


Article

# Synthesis, Characterization, and Self-Assembly of a Tetrathiafulvalene (TTF)–Triglycyl Derivative

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**Abstract:** In this work, we describe the synthesis, characterization, and self-assembly properties of a new tetrathiafulvalene (TTF)–triglycyl low-molecular-weight (LMW) gelator. Supramolecular organogels were obtained in various solvents via a heating–cooling cycle. Critical gelation concentrations (CGC) (range  $\approx$  5–50 g/L) and thermal *gel-to-sol* transition temperatures ( $T_{gel}$ ) (range  $\approx$  36–51 °C) were determined for each gel. Fourier transform infrared (FT-IR) spectroscopy suggested that the gelator is also aggregated in its solid state via a similar hydrogen-bonding pattern. The fibrillar microstructure and viscoelastic properties of selected gels were demonstrated by means of field-emission electron microscopy (FE-SEM) and rheological measurements. As expected, exposure of a model xerogel to I<sub>2</sub> vapor caused the oxidation of the TTF unit as confirmed by UV-vis-NIR analysis. However, FT-IR spectroscopy showed that the oxidation was accompanied with concurrent alteration of the hydrogen-bonded network.

**Keywords:** tetrathiafulvalene; triglycyl peptide; self-assembly; organogel

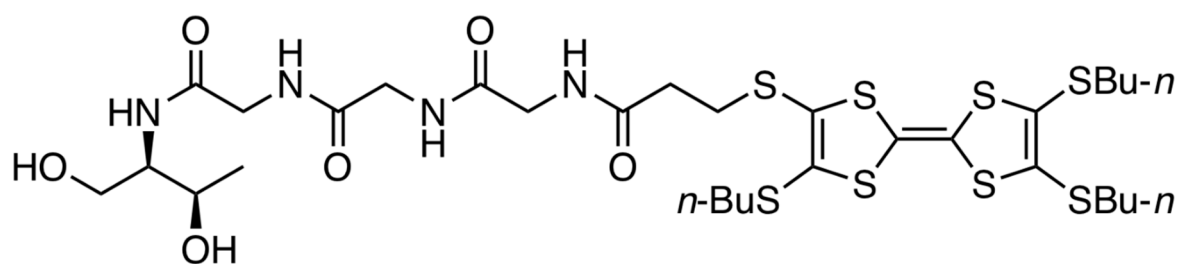
## 1. Introduction

Oligonucleotide conjugates carrying aromatic systems are important tools for a large number of applications due to their enhanced hybridization properties and their special fluorescence and chemical properties coming from the aromatic moieties [1–6]. Recently, tetrathiafulvalene (TTF) derivatives have been incorporated in oligonucleotides demonstrating the fluorescence-quenching properties of the DNA–TTF conjugates upon hybridization [7], the preservation of the electrochemical properties of TTF [8], an enhanced affinity to complementary sequences [9], and compatibility with the RNA interference mechanism for gene inhibition [10]. In the field of materials chemistry, TTF and its derivatives constitute electron donors that can form charge-transfer (CT) complexes and have been widely studied for the development of electrically conducting materials [11,12]. TTF moiety can be consecutively oxidized to the radical cation TTF<sup>•+</sup> and dication TTF<sup>2+</sup> by either electrochemical or chemical processes. Due to the reversibility of these transformations [13], TTF has become a general building block for the fabrication of switchable functional materials [14–17], including supramolecular self-assembled nanostructures [18–21].

The interesting properties of the TTF-oligonucleotides prompted us to synthesize a TTF derivative carrying a biodegradable triglycyl peptide linker between the oligonucleotide and the TTF molecule. This simple peptide sequence has been demonstrated to be hydrolyzed by cathepsin B present in endosomes [22]. During the preparation of the mentioned TTF derivative we accidentally observed the formation of gels in some solvents. In general, gels have received great attention during the last decade [23–34] due to their exceptional hierarchical architectures and potential applications in many important areas such as biomedicine, health care, and catalysis, among others [35–45]. Chemical gels are based on covalent bonds, usually cross-linked polymers [46–48], whereas physical gels (also called supramolecular gels) are normally fabricated using low-molecular-weight (LMW) molecules that self-assemble via non-covalent interactions. For this reason, supramolecular gels usually show reversible stimuli-responsive *gel-to-sol* transitions [23,24]. Systems based on both covalent and non-covalent interactions are also known [49]. Typically, the entanglement of 1D nanofibers of gelator molecules generates a 3D-network with the solvent molecules trapped into the interstices by capillary forces. This provides a solid-like appearance and viscoelastic features to physical gels [50–53].

Within this context, Jørgensen and co-workers reported in 1994 [54] the first TTF-based LMW gelator bearing bis-arborol units, which was used to fabricate a self-assembling “molecular” nanowire [55]. Since then, a large number of electroactive TTF-based LMW gelators and their gels have been described in the literature [56]. In general, the rational design behind these examples involves the connection of the TTF unit with groups capable of H-bonding, such as urea [57] or amide groups [58–63], to form extended structures. The incorporation of other molecular building blocks commonly used to form physical gels [64–67] or the use of metal–organic interactions [68–70] have also been employed to form TTF-based gels. Moreover, the inclusion of hydrophobic aliphatic chains is also employed to facilitate van der Waals interactions and stabilize the supramolecular aggregates [58–71]. If formation of CT complexes takes place, the TTF unit will bear positive charge, which can alter the interaction of neighboring TTF units and influence the intermolecular interactions arising from other groups involved in the self-assembly process. Indeed, the above-mentioned studies have confirmed the possibility of tuning the gel formation by CT interaction and oxidation. In addition, conductive gels have also been fabricated by simple mixing and mechanically grinding neutral TTF and an acceptor (i.e., tetracyanoquinodimethane) in ionic liquids [72]. More recently, gels made of TTF–dipeptide and tripeptide conjugates have been reported [73,74].

In this communication we describe the synthesis, characterization, and gelation properties of a new TTF–triglycyl-based LMW organogelator (Figure 1). As far as we are aware, a triglycyl peptide has not been used to fabricate TTF-based gelators.



**Figure 1.** Chemical structure of tetrathiafulvalene (TTF)–triglycyl derivative studied in this work.

## 2. Materials and Methods

### 2.1. Materials

Unless otherwise noted, all reagents and dry solvents were purchased from commercial suppliers (Sigma-Aldrich, St. Louis, MO, USA, TCI Chemicals, Tokyo, Japan) and used as received.

