Check for updates

OPEN ACCESS

EDITED BY Jan A. Plock, Aarau Cantonal Hospital, Switzerland

REVIEWED BY

Sofija Pejkova, University Clinic for Plastic and Reconstructive Surgery, North Macedonia Tito Brambullo, University of Padua, Italy

*CORRESPONDENCE Martin Kauke-Navarro kauke-navarro.martin@yale.edu Leonard Knoedler leonardknoedler@gmail.com

RECEIVED 11 August 2023 ACCEPTED 05 April 2024 PUBLISHED 23 April 2024

CITATION

Knoedler L, Huelsboemer L, Hollmann K, Alfertshofer M, Herfeld K, Hosseini H, Boroumand S, Stoegner VA, Safi A-F, Perl M, Knoedler S, Pomahac B and Kauke-Navarro M (2024) From standard therapies to monoclonal antibodies and immune checkpoint inhibitors – an update for reconstructive surgeons on common oncological cases. *Front. Immunol.* 15:1276306. doi: 10.3389/fimmu.2024.1276306

COPYRIGHT

© 2024 Knoedler, Huelsboemer, Hollmann, Alfertshofer, Herfeld, Hosseini, Boroumand, Stoegner, Safi, Perl, Knoedler, Pomahac and Kauke-Navarro. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

From standard therapies to monoclonal antibodies and immune checkpoint inhibitors – an update for reconstructive surgeons on common oncological cases

Leonard Knoedler^{1,2*}, Lioba Huelsboemer², Katharina Hollmann^{3,4}, Michael Alfertshofer⁵, Konstantin Herfeld^{6,7}, Helia Hosseini², Sam Boroumand², Viola A. Stoegner^{2,8}, Ali-Farid Safi^{9,10}, Markus Perl^{6,7}, Samuel Knoedler^{1,2}, Bohdan Pomahac² and Martin Kauke-Navarro^{2*}

¹Department of Plastic, Hand, and Reconstructive Surgery, University Hospital Regensburg, Regensburg, Germany, ²Division of Plastic Surgery, Department of Surgery, Yale New Haven Hospital, Yale School of Medicine, New Haven, CT, United States, ³Department of Pathology, Massachusetts General Hospital, Harvard Medical School, Boston, MA, United States, ⁴Faculty of Medicine, University of Wuerzbuerg, Wuerzburg, Germany, ⁹Division of Hand, Plastic and Aesthetic Surgery, Ludwig-Maximilians University Munich, Munich, Germany, ⁶Department of Internal Medicine III (Oncology and Haematology), University Hospital Regensburg, Regensburg, Germany, ⁷Leibniz Institute for Immunotherapy, Regensburg, Germany, ⁸Department of Plastic, Aesthetic, Hand and Reconstructive Surgery, Burn Center, Hannover Medical School, Hannover, Germany, ⁹Craniologicum, Center for Cranio-Maxillo-Facial Surgery, Bern, Switzerland, ¹⁰Faculty of Medicine, University of Bern, Bern, Switzerland

Malignancies represent a persisting worldwide health burden. Tumor treatment is commonly based on surgical and/or non-surgical therapies. In the recent decade, novel non-surgical treatment strategies involving monoclonal antibodies (mAB) and immune checkpoint inhibitors (ICI) have been successfully incorporated into standard treatment algorithms. Such emerging therapy concepts have demonstrated improved complete remission rates and prolonged progression-free survival compared to conventional chemotherapies. However, the in-toto surgical tumor resection followed by reconstructive surgery oftentimes remains the only curative therapy. Breast cancer (BC), skin cancer (SC), head and neck cancer (HNC), and sarcoma amongst other cancer entities commonly require reconstructive surgery to restore form, aesthetics, and functionality. Understanding the basic principles, strengths, and limitations of mAB and ICI as (neo-) adjuvant therapies and treatment alternatives for resectable or unresectable tumors is paramount for optimized surgical therapy planning. Yet, there is a scarcity of studies that condense the current body of literature on mAB and ICI for BC, SC, HNC, and sarcoma. This knowledge gap may result in suboptimal treatment planning, ultimately impairing patient

outcomes. Herein, we aim to summarize the current translational endeavors focusing on mAB and ICI. This line of research may serve as an evidence-based fundament to guide targeted therapy and optimize interdisciplinary anticancer strategies.

KEYWORDS

breast cancer, sarcoma, head and neck cancer, skin cancer, malignant melanoma, monoclonal antibody, immunotherapy, immune checkpoint inhibitors

1 Introduction

Cancer persists as a worldwide healthcare burden currently affecting 18 million US Americans (1). Malignant tumors are the second leading cause of death in the US population accounting for 609,360 deaths in 2022 (2). Annual cancer therapy costs amount to more than \$200 billion. About one-third of cancer patients report symptoms of reactive depression representing an additional burden beyond the tumor disease (3).

