Therapies for Hematopoietic Stem Cell Engraftment*. *Stem Cells International*,
731 2016.
- 732 118. Theil, A., et al., *Adoptive transfer of allogeneic regulatory T cells into patients with*
733 *chronic graft-versus-host disease*. *Cytherapy*, 2015. **17**(4): p. 473-486.
- 734 119. Koreth, J., et al., *Interleukin-2 and regulatory T cells in graft-versus-host disease*. *New*
735 *England Journal of Medicine*, 2011. **365**(22): p. 2055-2066.
- 736 120. Dall'Amico, R. and C. Messina, *Extracorporeal photochemotherapy for the treatment of*
737 *graft-versus-host disease*. *Therapeutic Apheresis*, 2002. **6**(4): p. 296-304.
- 738 121. Gatza, E., et al., *Extracorporeal photopheresis reverses experimental graft-versus-host*
739 *disease through regulatory T cells*. *Blood*, 2008. **112**(4): p. 1515-21.
- 740 122. Koehn, B.H., et al., *GVHD-associated, inflammasome-mediated loss of function in*
741 *adoptively transferred myeloid-derived suppressor cells*. *Blood*, 2015. **126**(13): p. 1621-
742 1628.

- 743 123. Ball, L.M., et al., *Multiple infusions of mesenchymal stromal cells induce sustained*
744 *remission in children with steroid-refractory, grade III–IV acute graft-versus-host*
745 *disease*. *British journal of haematology*, 2013. **163**(4): p. 501-509.
- 746 124. Ringdén, O., et al., *Mesenchymal stem cells for treatment of therapy-resistant graft-*
747 *versus-host disease*. *Transplantation*, 2006. **81**(10): p. 1390-1397.
- 748 125. Teshima, T., P. Reddy, and R. Zeiser, *Acute Graft-versus-Host Disease: Novel Biological*
749 *Insights*. *Biology of Blood and Marrow Transplantation*, 2016. **22**(1): p. 11-16.
- 750 126. Wolff, D., et al., *The Treatment of Chronic Graft-Versus-Host Disease*. *Dtsch Arztebl*
751 *Int*, 2011. **108**: p. 732-40.

752

753

754

755

756

757

758

759

760

761

762

763 **Figure legends**

764 **Figure 1. Initiation of GvHD.** Conditioning regimen leads to destruction of epithelial cells
765 and their integrity. Damaged epithelia secrete uric acid and adenosine triphosphate (ATP) that
766 results in production of pro-inflammatory cytokines. Pathogen recognition receptors (PRRs)
767 such as Toll-like receptors (TLRs), NOD-like receptors (NLRs) and P2XRs are activated by
768 pathogen associated molecular patterns (PAMPs) and danger associated molecular patterns
769 (DAMPs). These signals ultimately activates antigen presenting cells (APCs) that leads to
770 donor T cell activation. Adopted and modified from [37]

771 **Figure 2. Pathophysiology of acute GvHD.** Conditioning regimen cause profound damage to
772 the host tissues leading to release of inflammatory cytokines like TNF, IL-1. These cytokines
773 activate host APCs in phase I. In addition, loss of microbial diversity and metabolites thereof
774 leads to loss of epithelial and immune homeostasis. Host APCs activate mature donor T cells
775 present in stem cell inoculum in phase II. T cells subsequently proliferate and differentiate into
776 Th1 and Th17 type which are involved in activation of CD4 CTL, CD8 CTL and NK cells that
777 mediate tissue damage. In phase III, effector T cells together with storm of pro-inflammatory
778 cytokine attack the epithelial cells of skin, liver, lung and GI tract. This damage is further

779 supported by the LPS that has leaked through damaged intestinal mucosa which then recruit
780 myeloid cell to further produce pro-inflammatory cytokines and thus enhances the cytokine
781 storm. Adopted and modified from [13].

782 **Figure 3. Skin explant grades I –IV**

783 The outcome of the skin explant assay is histopathological damage ranging from grade I
784 GvHR (with minimal vacuolisation in the epidermis) to Grade II GvHR (with vacuolisation
785 and dyskeratotic bodies) to Grade III GvHR (with sub epidermal cleft formation) and finally
786 to Grade IV GvHR (with separation of the dermis from the epidermis).