## 2.2. Synthesis and Characterization of Compounds

### 2.2.1. General Remarks

Nuclear magnetic resonance (NMR) spectra,  $^1\text{H}$ -NMR (400 MHz), and  $^{13}\text{C}$ -NMR (100 MHz) were recorded using Varian Mercury 400 spectrometer at room temperature (RT). Tetramethylsilane (TMS) was used as internal reference (0 ppm) for  $^1\text{H}$  spectra recorded in  $\text{CDCl}_3$ . Chemical shifts for  $^1\text{H}$ -NMR and  $^{13}\text{C}$ -NMR were reported as  $\delta$ , parts per million (ppm), relative to the signal of the residual solvent ( $^1\text{H}$ -NMR:  $\text{CHCl}_3 = 7.26$  ppm, DMSO = 2.50 ppm;  $^{13}\text{C}$ -NMR:  $\text{CHCl}_3 = 77.16$  ppm, DMSO = 39.5 ppm;  $\text{CD}_3\text{OD} = 49.0$  ppm). Coupling constants ( $J$ ) are given in Hertz (Hz). Multiplicity notations: s = singlet, bs = broad singlet, d = doublet, t = triplet, q = quartet, dd = doublet of doublets, m = multiplet. Estimated error of reported values: 0.01 ppm ( $\delta$ ,  $^1\text{H}$ -NMR), 0.1 ppm ( $\delta$ ,  $^{13}\text{C}$ -NMR), 0.1 Hz ( $J$ , coupling constant). Copies of the NMR spectra are provided in the Supporting Information. Fourier transform infrared (FT-IR) spectra were recorded on a Thermo-Nicolet Avatar 380 FT-IR equipped with a SMART iTR sampling accessory. Electrospray ionization mass spectra (ESI-MS) were recorded on a Micromass ZQ instrument with single quadrupole detector coupled to an HPLC and high-resolution (HR) ESI-MS on an Agilent 1100 LC/MS-TOF instrument (Servei d'Espectrometria de Masses, Universitat de Barcelona). Flash chromatography was performed using silica gel (230–400 mesh) as stationary phase. Thin-layer chromatography (TLC) analysis were carried out on silica gel 60 F<sub>254</sub> aluminum sheets and visualized under UV light (365 nm).

### 2.2.2. Synthetic Procedures and Characterization of Compounds

**4,5-Bis-(butylthio)-1,3-dithiole-2-thione (3):** A mixture of bis(tetrabutylammonium) bis(1,3-dithiole-2-thione-4,5-dithiolate) zinc (**1**, TCI chemicals) (2.18 g, 2.30 mmol) and 1-bromobutane (**2**, Sigma-Aldrich) (1.24 mL, 11.50 mmol) in anhydrous acetonitrile (ACN) (30 mL) was refluxed for 1 h. The initially formed reddish solution turned into a green-yellowish mixture. Once the solution was cooled to room temperature (RT), the precipitate was filtered out and the solution mixture was concentrated to dryness under reduced pressure. The residue was dissolved in dichloromethane (DCM) (50 mL) and the organic phase was washed with water (4 × 30 mL). The organic phase was dried over anhydrous  $\text{MgSO}_4$  and concentrated to dryness under reduced pressure. The residue was purified by flash chromatography on silica gel (hexane/ethyl acetate (AcOEt) 1:1) affording the desired compound **3** as reddish oil (1.40 g, 98% yield). TLC in AcOEt/hexane 10:0.1,  $R_f = 0.85$ .  $^1\text{H}$ -NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 2.85 (t,  $J = 7.3$  Hz, 4H, =C-SCH<sub>2</sub>-), 1.67–1.59 (m, 4H, =C-SCH<sub>2</sub>-CH<sub>2</sub>-), 1.48–1.38 (m, 4H, =C-SCH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-), 0.91 (t,  $J = 7.3$  Hz, 6H, CH<sub>3</sub>).  $^{13}\text{C}$ -NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 211.49 (C=S), 136.34 (S-C-S), 36.42 (S-CH<sub>2</sub>-), 31.64 (S-CH<sub>2</sub>-CH<sub>2</sub>-), 21.63 (S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-), 13.51 (-CH<sub>3</sub>). IR (oil):  $\nu = 2954$  m, 2925 m and 2869 w (C-H stretch), 1461 m (H-C-H bend), 1060 s (C=S stretch), 884 m (H-C-S bend), 742 w (C-S stretch)  $\text{cm}^{-1}$ . HRMS (ESI<sup>+</sup>):  $m/z$ : calcd for  $\text{C}_{11}\text{H}_{19}\text{S}_5$  ( $[\text{M} + \text{H}]^+$ ) 311.0090; found 311.0084.

**2,3-Bis(2-cyanoethylthio)-6,7-bis(butylthio)tetrathiafulvalene (5):** A mixture of 4,5-bis(2-cyanoethylthio)-1,3-dithiol-2-one (**4**, Sigma-Aldrich) (1 g, 3.47 mmol) and compound **3** (1.3 g, 4.16 mmol) was dried by successive evaporation of dry ACN (×3) followed by storage in a desiccator. The residue was dissolved in trimethyl phosphite (40 mL) and the solution heated gradually to reach 110 °C under argon atmosphere. The solution was stirred for 2 h at 110 °C and subsequently allowed to cool RT. The mixture was concentrated to dryness and the residue was treated with toluene and evaporated (×4) in order to eliminate the excess of trimethyl phosphite. The obtained residue was purified by flash chromatography on silica gel (DCM/hexane 8:2 to pure DCM) affording the desired compound **5** as a yellow solid (1.22 g, 64% yield). TLC in DCM/hexane 10:0.1,  $R_f = 0.38$ .  $^1\text{H}$ -NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 3.07 (t,  $J = 7.2$  Hz, 4H, =C-SCH<sub>2</sub>-CH<sub>2</sub>-CN), 2.81 (t,  $J = 7.2$  Hz, 4H, =C-SCH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 2.72 (t,  $J = 7.2$  Hz, 4H, =C-SCH<sub>2</sub>-CH<sub>2</sub>-CN), 1.64–1.56 (m, 4H, C-SCH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 1.47–1.38 (m, 4H, C-SCH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 0.91 (t,  $J = 7.3$  Hz, 6H, CH<sub>3</sub>).  $^{13}\text{C}$ -NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 127.95 and 127.82 (S-C-S), 117.40 (CN), 114.75 (C=C), 106.38 (C=C), 36.02 (S-CH<sub>2</sub>-), 31.70 (S-CH<sub>2</sub>-CH<sub>2</sub>-),

31.23 (S-CH<sub>2</sub>-CH<sub>2</sub>-CN), 21.60 (S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-), 18.89 (S-CH<sub>2</sub>-CH<sub>2</sub>-CN), 13.57 (-CH<sub>3</sub>). IR (solid):  $\nu$  = 2954 m, 2924 m and 2869 m (C-H stretch), 2249 w (C≡N stretch), 1457 w (H-C-H bend), 1415 m-s (H-C-H bend), 889 s (H-C-S bend), 771 s (C-S stretch) cm<sup>-1</sup>. HRMS (ESI<sup>+</sup>): *m/z*: calcd for C<sub>20</sub>H<sub>27</sub>N<sub>2</sub>S<sub>8</sub> ([M + H]<sup>+</sup>) 550.9940; found 550.9956.