In most solid malignancies, advanced or high-risk cancer commonly warrants a joint approach including local treatment and systematic therapy (4-9). Over the past decades, chemotherapeutics and radiotherapy have been considered the standard (neo-)adjuvant therapy strategies. However, recent scientific advances have proposed novel treatment protocols and broadened the therapeutic arsenal of healthcare providers (10).

The advent of monoclonal antibodies (mAB) (e.g., the antihuman epidermal growth factor receptor 2 (HER-2)/neu antibody trastuzumab) and immune checkpoint inhibitors (ICI) (e.g., programmed cell death protein 1 (PD-1), programmed cell death protein ligand 1 (PD-L1), and cytotoxic T-lymphocyte antigen 4 (CTLA-4) inhibitors) has heralded a paradigm shift in oncology (11, 12). ICI target the immune system evasion mechanism of tumor cells based on self-tolerance-inducing proteins on the cell surface. Common target structures include PD-1 and its ligand PD-L1, both inhibitors of antitumoral T cells (13, 14). The combined positive score (CPS) is used to judge the level of PD-L1 expression, by evaluating the proportion of potential PD-L1 expression, encompassing both tumor and immune cells, relative to the overall count of living tumor cells (15). The clinical effects of mAB (as targeted therapy) center on the stimulation of different innate immune effector processes, direct antibody-mediated toxicity, and complement activation (16). Specific cell surface antigens and receptors, like, epidermal growth factor receptor (EGFR), and vascular endothelial growth factor receptor (VEGFR), represent established targets for mAB (17). The therapeutic potency and clinical efficacy have led to increased rates of complete remissions and prolonged progression-free survival (PFS) even in advanced neoplasms (18). However, the administration of ICI and mAB can induce various side effects. For

instance, ICI have been associated with pneumonitis, hepatitis, and myocarditis, while mAB have been implicated with cytokine release syndrome, pneumonitis, cytopenia, or cardiac failure (19–24).

Knowledge of the efficacy, safety, and drug interactions of these emerging treatments is pivotal to leveraging systemic therapies with local surgical strategies (25). There is a scarcity of research work condensing the scientific literature on mAB and ICI for the surgical readership. This knowledge gap leads to untapped therapy potential impairing patient outcomes and survival rates.

Additionally, current cancer guidelines recommend a close interlocking of cancer resection and reconstructive surgery to restore the functionality and aesthetics of the affected body region. A mounting body of evidence has underscored the clinical impact of reconstructive surgery to support patient rehabilitation and improve quality of life (26–29).

Therefore, we aim to summarize the current body of evidence and clinical trials on mAB, ICI, and other immunotherapies for the most common cancer entities treated by reconstructive surgeons (i.e., head and neck cancer (HNC), breast cancer (BC), skin cancer (SC), and sarcoma). This line of research may help reconstructive surgeons develop a more comprehensive understanding of cancer therapies and optimize the perioperative workflow.

2 Head and neck cancer – basic information

HNC describes a heterogeneous and complex group of malignancies, categorized based on their anatomical location in the oral cavity, oropharynx, nasopharynx, hypopharynx, and larynx. The majority of HNC (90%) stems from the epithelial lining and is summarized as HNSCC (30–32). Given the majority of HNSCC within HNC, this review focuses on HNSCC and its current neoadjuvant and adjuvant immunotherapeutic modalities. Worldwide, HNSCC is accounting for more than 700,000 new cases and 300,000 deaths per year, representing the sixth most common cancer (33, 34). The overall incidence of HNSCC is proposed to increase by 30% unitl 2030, both in developed and developing countries (32, 35). Significant epidemiological variations in the incidence of HNSCC include tobacco and alcohol consumption,

environmental pollutants, and viral infections, namely human papilloma virus (HPV) and Epstein-barr virus (EBV) infections (31, 34). A substitute biomarker for HPV infection is protein p16, which is upregulated by HPV virus proteins (36–38). The American Cancer Society estimated 54,540 new oral cavity and pharynx cancer cases for both sexes in the US of which about 75% occur in male patients (39). The gender dysbalance might be due to a higher prevalence of behavioural risk factors (e.g., nicotine or alcohol abuse) in males (40). Besides these, genetic syndromes (e.g., Fanconi anemia) may contribute to HNSCC (41).

2.1 Standard therapy for head and neck cancer

The neoadjuvant therapy typically involves chemotherapy, with a preference for platinum-based compounds like cisplatin, often used in conjunction with taxanes or 5-fluorouracil (5-FU), and might succeeded by radiotherapy or chemoradiotherapy (which may warrant further surgical management due to side-effects) to further halt the tumor's progression. For locally advanced HNSCC, the primary approach is commonly surgery followed by adjuvant radiation therapy or definitive simultaneous chemoradiotherapy, with surgery (in an organ-preservation approach) kept as a salvage option (42). As delineated in the National Comprehensive Cancer Network (NCCN) Guidelines, Version 2.2020, salvage surgery can also present a curative opportunity for patients enduring isolated resectable recurrences. Treatment decisions are contingent upon several factors, such as the patient's prior therapy, location, and the extent of recurrence. The treatment modality for unresectable diseases generally mirrors the strategy deployed for metastatic HNSCC. Yet, patients with unresectable HNSCC may still undergo definitive radiotherapy. For metastatic or widespread recurrent HNSCC, systemic therapy is the mainstay of treatment. This typically involves the use of chemotherapy, with platin derivates, taxanes, and 5-FU among others, utilized individually or in various combinations. In such settings, targeted therapies like the EGFR-inhibitor cetuximab have also proven efficacious (43).