787

788

Provisional

Figure 01.TIF

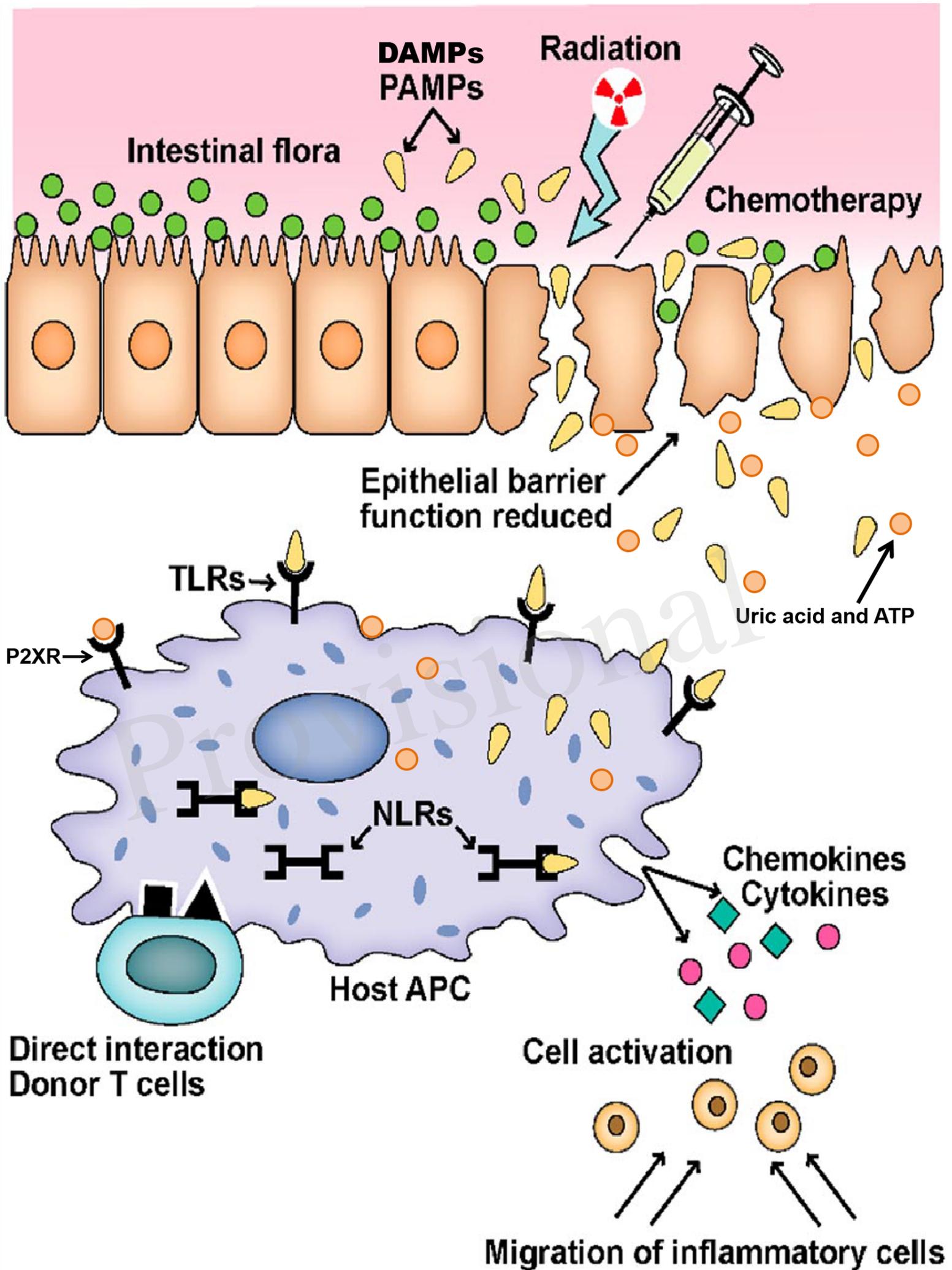