2-(2-Cyanoethylthio)-3,6,7-tris(butylthio)tetrathiafulvalene (**6**): A solution of CsOH·H<sub>2</sub>O (365 mg, 2.12 mmol) in dry methanol (MeOH) (15 mL) was added dropwise over a period of 30 min to a stirred solution of TTF derivative **5** (1.13 g, 2.07 mmol) in tetrahydrofuran (THF) (40 mL). The resulting orange solution was stirred for 30 min at RT. To this solution, 1-bromobutane (**2**) (0.44 mL, 4.14 mmol) dissolved in THF (15 mL) was added and the resulting mixture was stirred overnight (o.n.) at RT under nitrogen atmosphere. The mixture was concentrated to dryness and the resulting oil was purified by flash chromatography on silica gel (DCM/hexane 1:1→2:1) giving the desired compound **6** as an orange solid (1.09 g, 89% yield). TLC in DCM/hexane 2:1, *R<sub>f</sub>* = 0.45. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm): 3.01 (t, *J* = 7.2 Hz, 2H, SCH<sub>2</sub>-CH<sub>2</sub>-CN), 2.87–2.78 (m, 6H, SCH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 2.68 (t, *J* = 7.2 Hz, 2H, SCH<sub>2</sub>-CH<sub>2</sub>-CN), 1.65–1.57 (m, 6H, SCH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 1.47–1.38 (m, 6H, SCH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 0.93–0.89 (m, 9H, CH<sub>3</sub>). <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm): 133.71, 127.92, 127.61 and 121.83 (-CH<sub>2</sub>-S-C-S-), 117.53 (CN), 112.36 and 108.27 (S<sub>2</sub>C=CS<sub>2</sub>), 36.02 and 35.98 (S-CH<sub>2</sub>-), 31.77 and 31.73 (S-CH<sub>2</sub>-CH<sub>2</sub>-), 31.24 (S-CH<sub>2</sub>-CH<sub>2</sub>-CN), 21.62 (S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-), 18.69 (S-CH<sub>2</sub>-CH<sub>2</sub>-CN), 13.57 and 13.53 (-CH<sub>3</sub>). IR (solid):  $\nu$  = 2954 s, 2925 s and 2870 m (C-H stretch), 2250 w (C≡N stretch), 1463 m (H-C-H bend), 1412 m (C-H stretch), 890 s (H-C-S bend), 772 m-s and 742 m-s (C-S stretch) cm<sup>-1</sup>. HRMS (ESI<sup>+</sup>): *m/z*: calcd for C<sub>21</sub>H<sub>31</sub>NS<sub>8</sub> 554.0300; found 554.0288.

3-[[4',5,5'-Tris(butylsulfanyl)-2,2'-bi-1,3-dithiol-4-yl]sulfanyl]propionic acid (**8**): (2-Cyanoethyl)sulfanyl TTF derivative **6** (1.01 g, 1.82 mmol) was dissolved in anhydrous THF (40 mL) and a solution of CsOH·H<sub>2</sub>O (0.32 g, 1.9 mmol) in anhydrous MeOH (15 mL) was added. After stirring the reaction mixture at RT for 30 min, a suspension of 3-bromopropionic acid (**7**, Sigma-Aldrich) (0.56 g, 3.6 mmol) and freshly annealed K<sub>2</sub>CO<sub>3</sub> (1.39 g, 10 mmol) in anhydrous MeOH (15 mL) was added. The reaction mixture was stirred at RT for 3 h and concentrated to dryness. The obtained residue was dissolved in Et<sub>2</sub>O (230 mL) and 1M HCl aqueous solution (150 mL) was added slowly. The organic phase was washed with water (100 mL) and brine (100 mL), dried over anhydrous MgSO<sub>4</sub> and concentrated to dryness under reduced pressure. The obtained residue was purified by chromatography on silica gel (DCM/MeOH 100:0→90:10) affording the desired compound **8** as an orange solid (0.95 g, 91% yield). TLC in DCM/MeOH 10:0.5, *R<sub>f</sub>* = 0.50. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm): 3.03 (t, *J* = 7.2 Hz, 2H, SCH<sub>2</sub>-CH<sub>2</sub>-COOH), 2.83–2.79 (m, 6H, SCH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 2.72 (t, *J* = 7.2 Hz, 2H, SCH<sub>2</sub>-CH<sub>2</sub>-COOH), 1.63–1.56 (m, 6H, C-SCH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 1.47–1.37 (m, 6H, C-SCH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 0.91 (t, *J* = 7.2 Hz, 9H, CH<sub>3</sub>). <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm): 176.78 (COOH), 131.10, 127.87, 127.61 and 124.70 (-CH<sub>2</sub>-S-C-S-), 111.17 and 109.22 (S<sub>2</sub>C=CS<sub>2</sub>), 35.99 and 36.96 (S-CH<sub>2</sub>-), 34.41 (SCH<sub>2</sub>-CH<sub>2</sub>-COOH), 31.74 (S-CH<sub>2</sub>-CH<sub>2</sub>-), 30.48 (SCH<sub>2</sub>-CH<sub>2</sub>-COOH), 21.63 (S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-), 13.57 and 13.54 (-CH<sub>3</sub>). IR (solid):  $\nu$  = 2956 m, 2912 m and 2854 m (C-H stretch), 1704 s (C=O stretch), 1465 m (H-C-H bend), 1438 m-s (O-H bend), 1294 m (C-O stretch), 947 m, broad (O-H bend), 887 m (H-C-S bend), 788 m-s, 772 m-s and 740 m-s (C-S stretch) cm<sup>-1</sup>. HRMS (ESI<sup>-</sup>): *m/z*: calcd. for C<sub>21</sub>H<sub>31</sub>O<sub>2</sub>S<sub>8</sub> 571.0095; found 571.0093.

*N*-[*N'*-(*Tert*-butoxycarbonyl)glycylglycylglycyl]-L-threoninol (**13**): The *N*-protected tripeptide Boc-(Gly)<sub>3</sub>-OH (**9**, Bachem) (1.05 g, 3.54 mmol) along with 4-nitrophenol (**10**, Fluka) (0.59 g, 4.25 mmol) were dissolved in anhydrous pyridine (15 mL) under argon and the solution was cooled at 0 °C. Then, 1,3-dicyclohexylcarbodiimide (DCC) (0.88 g, 4.25 mmol) was added under argon and the reaction was stirred at 0 °C for 30 min and then at RT overnight. After this time, the precipitate was filtered out and the solvent was removed under vacuum. The obtained residue was dissolved in toluene and concentrated to dryness under reduced pressure (×3). The resulting 4-nitrophenyl ester **11** was used in the next step without further purification.

The activated acid and (2*R*, 3*R*)-2-aminobutane-1,3-diol (**12**, L-threoninol, Sigma-Aldrich) (0.41 g, 3.89 mmol) were dissolved in anhydrous DMF (15 mL) under argon and stirred at RT for 3 h. After this time, the solvent was removed under reduced pressure and the residue was dissolved in toluene and concentrated to dryness ( $\times 3$ ). The obtained residue was purified by flash chromatography on silica gel (crude adsorbed onto the silica, DCM/MeOH 100:5 $\rightarrow$ 100:15) affording the desired compound **13** as a white solid (1.13 g, 87%). TLC in DCM/MeOH 100:10,  $R_f = 0.35$ .  $^1\text{H-NMR}$  (400 MHz, DMSO- $d_6$ )  $\delta$  (ppm): 8.07–8.01 (m, 2H, NH), 7.25 (d,  $J = 8.0$  Hz, 1H, NH), 6.95 (t,  $J = 6.0$  Hz, 1H, NH), 4.52–4.50 (br, 2H, OH), 3.85–3.83 (m, 1H, OH-CH-CH<sub>3</sub>), 3.71–3.68 (m, 4H, NH-CH<sub>2</sub>-CO), 3.62–3.56 (m, 2H, NH-CH-CH<sub>2</sub>), 3.56 (d,  $J = 8.0$  Hz, 2H, NH-CH<sub>2</sub>-CO), 3.44–3.37 (m, 1H, OH-CH<sub>(A)</sub>H<sub>(B)</sub>-CH), 3.31–3.26 (m, 1H, OH-CH<sub>(A)</sub>H<sub>(B)</sub>-CH), 1.35 (s, 9H, (CH<sub>3</sub>)<sub>3</sub>-C), 0.95 (d,  $J = 6.4$  Hz, 3H, CH<sub>3</sub>-CH-CH).  $^{13}\text{C-NMR}$  (100 MHz, DMSO- $d_6$ )  $\delta$  (ppm): 170.58 (NH-CO), 169.67 (NH-CO), 169.37 (NH-CO), 156.50 (O-CO-NH), 78.83 (CH<sub>3</sub>-C), 64.88 (CH<sub>3</sub>-CH), 61.12 (OH-CH<sub>2</sub>-CH), 56.43 (NH-CH-CH), 43.99 (NH-CH<sub>2</sub>-CO), 42.77 (NH-CH<sub>2</sub>-CO), 42.70 (NH-CH<sub>2</sub>-CO), 28.87 (CH<sub>3</sub>-C), 20.73 (CH<sub>3</sub>-CH). IR (solid):  $\nu = 3428$  w, broad, 3294 m, broad and 3094 w, broad (N-H and O-H stretch), 2963 w and 2929 w (C-H stretch), 1683 m and 1632 s (C=O stretch), 1525 s (N-H bending), 1481 w–1394 w (C-H bend), 1279 m, 1244 s, 1156 m, 1123 w, 1102 w, 1076 m, 1048 m and 1025 m (C-O stretch or C-N stretch), 947 m and 857 m (C-H bend), 654 m, broad (N-H out of plane bend). HRMS (ESI<sup>+</sup>):  $m/z$ : calcd. for C<sub>15</sub>H<sub>29</sub>N<sub>4</sub>O<sub>7</sub> ([M + H]<sup>+</sup>) 377.2036 found 377.2031;  $m/z$ : calcd. for C<sub>15</sub>H<sub>28</sub>N<sub>4</sub>NaO<sub>7</sub> ([M + Na]<sup>+</sup>) 399.1856 found 399.1856.