In summary, the standard treatment approach for HNSCC, either applied in a neoadjuvant or adjuvant setting, currently consists of surgery and (radio-)chemotherapy but is associated with considerable morbidity and unfavorable prognoses yielding poor five-year survival rates. A landmark Indian multi-institution study reported a five-year cumulative survival (FCS) of 68% for surgical treatment with radiation for oral cavity cancer and 60% for locally advanced stages. Chemoradiation of irresectable cancer compared to radiation alone showed an improvement of 15% in survival rate for oro- and hypopharyngeal cancers with 40% FCS (44). Resistance to radio- and chemotherapy and relapse have been described as the main factors contributing to low survival rates. Additionally, the genetic variety within HNSCC has impeded the process of identifying specific targets and precision therapies. Therefore, gaining a deeper understanding of the pathophysiology of HNSCC is crucial for the advancement of therapy (30). Recent clinical trials indicate that especially mAB immunotherapy can potentially transition paradigms in the treatment of HNSCC (45).

2.2 The role of monoclonal antibodies in head and neck cancer therapy

Early favor was assigned to cetuximab, an EGFR-inhibiting mAB. EGFR is a receptor tyrosine kinase upregulated in many malignancies. Its overexpression or overactivation sets off downstream pro-oncogenic pathways, promoting cell proliferation and inhibiting apoptosis (45, 46).

Initially examined in combination with radiotherapy, additional cetuximab showed longer locoregional control (24.4 months) compared to radiation alone (14.9 months). Furthermore, cetuximab and radiotherapy achieved significantly longer overall survival (49.0 months) than radiotherapy alone (29.3 months). Side effects did not differ significantly in both treatment regimens, with the exception of acneiform rash and infusion reactions (47).

In a subsequent phase III trial, including 891 patients, of concurrent accelerated radiation plus cisplatin in combination with cetuximab or alone for stage III to IV HNSCC, the addition of cetuximab led to more interruptions in radiation therapy and more grade III to IV radiation mucositis but did not improve three-year PFS or overall survival. Notably, patients with $p16^+$ oropharyngeal carcinoma demonstrated better three-year PFS and overall survival compared to those with $p16^-$ oropharyngeal carcinoma, while tumor EGFR expression did not significantly affect the outcome (48).

However, when examined in combination with chemotherapy in the palliative setting, cetuximab in combination with platinumbased chemotherapy and 5-FU, can prolong the median overall survival from 7.4 months to 10.1 months when applied as a first-line treatment for patients with recurrent or metastatic (R/M) HNSCC (49). Therefore, cetuximab was the first introduced targeted therapy for HNSCC (50). Under special recommendations by the National Comprehensive Cancer Network (NCCN), cetuximab is currently considered to be an option for the treatment of patients with R/M HNSCC alongside cisplatin (45, 51). It can be utilized for both HPV-positive and HPV-negative cases and in an adjuvant, neoadjuvant, or combination setting (52).

Skin toxicities have presented as a major side effect in treatment with cetuximab (53). Therefore, a phase IV study is designed to distinguish between skin reactions caused by radiation and those triggered by cetuximab to assess the incidence and severity of these adverse effects (trial identifier: NCT01553032).

Bevacizumab is humanized mAB targeting VEGF-A. VEGF in general is a stimulator of angiogenesis, a crucial factor for tumor growth and metastasis. Its overexpression has been associated with unfavorable outcomes in HNC (45). Several pro-angiogenic factors besides VEGFR, like EGFR, fibroblast growth factor receptor (FGFR), and platelet-derived growth factor receptor (PDGFR), are upregulated. Therefore, along with bevacizumab, there have been other angiogenesis-inhibiting drugs tested, yielding varying degrees of success (54).

However, till 2019, only bevacizumab has been investigated in phase III clinical trials (54, 55). Still, bevacizumab is not recommended by NCCN due to haematologic and other toxicity effects (56). It has been tested alongside docetaxel, a taxane. Taxanes are a group of chemotherapy drugs, which are a pivotal part of chemotherapeutic regimens, especially in breast, ovarian, non-small cell lung, and head and neck cancer. Acting as microtubule inhibitors by impeding the cell cycle at the G2/M phase, this drug class triggers the activation of cellular apoptosis pathways, thus inhibiting cancer growth (57, 58).