Figure 02.TIF

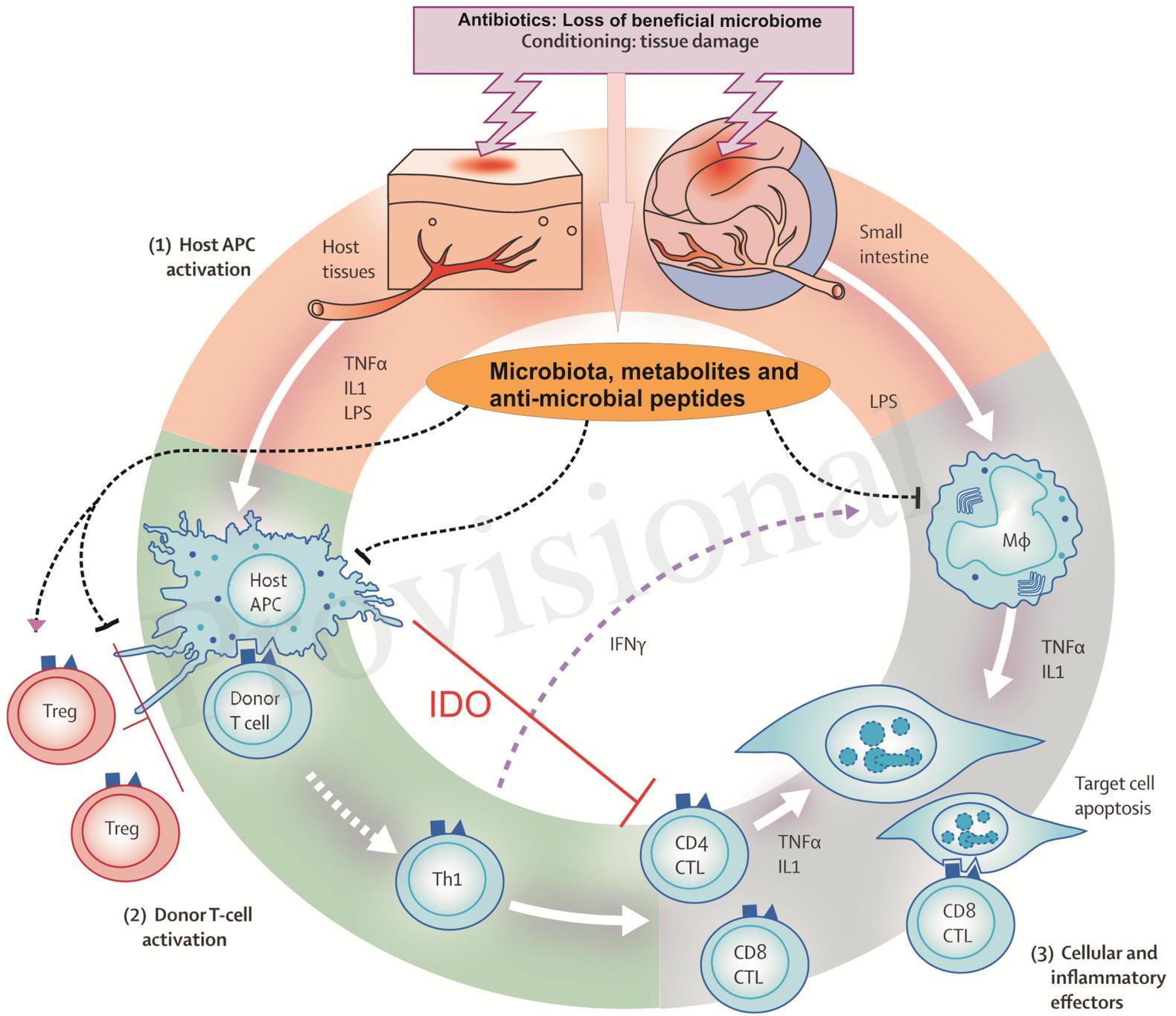


Figure 03.TIF