*L*-Threoninol-(Gly)<sub>3</sub>-TTF derivative (**14**): Compound **13** (0.87 g, 2.32 mmol) was dissolved in DCM (10 mL) and TFA (1 mL) was added slowly at RT. Stirring was continued until TLC showed no remaining starting material (ca. 1–1.5 h) and the mixture was concentrated to dryness under reduced pressure. The obtained residue was dissolved in toluene and concentrated to dryness under reduced pressure. This process was repeated three times to remove traces of TFA to yield the TFA salt of the deprotected amino compound (colorless oil). This crude TFA salt was dried in vacuo and used in the next step without purification.

{[4',5,5'-Tris(butylsulfanyl)-2,2'-bis-1,3-dithiol-4-yl]sulfanyl}propanoic acid (**8**, 0.95 g, 1.66 mmol) along with *N*-hydroxysuccinimide (NHS) (Sigma-Aldrich) (0.21 g, 1.83 mmol) were dissolved in anhydrous THF (40 mL) under argon and the solution was cooled at 0 °C. Then, DCC (0.38 g, 1.83 mmol) was added under argon and the reaction was stirred at 0 °C for 30 min and at RT overnight. The precipitate was filtered out and the solvent was removed under reduced pressure. The resulting *N*-hydroxysuccinimide ester was used in the next step without further purification. The active ester was dissolved in anhydrous DMF (30 mL). The TFA salt of the deprotected amino compound was dissolved in DMF (30 mL) and *N,N*-diisopropylethylamine (DIEA) (405  $\mu\text{L}$ , 2.32 mmol) was added. Then, the solution of the active ester was added under argon and the mixture was allowed to react at RT for 30 min. The solvent was removed under reduced pressure and the residue was dissolved in toluene and concentrated to dryness ( $\times 3$ ). The crude compound was washed with hexane (5  $\times$  70 mL) and with diethyl ether (5  $\times$  70 mL). The residue was dissolved in DCM (300 mL) and the organic phase was washed with 5% aqueous NaHCO<sub>3</sub> (2  $\times$  75 mL) and saturated aqueous NaCl (75 mL), dried over anhydrous MgSO<sub>4</sub>, and concentrated to dryness under reduced pressure. The crude residue was purified by chromatography on silica gel (DCM/MeOH 100:0.5 $\rightarrow$ 90:10) obtaining a reddish solid (868 mg, 62%). TLC (DCM/MeOH 100:1)  $R_f = 0.10$ .  $^1\text{H-NMR}$  (400 MHz, CD<sub>3</sub>OD)  $\delta$  (ppm): 7.83–7.52 (NH zone), 3.98–3.95 (m, 1H, OH-CH-CH<sub>3</sub>), 3.91–3.69 (m, 6H, NH-CH<sub>2</sub>-CO), 3.82–3.79 (m, 1H, NH-CH-CH<sub>2</sub>), 3.66–3.62 (m, 1H, OH-CH<sub>(A)</sub>H<sub>(B)</sub>-CH), 3.60–3.55 (m, 1H, OH-CH<sub>(A)</sub>H<sub>(B)</sub>-CH), 3.09 (t,  $J = 7.2$  Hz, 2H, SCH<sub>2</sub>-CH<sub>2</sub>-CONH), 2.87–2.81 (m, 6H, SCH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 2.64 (t,  $J = 7.2$  Hz, 2H, SCH<sub>2</sub>-CH<sub>2</sub>-CONH), 1.64–1.56 (m, 6H, C-SCH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 1.49–1.40 (m, 6H, C-SCH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 1.13 (d,  $J = 6.4$  Hz, 3H, CH<sub>3</sub>-CH-CH), 0.92 (t, 9H,  $J = 7.2$  Hz, SCH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>).  $^{13}\text{C-NMR}$  (100 MHz, CD<sub>3</sub>OD)  $\delta$  (ppm): 172.71, 171.21, 170.82 and 170.44 (NH-CO), 129.50, 127.78, 127.10 and 126.18 (-CH<sub>2</sub>-S-C-S-), 110.15 and 109.49 (S<sub>2</sub>C=CS<sub>2</sub>), 65.80 (CH<sub>3</sub>-CH), 61.35 (OH-CH<sub>2</sub>-CH), 56.34 (NH-CH-CH), 42.49, 42.43 and 42.17 (NH-CH<sub>2</sub>-CO), 35.52, 35.45, 35.40 and 35.38 (S-CH<sub>2</sub>- and SCH<sub>2</sub>-CH<sub>2</sub>-CONH), 31.59, 31.56 and 31.55

(S-CH<sub>2</sub>-CH<sub>2</sub>), 31.02 (SCH<sub>2</sub>-CH<sub>2</sub>-CONH), 21.16 and 21.14 (S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-), 18.92 (CH<sub>3</sub>-CH), 12.54 and 12.51 (S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>). IR (solid):  $\nu$  = 3280 m, broad and 3086 w, broad (O-H and N-H stretch), 2955 m-w, 2926 m-w and 2869 w (C-H stretch), 1642 s (C=O stretch), 1553 m-s (N-H bend), 1456 m-w, 1417 m-w and 1376 m-w (C-H bend or O-H bend or H-C-H bend), 1249 m, 1099 m and 1028 m (C-O stretch and N-H stretch), 886 w (H-C-S bend or C-H bend), 771 m (C-S stretch), 684 m, broad (N-H out of plane). HRMS (ESI<sup>+</sup>):  $m/z$ : calcd for C<sub>31</sub>H<sub>52</sub>N<sub>4</sub>O<sub>2</sub>S<sub>8</sub> ([M + H]<sup>+</sup>) 831.1574 found 831.1613;  $m/z$ : calcd for C<sub>31</sub>H<sub>52</sub>N<sub>4</sub>NaO<sub>2</sub>S<sub>8</sub> ([M + Na]<sup>+</sup>) 853.1394 found 853.1416.

### 2.3. Preparation and Characterization of Gel Materials

Anhydrous solvents used for gelation tests were purchased from commercial suppliers. Gelation tests were carried out in screw-capped glass vials (4 cm length × 1 cm diameter) having a specific amount of the TTF derivative **14** and the desired solvent (1 mL). The mixture was heated with a standard heat gun until complete dissolution. Then, the clear solution was allowed to cool down to RT. The material was initially classified as gel if it did not flow by turning the vial upside-down. The viscoelastic nature was further confirmed by rheology of a model system.