A phase II interventional study including 30 patients with stage III or IV HNSCC treated with a combination of bevacizumab, docetaxel, and radiotherapy to determine the treatment-specific time to progression. The three-year PFS was 61.7% with an overall survival of 68.2%. Locoregional recurrence-free survival was 84.5% and distant metastasis-free survival was 80.5%. The most common non-hematologic toxicities were dysphagia, mucositis, and dermatitis. No grade V toxicities were observed, however, more than 80% of patients developed ≥ grade III lymphopenia and two patients suffered from hemorrhage. Still, Yao et al. claimed this regimen as tolerable and effective in HNSCC (59). A phase III randomized trial (E1305) showed improved response rate and PFS with bevacizumab in addition to conventional chemotherapy in the first-line therapy of R/M HNSCC. Overall response rates were 35.5% with bevacizumab alongside chemotherapy and 24.5% with chemotherapy alone. However, there was no significantly improved overall survival, and toxicities were elevated, including a higher rate of treatment-related grade 3 to 5 bleeding events (6.7% vs. 0.5%) and treatment-related deaths (9.3% vs. 3.5%) with added bevacizumab vs. chemotherapy alone. Median overall survival with bevacizumab plus chemotherapy was 12.6 months and 11.0 months with chemotherapy alone. Moreover, at 2, 3, and 4 years, the overall survival rates were 25.2% vs. 18.1%, 16.4% vs. 10.0%, and 11.8% vs. 6.4% for bevacizumab in addition to chemotherapy vs. chemotherapy alone (60) (Table 1; Figure 1).

2.3 Immune checkpoint inhibitors for the treatment of head and neck cancer

In 2016, ICI were introduced for the treatment of HNSCC, when the anti-PD-1 monoclonal antibodies pembrolizumab and nivolumab received approval from the FDA for the treatment of patients with R/M HNSCC. The consent was supported by two landmark trials, stating that treatment with nivolumab led to longer overall survival compared to standard treatment options, and pembrolizumab showed adequate tolerability and clinically relevant antitumor activity in R/M HNSCC (51, 61, 62).

CheckMate 141, the first reported randomized phase III trial of a PD-1 inhibitor in HNSCC, CheckMate 141, the first reported randomized phase III trial of a PD-1 inhibitor in HNSCC, focused on enrolling patients who had experienced disease progession within six months of undergoing platinum-based chemotherapy. Regardless of tumor PD-L1 status, 361 patients were enrolled in this study. Patients were randomized to receive either 3mg/kg nivolumab every 2 weeks or investigator's choice of weekly systemic standard therapy (methotrexate, weekly docetaxel, or cetuximab. Patients treated with nivolumab exhibited improved median overall survival (7.5 months vs 5.1 months) and a higher overall response rate (13.3% vs. 5.8%) compared to those receiving chemotherapy. At the first interim

analysis, the estimated one-year overall survival was notably higher with nivolumab (36% vs. 16,6%). Moreover, 13.1% patients on nivolumab experienced grade 3/4 treatment related adverse events, as opposed to 35.1% of patients on standard therapy. Upon 2-year follow-up, patients who received nivolumab had a median overall survival of 7.7 months, while those who underwent chemotherapy had a median overall survival of 5.1 months. Antibodies blocking PD-1, namely pembrolizumab and nivolumab, have been added to the treatment of HNSCC (15, 25). Further, Gillison et al. analyzed longterm outcomes with nivolumab as a firstline treatment in recurrent or metastatic HNSCCs. In their 2-year randomized, phase III trial, they found that nivolumab (n=50) improved overall survival compared with the investigator's control therapy (i.e., chemotherapy only (n=26)) from a median of 3.3 months to 7.7 months (63). Wu et al. prospected that more effective cancer immunotherapy can be achieved by using anti-PD-L1 agents in combination with other therapeutics (64). In fact, preclinical evidence from different cancer entities lends supports to therapeutic synergies between chemo- and immunotherapy, primarily due to the activation of innate immune pathways (65, 66).

The palliative first-line therapy standard has fundamentally changed due to the phase III KEYNOTE-048 study, where pembrolizumab alone or in combination with cisplatin, and 5-FU was compared to cetuximab-chemotherapy, in the above-described combination of cetuximab, platinum-based chemotherapy, and 5-FU, in R/M HNSCC. In this study, 301 patients received pembrolizumab alone, 281 pembrolizumab and chemotherapy, and 300 cetuximab and chemotherapy. Both pembrolizumab and pembrolizumab-chemotherapy treatments were compared to cetuximab-chemotherapy. Particularly in tumors with high PD-L1 expression (CPS \geq 20), pembrolizumab monotherapy extended overall survival to 14.9 months compared to 10.3 months, with a significantly better side-effect profile. Albeit the response rate was lower (23.3% vs. 36.1%) compared to cetuximab-chemotherapy. Overall survival also benefited (given a CPS \geq 1) at 12.3 months compared to 10.3 months, but 38.9% of this group initially progressed. Overall survival with pembrolizumab in combination with cisplatin, and 5-FU, also significantly extended at CPS \geq 1, with a similar remission rate, but with a higher side effects rate than pembrolizumab alone, yet comparable to cetuximab-chemotherapy. Patients were followed-up for four years. The median overall survival improved by a minimum of 3.6 months when pembrolizumab was applied either way. In conclusion, first-line pembrolizumab and pembrolizumab-chemotherapy treatment were considered beneficial in R/M HNSCC (67, 68).