Critical gelation concentration (CGC) corresponds to the minimum gelator concentration required for gelation. The values were obtained by continuously adding aliquots of solvent (0.02–0.1 mL) into vials having the TTF derivative **14** and performing either the heating or cooling until no gelation was observed. The waiting time used to define the state of the material was ~12 h. Concentrations above 200 g/L were not tested.

Thermal *gel-to-sol* transition temperature ( $T_{\text{gel}}$ ) values were determined using a calibrated thermoblock at 1 °C/5 min (Figure S1). The temperature at which the bulk gel started to break was defined as  $T_{\text{gel}}$ . Due to potential effects of thermal history and hysteresis [75], the apparatus has been previously calibrated with samples measured by differential scanning calorimetry (DSC) [76].

Rheological measurements were performed with an AR 2000 Advanced rheometer (TA Instruments, New Castle, DE, USA). Conditions: 1000 mm gap, torque =  $5 \times 10^{-4}$  N/m, 25 °C, plain-plate geometry (20 mm, stainless steel), sample volume = 2 mL. Dynamic strain sweep (DSS) measurements were performed between 0.01 and 100% strain. For dynamic frequency sweep (DFS) measurements, frequency was varied from 0.1 to 10 Hz at 0.1% strain. Finally, dynamic time sweep (DTS) measurements were carried out within the linear regime at 0.1% strain and 1 Hz frequency.

FT-IR spectra were obtained at RT with a Cary 630 FTIR spectrometer (Agilent Technologies, Santa Clara, CA, USA) equipped with an attenuated total reflection (ATR) accessory (32 scans, spectral resolution = 8 cm<sup>-1</sup>).

Field emission scanning electron microscopy (FE-SEM) of xerogels was carried out with a Zeiss Merlin, Field Emission Scanning Electron Microscope (accelerating voltage = 10 kV). Xerogels, prepared by freeze-drying the gels, were placed on top of a tin plate and shielded with Pt (40 mA, 30–60 s; film thickness = 5–10 nm). Images were by Servicio General de Apoyo a la Investigación-SAI (University of Zaragoza).

Images under polarized light were obtained using a Wild Makroskop M420 optical microscope equipped with a Canon Power shot A640 camera.

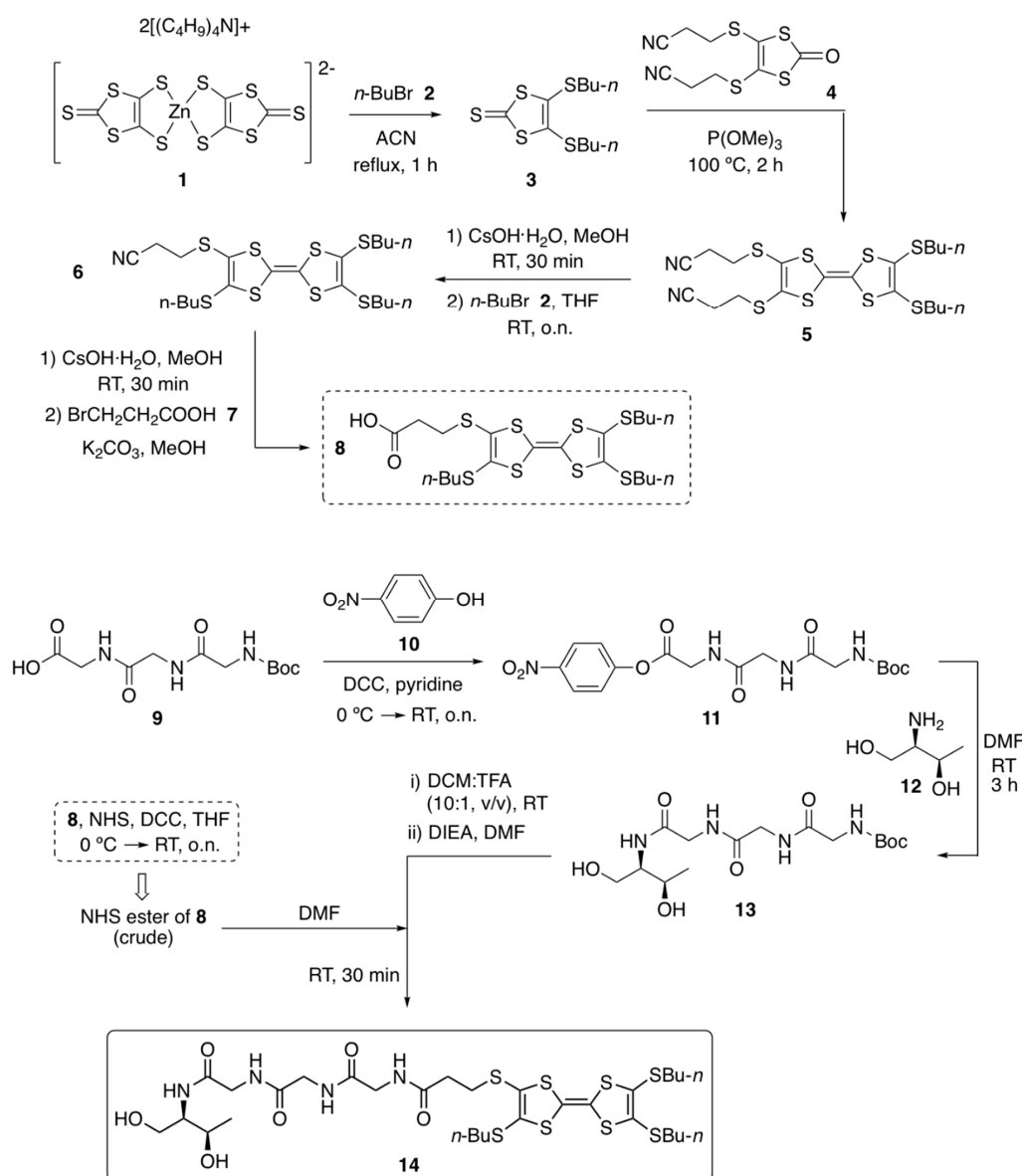
UV-vis spectra were obtained with an Ocean Optics, Flame spectrometer with a DH-2000-BAL light source or on a Varian Cary 50 UV spectrophotometer. Quartz-glass cuvettes of 0.5 cm thickness were used.

Iodine doping was achieved by exposing the xerogel to I<sub>2</sub> vapor for *circa* 120 min in a sealed glass beaker with I<sub>2</sub> crystals.

### 3. Results and Discussion

#### 3.1. Synthesis of TTF–Triglycyl Derivative

The synthesis of the TTF–triglycyl derivative **14** was achieved by assembling the L-threoninol, and triglycine units with the TTF carboxylic derivative **8** (Scheme 1). First, compound **8** was prepared following the procedure described in the literature [10] with slight modifications. The first step consisted in the synthesis of 4,5-bis-(butylthio)-1,3-dithiole-2-thione (**3**) by alkylation of the zinc complex of bis(1,3-dithiole-2-thione-4,5-dithiolate) tetrabutylammonium salt (**1**) with 1-bromobutane (**2**) as described for similar compounds by Simonsen and co-workers [77]. Phosphite-induced coupling of compounds **3** and **4** [77] gave the symmetric TTF derivative **5** in good yield (64%). Selective sequential deprotection of one 2-cyanoethyl group followed by alkylation with 1-bromobutane (**2**) gave the TTF derivative **6** in very good yield (89%). A second round of sequential deprotection of the second 2-cyanoethyl group followed by alkylation with 3-bromopropionic acid (**7**) [78] gave the desired TTF carboxylic derivative **8** in 91% yield.



Scheme 1. Synthesis of TTF–triglycyl derivative **14**.



Then Boc-protected triglycine **9** was activated with DCC and 4-nitrophenol (**10**). Then, the obtained nitrophenyl active ester **11** was reacted with L-threoninol (**12**) yielding the *t*-butoxycarbonyl (Boc)-protected triglycine-L-threoninol derivative **13**. This compound was treated with a TFA solution in DCM to remove the Boc-amino protecting group yielding the corresponding L-threoninol-triglycine trifluoroacetate salt that was subsequently used without purification for the coupling with compound **8** in the presence of Hünig's base. This reaction was done via classical activation of the carboxyl group with DCC and NHS, affording the desired TTF-based gelator **14** in 24% (isolated yield).

### 3.2. Gelation Properties

The gelation ability of TTF-triglycyl derivative **14** was studied for a number of anhydrous solvents of different nature (i.e., apolar, polar aprotic, polar protic) using the standard heating-cooling treatment. Materials that did flow upon inversion of the vial upside-down were initially classified as gels. One of the gels was used later as a model example to demonstrate its viscoelastic nature by oscillatory rheological measurements (see below). Compound **14** was found to be soluble in methanol, ethanol, isopropanol, dimethylformamide, dimethyl sulfoxide, and tetrahydrofuran, whereas it resulted insoluble in diethyl ether, acetonitrile, *n*-hexane, and cyclohexane. In contrast, opaque gels were obtained in 10 solvents at concentrations varying between  $5 \pm 1$  and  $50 \pm 10$  g/L (Table 1).

**Table 1.** Gelation ability of TTF-triglycyl derivative **14**, critical gelation concentration (CGC), gelation time, and *gel-to-sol* transition temperature ( $T_{gel}$ ).<sup>1</sup>

Solvent <sup>2</sup>	CGC (g/L)	Gelation Time (min)	$T_{gel}$ (°C)
Acetone	$40 \pm 10$	$60 \pm 10$	$36 \pm 3$ <sup>4</sup>
Benzene	$6 \pm 1$	$78 \pm 5$	$37 \pm 2$
Benzonitrile	$35 \pm 10$	$55 \pm 5$	$42 \pm 2$
Chlorobenzene	$9 \pm 1$	$78 \pm 5$	$38 \pm 2$
Chloroform	$18 \pm 2$	$10 \pm 2$	$50 \pm 3$
Dichloromethane	$29 \pm 2$	$10 \pm 2$	$49 \pm 2$
1,4-Dioxane	$50 \pm 10$	$60 \pm 10$	$51 \pm 3$
Ethyl acetate	$5 \pm 1$	$10 \pm 2$	$47 \pm 2$
Toluene	$9 \pm 1$	$90 \pm 10$	$41 \pm 2$
Xylene <sup>3</sup>	$8 \pm 1$	$90 \pm 10$	$46 \pm 2$

<sup>1</sup> Gels were obtained upon a heating-cooling cycle (see Experimental Section). The use of anhydrous solvents was found to be very critical to obtain the lowest possible critical gelation concentration (CGC) values. In general, the use non-anhydrous solvents either inhibited the gelation or raised the CGC up to 3–4 times in comparison to anhydrous solvents. Error values were calculated from three randomized experiments. <sup>2</sup> Solvent volume = 1 mL.

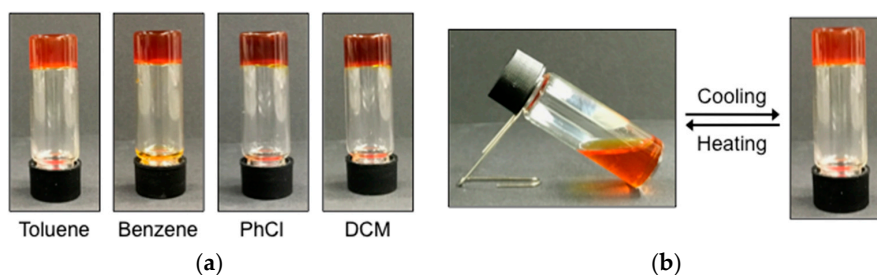
<sup>3</sup> Xylene was used as mixture of isomers. <sup>4</sup> Partial gel formation was observed. In this case,  $T_{gel}$  value corresponds to the isolated gelled portion.

These gels remained stable for at least two months when stored in sealed vials at RT. Their color varied from bright orange to dark orange-brown depending on the solvent (Figure 2a). The supramolecular gels showed typical thermoreversibility (Figure 2b), and their opaque appearance suggested the presence of aggregates larger than the wavelength of visible light ( $\lambda = 380\text{--}780$  nm), which was in agreement with electron microscopy (see below). Further investigations were carried out using only some selected gels as representative materials.

As mentioned in the introduction, it is well established that the redox TTF core in this type of LMW also provides both  $S \cdots S$  and  $\pi \cdots \pi$  interactions during molecular aggregation, whereas the peptide chain drives molecular aggregation via hydrogen bonding [73,74]. Comparison of Fourier transform infrared (FT-IR) spectra of solid TTF derivative **14** with that of the xerogel (i.e., prepared by freeze-drying the corresponding organogel) and **14** in solution (i.e., below the CGC) showed only very small frequency shifts (i.e., red-shift  $\Delta\nu \sim 5$   $\text{cm}^{-1}$ ) for characteristic bands such as C=O stretching ( $\sim 1641$   $\text{cm}^{-1}$ ), N-H amide I (br  $\sim 3280$   $\text{cm}^{-1}$ ), and N-H amide II (bending vibration) ( $\sim 1553$   $\text{cm}^{-1}$ ) (Figure S2). These values suggest that **14** may also be aggregated in the solid state and in solution via



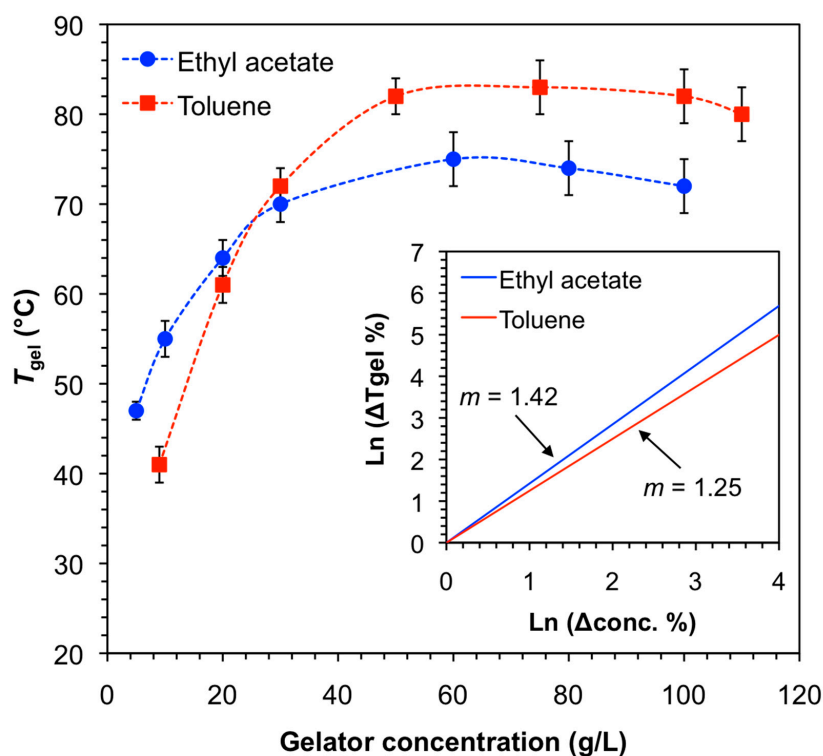
similar intermolecular hydrogen-bonding pattern between amide groups, most likely in the parallel  $\beta$ -sheet conformation [79], as well as van der Waals interactions between the alkyl chains.