Further, PD-1 levels among HNSCC patients infected with HPV have been shown to be elevated (36–38). Additionally, PD-L1 expression is linked to the primary tumor location, postoperative recurrence, survival rate, and PD-1 and p16 expression.

A lower probability for postoperative recurrence was linked to PD-L1⁺/PD-1⁺ (2.5-fold). PD-L1/PD-1 expression, linked to p16 and HPV, in HNSCC patients also led to better overall survival. PD-L1 expression was positively associated with the time of overall survival compared to negative PD-L1 expression (overall survival rate: 82.8% vs 53.5%; overall survival 36.66 months vs. 30.54

Cancer entity	Study/ Trial number	Treatment regimen/Antibody	Cancer specification	Drug targets
Monoclonal Antibodies				
Head and neck cancer	NCT01553032 (Phase IV)	Cetuximab	Locally advanced (Stage III, IVA or IVB) non- metastatic HNC	EGFR
Breast cancer	NCT00411788 (Phase II)	Trastuzumab + Rapamycin	positive metastatic breast cancer	HER-2
Breast cancer	NCT03765983 (Phase II)	Trastuzumab + GDC-0084	positive metastastic breast cancer	HER-2
Immune Checkpoint Inhibitors				
Head and neck cancer	NCT03818061 (Phase II)	Atezolizumab + Bevacizumab	Advanced/metastatic HNSCC	PD-L1 + VEGFR
Head and neck cancer	NCT05187338 (Phase II)	Ipilimumab + Pembrolizumab + Durvalumab	Advanced solid Tumors	CTLA-4 + PD-1 + PD-L1
Head and neck cancer	NCT03765918 (Phase II)	Pembrolizumab	Previously untreated, resectable, locally advanced HNSCC	PD-1

TABLE 1 Summary of clinical trials on monoclonal antibodies and immune checkpoint inhibitors for the treatment of the cancer entities presented in this review (i.e., head and neck cancer, breast cancer, malignant melanoma, sarcoma).

months). In conclusion, these findings promote PD-1-targeted immunotherapy for treatment in HNSCC, specifically in HPV-positive cases (36). Therefore, it is paramount to differentiate between HPV-positive versus HPV-negative HNSCC.

Ongoing studies are investigating other potential immune implications and combinations of cancer-related pathways beyond PD-1 or EGFR An active, not recruiting study aims to investigate the clinical and biological effects of the therapy R/M HNSCC with atezolizumab, targeting PD-L1, in combination with bevacizumab, mAB against VEGF (trial identifier: NCT03818061). Of note, atezolizumab in combination with bevacizumab has been tested in a phase II study for patients with unresectable or metastatic mucosal melanoma, demonstrating a median progression-free survival of 8.2 months, along with an objective responste rate of 45.0% (69). To avoid the above-mentioned ICI treatment failure, pembrolizumab will be used in combination in several studies. For instance, a triple combination of CTLA-4, PD-1, and PDL-1 blocking ICI, ipilimumab, pembrolizumab, and durvalumab, is about to be tested for 100 patients with advanced solid tumors including HNSCC in a recruiting phase I/II trial (trial identifier: 'NCT05187338) (Table 1; Figure 2).

2.4 Additional therapeutic approaches in head and neck cancer

Tyrosine kinase inhibitors (TKI) work by inhibiting pathways that regulate cell growth, division, and survival but do not belong to



Summary of monoclonal antibodies and their pathways for the treatment of cancer entities presented in this review (i.e., head and neck cancer, breast cancer, malignant melanoma, sarcoma).



the group of mAB and ICI from a pharmacological standpoint (70). Yet, these agents can block multiple angiogenic signaling pathways including those of VEGFR, EGFR, FGFR, and PDGFR. Examples are cabozantinib, axitinib, and lenvatinib (55, 71).

A phase II trial investigated cabozantinib, a multi-TKI inhibiting VEGFR 1-3 among other receptors, in combination with pembrolizumab in the treatment of patients with R/M HNSCC. Median overall survival was 22.3 months with median PFS of 14.6 months. Promising clinical activity and tolerability was concluded, however no correlation between tumor mutational burden (TMB) and clinical outcome was observed, warranting for further investigation for similar combinations for HNSCC (71).

2.5 Immunotherapies in head and neck cancer standard therapy – where do we stand?

ICI such as pembrolizumab and nivolumab have been implemented into the treatment algorithm for R/M HNSCC and are an integral part of the most recent American Society of Clinical Oncology (ASCO) Guidelines (72).