**Figure 2.** (a) Photographs of upside-down vials having selected gels made of **14** in toluene, benzene, chlorobenzene (PhCl), and dichloromethane (DCM) at the CGC (see Table 1); (b) illustration of the thermal *sol-gel* transition of the gel made of **14** in toluene at the CGC.

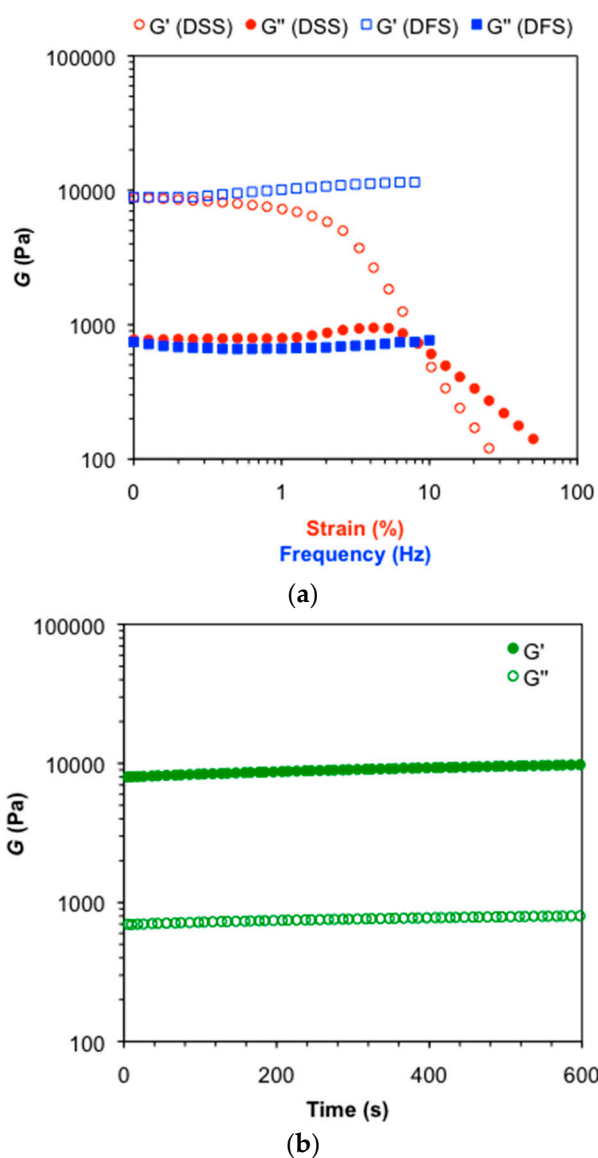
### 3.3. Characterization of Organogels

In general, relatively low *gel-to-sol* transition temperatures ( $T_{gel}$ ) were determined for all gels ranging from 36 to 51 °C ( $\pm 3$ ) (Table 1). As typically observed with most supramolecular gels,  $T_{gel}$  gradually increased with increasing gelator concentration due to the generation of denser networks (Figure 3). Taking the gels prepared in toluene and ethyl acetate as representative systems, a plateau region was reached before the gels collapsed into partial and inhomogeneous gels that constantly lose solvent over time. The increment of the  $T_{gel}$  until the plateau region was  $\sim 39$  °C for toluene and  $\sim 25$  °C for ethyl acetate with respect to the initial values obtained at their CGC. Linear Ln-Ln plot (Figure 3, inset) using the percentage increases of  $T_{gel}$  showed that the percentage increment of  $T_{gel}$  was  $\sim 1.1$ -fold higher for the gel made in ethyl acetate. Further research is necessary to understand the reasons behind such differences.



**Figure 3.** Variation of  $T_{gel}$  with increasing gelator concentration for the gels made in toluene and ethyl acetate. Inset: Normalized Ln-Ln plot of  $T_{gel}$  percentage increases.

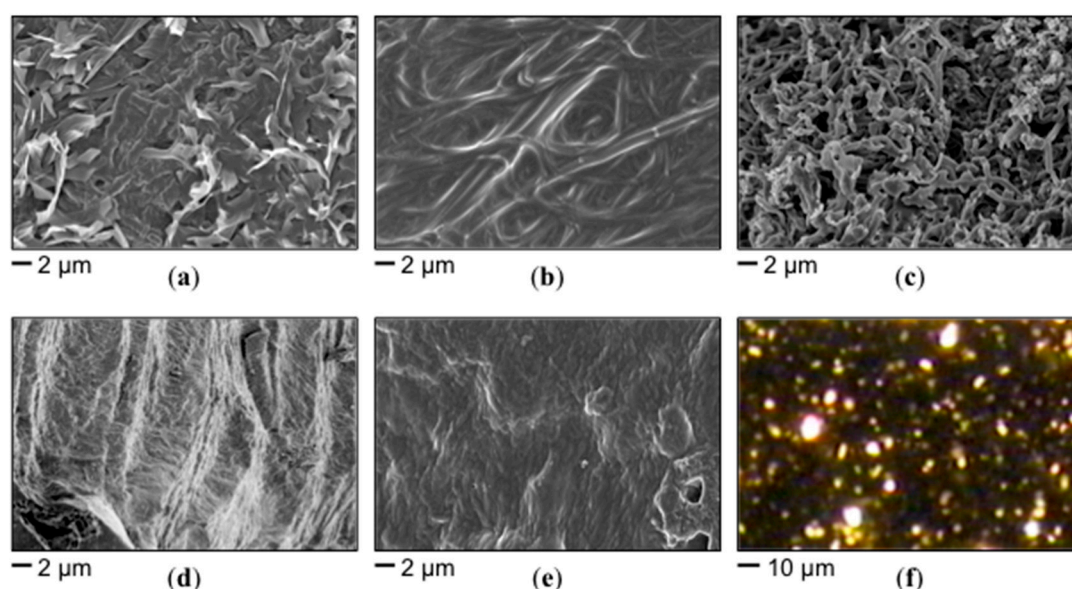
Dynamic rheological experiments (Figure 4) for the model gel made of **14** in toluene confirmed its viscoelastic nature. Within the linear regime established by dynamic frequency sweep (DFS) and dynamic strain sweep (DSS) measurements, the storage modulus  $G'$  was found to be one order of magnitude higher than the loss modulus  $G''$  with low frequency dependency (i.e.,  $G' \approx 8.9 \pm 0.5$  kPa,  $G'' \approx 0.75 \pm 0.1$  kPa) (Figure 3a). Moreover, the gels were brittle in nature, which was confirmed by its destruction at low frequency and  $\sim 7\%$  of strain (Figure 4a). Finally, dynamic time sweep (DTS) measurements at 0.1% strain and 1 Hz frequency confirmed the stability of the gel over time at RT (Figure 4b).



**Figure 4.** (a) Dynamic frequency sweep (DFS), dynamic strain sweep (DSS), and (b) dynamic time sweep (DTS) measurements for the model gel made of **14** in toluene (9 g/L). The  $\tan \delta$  (i.e.,  $G''/G'$ , damping coefficient) was reproducible in random experiments.

Morphological studies of selected organogels were conducted by field emission scanning electron microscopy (FE-SEM) of the corresponding xerogels (Figure 5). In general, fibrillar networks with high aspect ratio were observed for several organogels made of **14**. As observed with other supramolecular self-assembled gels, interactions between solvent–aggregate, solvent–gelator, and gelator–gelator molecules are also regulated by the solvent nature and have a major effect on the microstructure of

the materials. For instance, dense and entangled leaf-like structures were observed for the material obtained in ethyl acetate (Figure 5a), whereas the samples prepared in benzonitrile showed a smooth surface with fibrillar structures of  $\sim 0.5\text{--}1\ \mu\text{m}$  in diameter (Figure 5b). A highly dense and porous network made of fibers up to  $\sim 1\ \mu\text{m}$  in diameter was observed for the gel made in 1,4-dioxane (Figure 5c). Surfaces made of smaller fibers of  $\sim 50\text{--}100\ \text{nm}$  in diameter resembling a mountainous valley were observed for the specimens prepared in toluene (Figure 5d). However, fibrillar-globular structures with diameters between  $\sim 100$  and  $500\ \text{nm}$  were distinguished when the solvent used for the preparation of the gel was dichloromethane (Figure 5e). Although we systematically took pictures of the bulk samples at different magnifications and locations on the grid in order to identify possible artifacts in the microstructures, it should be considered that such artifacts can be produced during the preparation and visualization of the samples and, therefore, the conclusions derived from these images should not be oversold. Furthermore, the typical anisotropic growth in this type of materials, suggesting well-ordered molecular packing, is also evidenced by the observation of birefringent domains under polarized light (Figure 5f), which is important for optical applications [80].

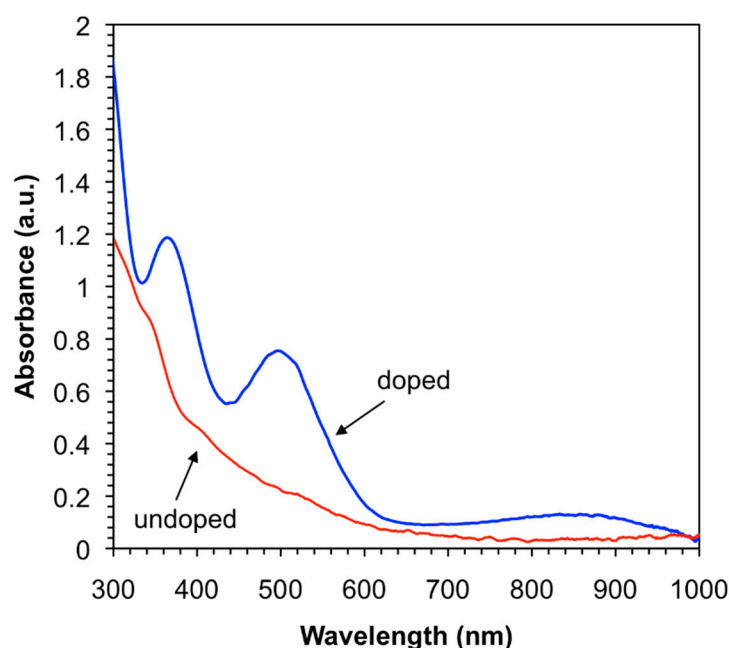


**Figure 5.** Representative FE-SEM photographs of xerogels prepared by freeze-drying the organogels made of **14** in (a) ethyl acetate (5 g/L); (b) benzonitrile (35 g/L); (c) 1,4-dioxane (50 g/L); (d) toluene (9 g/L); (e) dichloromethane (29 g/L); (f) birefringent domains visualized under polarized light for the xerogel derived from the gel made of **14** in dichloromethane (29 g/L).

### 3.4. Effect of Iodine-Doping

Iodine-doping of the fibrous xerogel derived from a model gel made in DCM confirmed the oxidation of the TTF moiety in the gelator preserving the self-assembled network structure. UV-vis-NIR spectrum of the xerogel after exposure to  $\text{I}_2$  vapor showed new absorption bands centered at approximately 360, 500, and 850 nm (Figure 6). The absorption bands at 360 and 500 nm are typically ascribed to intramolecular electronic transitions involving the TTF cation radical, whereas the broad band located at 850 nm indicates the formation of a full CT state between cation radicals of self-stacked TTF units (i.e.,  $\text{TTF}^+\text{I}^-$ ). The increase in the absorbance above 600 nm is in agreement with the formation of a partial mixed-valence CT state between TTF-containing neutral and cation radicals (i.e.,  $(\text{TTF})(\text{I})_n$  with  $n < 1$ ) [58,73]. Unfortunately, the intermolecular hydrogen-bonded network seemed to be altered during the oxidation as shown by visible changes in the intensity and position of the IR absorption bands associated to N-H and C=O stretching vibrations of the amides (Figure S3). Unfortunately, although iodine can create the necessary unfilled states required for conduction,

the instability of the fibrillar network prevented us from obtaining reproducible conductivity values. As observed with other similar gels, preliminary experiments also showed a *gel-to-sol* transition after exposure of the wet gel to iodine vapors.



**Figure 6.** UV-vis-NIR spectra before and after doping with  $I_2$  the xerogel prepared from the organogel made of **14** in DCM (29 g/L).

#### 4. Conclusions

In conclusion, TTF–triglycyl derivative **14** was synthesized and fully characterized by standard techniques. This compound was found to act as a LMW gelator for a series of organic solvents after a heating–cooling cycle. Therefore, the presence of aromatic rings on the peptide linked to the TTF unit is not a requirement for the formation of supramolecular gels using TTF-peptide conjugates. The critical gelation concentrations ranged from ~5 to 50 g/L, and the thermal *gel-to-sol* transition temperatures varied from ~36 to 51 °C. FE-SEM imaging of the xerogels, prepared by freeze-drying the organogels, showed fibrillar microstructures with distinctive features depending on the solvent used to prepare the gels. Moreover, the viscoelastic and brittle nature of a model gel system was also supported by dynamic rheological experiments. Finally, UV-vis-NIR analysis of a model xerogel confirmed the oxidation of the TTF unit upon exposure to  $I_2$  vapor. Nevertheless, FT-IR spectroscopy showed that the oxidation was accompanied with concurrent alteration of the intermolecular hydrogen-bonded network.

**Supplementary Materials:** The following are available online at <http://www.mdpi.com/2076-3417/8/5/671/s1>, NMR spectra, FT-IR spectra, Figure S1: Thermoblock used for  $T_{gel}$  determinations. (A) Front view of the set-up. (B) Top view of the set-up during a typical experiment. The set-up was previously calibrated with literature data for known compounds, Figure S2: FT-IR spectra of solid **14**, **14** dissolved in DCM, and xerogel, i.e., prepared by freeze-drying the organogel made of **14** in DCM (29 g/L), Figure S3: FT-IR spectra of the xerogel, obtained from the corresponding organogel made of **14** in DCM (29 g/L), before and after iodine-doping.

**Author Contributions:** Pérez-Rentero Sónia carried out the synthesis and characterization of all compounds, and observed for the first time the aggregation tendency of the gelator. Häring Marleen, Saldías César, and Díaz Díaz David carried out the rest of the experiments. Díaz Díaz David and Eritja Ramon directed the project. All authors participated in writing the manuscript.

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