For patients with R/M HNSCC, PD-L1 immunohistochemistry and TMB testing are recommended. Pembrolizumab has emerged as a primary treatment option for R/M HNSCC, either paired with platinum/5-FU or used in isolation for patients with a CPS of ≥ 1 (43, 73). In general, PD-L1 CPS of ≥ 1 or TMB of ≥ 10 suggests a positive clinical response to PD-1 inhibitors. Alternatively, based on the performance status, such patients can be administered a combination of pembrolizumab, platinum, and 5-FU. Interestingly, this therapy approach can be employed for cases with CPS < 1, too. Further, pembrolizumab or nivolumab should be administered to patients with platinum-refractory R/M HNSCC, regardless of CPS status (74). ICI targeting PD-1 should be considered if the tumor has progressed after platinum-based therapy. Radiation therapy is recommended as a therapeutic option concurrently with immunotherapy for palliation or local control in patients with oligometastatic HNSCC. Yet, it is not recommended to be given in order to enhance response to immunotherapy outside of clinical trials. In the specific case of TMB-high R/M HNSCC or PD-L1-positive R/M salivary gland cancer, pembrolizumab is recommended as another therapy option (75).

In conclusion, immunotherapy is the new standard approach for the treatment of R/M HNSCC. Positive results have been achieved in neoadjuvant settings, and combination therapy, including TKI, seems to be a promising strategy but warrants further research and broad clinical studies (76). Currently approaching this concern, the KEYNOTE-689 trial, a phase III randomized, open-label study, is designed to assess pembrolizumab in combination with standard treatment – radiotherapy with or without cisplatin – as both neoadjuvant and adjuvant therapy for patients with previously untreated, resectable, locally advanced HNSCC (trial identifier: NCT03765918).

3 Breast cancer – general information

Female BC has taken the lead as the most frequently diagnosed cancer according to global cancer statistics in 2020. Approximately

2.3 million new cases of BC and 680.000 deaths due to BC were estimated worldwide, making it the second leading cause of cancer-related death among women (39, 77).

BC is a generic term referring to a group of heterogeneous malignancies, clinically classified into three primary subtypes based on immunohistochemical markers. Those markers include the status of hormone receptors, either estrogen (ER) or progesterone receptor (PR), and HER-2. Therefore, there are luminal ER⁺ and PR⁺, HER-2⁺, and triple negative BC (TNBC) (78). ER is involved in the main regulation of BC cell growth, PR is considered a significant indicator in carcinogenesis acting as an interdependent partner alongside ER (79). Roughly 15-20% of BC cases exhibit amplification of the HER-2 oncogene, situated on chromosome 17q12. HER-2 is a membrane tyrosine kinase receptor expressed in breast tissue, normal and malignant, indicating pathways for cell proliferation and differentiation. In BC cells, an elevated level of HER-2 is linked to poor prognosis, increased cell proliferation, angiogenesis, and evolution of metastasis (80, 81). Even with adjuvant chemotherapy, the five-year survival rate in metastatic breast cancer remains less than 30% (82, 83). Metastatic recurrence poses a significant challenge for women diagnosed with BC, with 20-30% mortality due to metastatic disease BC (78, 84).

Mutations in the tumor suppressor genes BRCA1 and BRCA2 are responsible for the majority of hereditary BC. Identifying these variants in patients is crucial for determining eligibility for targeted treatment using poly-ADP-ribose polymerase (PARP) inhibitors. Consequently, BRCA1/2 genomic testing is imperative for BC (85). Due to its reliability, accessibility, and simplicity, Sanger DNA sequencing has been recognized as the gold standard for BRCA testing. However, an increase in the application of next-generation sequencing (NGS) technology, which offers recognition of Large Genomic Rearrangements (LGRs) and cost-effectiveness, is observed (86).

3.1 Standard therapy for breast cancer – on the pulse of time?

Breast cancer therapy is individualized based on the cancer's type, stage, and patient's preference and health factors. Treatment strategies involve surgery, radiation, chemotherapy, hormonal, and targeted therapies.

Neoadjuvant therapy was initially investigated in inoperable, locally advanced BC, but has been integrated into the standard of care, particularly in HER-2⁺ early-stage BC, for 2 cm N0 (according to the TMN classification) or all N⁺ BC subtypes, and TNBC (87). Due to the most recent ASCO guidelines, neoadjuvant therapy is suitable for inflammatory BC patients, and for patients with posttherapeutic tumor residuals. Further, neoadjuvant therapy is applied to reduce the extent of surgery in the breast and axilla and to implement individualized post-neoadjuvant strategies. While TNBC patients with clinically node-positive or T1c disease should receive an anthracycline and taxane regimen prior to surgery, T1aN0 and T1bN0 HER-2⁺ patients have no recommendation for routine neoadjuvant treatment. Postmenopausal patients with hormone receptors-positive, HER-2⁻ BC can be administered hormone therapy to downstage disease preoperatively (88). Moreover, tamoxifen or aromatase inhibitors may be used for hormone receptor-positive tumors. Trastuzumab may be given in HER-2-positive tumors alongside chemotherapy. Primary surgery has been proven to be highly effective. Yet, its clinical impact can be potentiated by including the principles above (89).

Adjuvant therapy after surgery is based on the same pillars as neoadjuvant strategy, namely chemotherapy, radiation therapy, hormonal therapy, and targeted therapy, e.g., anti-HER-2 drugs like trastuzumab for HER-2⁺ tumors (78). While radiotherapy remains crucial in adjuvant BC therapy, ER^+ tumors are commonly treated with five to ten years of endocrine therapy and/or chemotherapy. Tamoxifen, for example, is a medication classified as a selective ER modulator, which is used for the treatment of BC in both men and women (and as a prophylactic agent for BC risk reduction) (78, 90).

3.2 The status quo of monoclonal antibodies in breast cancer therapy

mAB have versatile roles in BC therapy, including directly attacking cancer cells, but also facilitating drug delivery to specific targets, inhibiting cell growth, and blocking immune system evasion. While mAB have proven to be efficient in treating HER-2⁺ BC, their potential remains unexplored for other subtypes, particularly TNBC (79, 91).

Trastuzumab was the first FDA-approved anti-HER-2 mAB in 1998, and remains the first-line therapy for early, advanced, and (in combination therapy) also for metastatic, HER-2⁺ BC (92). By binding to the extracellular domain of HER-2 on tumor cells, it stops the downstream pathways and therefore their proliferation (79). Resistance to trastuzumab-based therapy poses a significant challenge, thus anticipation for enhanced therapeutic outcomes lies within investigating drug combinations that target multiple relevant pathways. Interestingly, trastuzumab is already applied in combination with other agents, e.g. anthracyclines and taxanes, in BC. For metastatic BC, many other drugs are trialed besides trastuzumab (80). Further examples of FDA-approved mAB for BC treatment are pertuzumab, another anti-HER-2 mAB, and bevacizumab. Bevacizumab targets VEGF, therefore acting as an anti-angiogenic drug. Erinjeri et al. retrospectively analyzed 1,108 port placements in patients who were treated with bevacizumab. Ports had to be removed in 120 patients of which 11 were BC patients. Following port removal one BC patient presented with wound healing disorders. Overall, patients with an interval between bevacizumab therapy and surgery of less than 14 days demonstrated a significantly higher risk of wound healing disorders (93). This finding can be due to the half-life of bevacizumab which may vary between 11 and 50 days. The GeparQuinto study by Gerber et al. included 127 study sites and looked into outcomes following BC surgery. The authors found no overall rise in surgical complications when bevacizumab was administered concurrently with neoadjuvant anthracycline- and taxane-based chemotherapy for early or locally advanced BC. Overall, 38.1% of all patients included in the study suffered surgical complications, such as bleeding, necrosis, wound

infections, and abscesses. In the group of patients, who were treated without bevacizumab, surgical complications occurred in 10.9% vs. 15.0% when treated with bevacizumab showing no statistical significance. Yet, within specific patient subsets, such as those undergoing breast-conserving surgery or requiring repeated surgical intervention to attain clear margins, the incidence of complications significantly increased among those treated with bevacizumab. Among patients undergoing breast-conserving surgery (N=53), surgical complications occurred in 7.3% following treatment without bevacizumab, and in 13.4% following treatment with bevacizumab (94). Interestingly, to date, no trials correlated ICI in BC therapy with an increased risk of lymphedema, a common complication following breast cancer resection, especially when combined with lymph node dissection (95). Golshan et al. investigated the neoadjuvant administration of bevacizumab in TNBC. While 28 patients received single-agent cisplatin prior to definitive surgery, 51 patients were administered neoadjuvant cisplatin plus bevacizumab preoperative. The authors reported that the use of expanders or implants might be problematic for patients treated with bevacizumab with a trend towards more wound-related events in this group despite not yielding significant differences to the cisplatin-only group. Yet, they emphasized that future research was warranted to deduce concrete recommendations (Supplementary Material S1).

3.3 What do the guidelines say? – immune checkpoint inhibitors in the standard therapy for breast cancer

Immunotherapy has emerged as a promising lifeline for TNBC patients. It is, therefore, recommended, that all patients eligible for immunotherapy treatment undergo tumor tissue PD-L1 testing, regardless of their prior immunotherapy in adjuvant or neoadjuvant settings.

Current immunotherapies regimens are recommended to include anthracyclines and a taxane, with or without carboplatin. Adjuvant immunotherapy should be balanced carefully, especially regarding potential toxicities. Neoadjuvant pembrolizumab and atezolizumab are recommended based on improved pathological complete response rates, regardless of PD-L1 status. Patients with inoperable advanced or metastatic TNBC should undergo tumor tissue testing for PD-L1 and comprehensive genomic profiling, including TMB and MSI tests.

Pembrolizumab alongside chemotherapy, is beneficial for patients with tumors expressing PD-L1 with a CPS of \geq 10. It is recommended as a first-line treatment when combined with nabpaclitaxel, or a carboplatin and gemcitabine combination. However, it is also still only recommended when PD-L1 expression has a CPS of \geq 10. High-risk early-stage TNBC patients can benefit from pembrolizumab with chemotherapy before and after surgery.

Further studies are expected to enhance treatment for earlystage TNBC and other subtypes. Future research may also further explore immunotherapy's impact on hormone receptor-positive and -negative subtypes, and ideal chemotherapy combinations involving ICI (96).

4 Skin cancer – from clinical classifications to genetic profiling

SC is the predominant form of cancer worldwide, accounting for a variety of malignancies. It is generally categorized as non melanoma skin cancer (NMSC), including basal cell carcinoma (BCC) and squamous cell carcinoma (SCC), or as malignant melanoma (MM) arising from the malignant transformation of melanocytes (i.e., melanin-producing cells in the basal layer of the epidermis). Of note, melanocytes originate from the neural crest, indicating that MM can be found in any location, where neural crest cells migrate such as the skin, but also the gastrointestinal tract, the brain or the eye (97, 98). Due to significantly higher mortality rates in MM compared to other SC, this review will focus on MM (97, 98). Of note, immunotherapies (especially ICI) have been shown to be effective therapy agents in NMSC (99).

Multiple epidemiological studies have provided evidence of rising incidence in MM over the past decades (98, 100-102). Global cancer statistics in 2020 reported 325,000 new cases and 57,000 new deaths for MM versus 6.3 million cases and 56,000 deaths for NMSC, underlying the high mortality of MM disease (32). MM commonly affects younger patients compared to other solid tumors and is more common for White people than Black persons or Asians (97, 103). MM development involves multiple factors arising from an interaction between endogenous factors, like genetic predisposition, and exogenous factors. UV solar radiation exposure is the main risk factor for cutaneous malignancies (100, 104). The increased occurrence of UV-induced cytidine to thymidine transitions is associated with a significantly higher mutation rate in MM than in other solid tumors (105, 106). Increased sensitivity to UV radiation, especially in white patients increases the risk of developing MM as pigmentation and melanin are crucial for shielding melanocytes and keratinocytes from UV lights. That is why individuals with lighter skin (i.e., phototypes I and II) face a greater risk for SC (107). Family history is another pivotal risk factor for MM development, while mutations in the tumor suppressor gene cyclin-dependent kinase inhibitor (CDKN2A) are the most common ones responsible for hereditary MM (108, 109). Somatic driver mutations for melanoma include, for example, B rapidly accelerated fibrosarcoma (BRAF), RAS, CDKN2A, neurofibromatosis type 1 (NF1), and phosphatase and tensin homolog gene (PTEN) (110, 111).

4.1 Standard treatment approach for malignant melanoma

For local MM therapy, the primary treatment is surgical removal, showing high survival rates. However, survival rates decrease significantly after metastasis (112, 113). Before the advent of immune checkpoint inhibitors, the five-year relative survival rate for patients with stage IV, metastatic MM was around 10%, depending on the location and metastasis burden. For comparison, the five-year relative survival rate for MM stage 0 (i.e., melanoma in situ) is 97% (97). Beyond surgery, immunotherapy plays a key role in

standard therapy for MM (Please see "From Bench to Beside – Immunotherapy as Standard Therapy for Malignant Melanomas). Applying immunotherapy in MM treatment, improved survival rates. The five-year overall survival in patients with advanced melanoma was 52% when receiving nivolumab and ipilimumab, and 44% when receiving nivolumab alone (114).

4.2 Monoclonal antibodies and immune checkpoint inhibitors for malignant melanoma – effective and targeted therapies?

4.2.1 Monoclonal antibodies for malignant melanoma

Relatlimab, a mAB targeting LAG-3, plus nivolumab, a PD-1targeting mAB, have been approved by the FDA as a first-line treatment for advanced-stage melanoma (115). While inhibition of PD-1, and PD-L1 mostly (not exclusively) affects CD8⁺ T cells, inhibition of LAG-3 is proposed to affect multiple immune cell subtypes, since LAG-3 is found to be more broadly expressed on various cell lines (116). To investigate relatlimab's specific cellular impact on the immune system, Huuhtanen et al. performed a phase I single-cell characterization trial. The authors evaluated relatlimab plus nivolumab, stating that the combination of those drugs, inhibiting LAG-3 and PD-1 affected not only CD8⁺ T cells but also natural killer cells (NK cells) and regulatory T cells (T_{regs}). Adaptive NK cells (i.e., specialized NK cells carrying the potential for immunological memory) experienced an enhancement in responders and transcriptomic changes, while the number of T_{regs} increased. Interestingly, transcriptome analysis showed that the metabolic activity of Tregs decreased. Furthermore, the clonality of T cell receptors, as well as the cytotoxic and NK cell-like activity of their enlarging CD8⁺ T cell clones increased in therapy-responders (117) (Table 1; Figure 1).

Molecular and cellular factors as suitable targets for checkpoint inhibitors in MM have been identified, including PD-L1, major histocompatibility complex (MHC-I), TMB and BRAF mutation, and T cell infiltration. Additionally, MITF, a transcription factor involved in the development and survival of melanocytes, is proposed to have a mechanistic relevance in MM treatment.

Primary targets in melanoma treatment are PD-1, PD-L1, and CTLA-4. Pembrolizumab and nivolumab target PD-1 to enhance the immune response against melanoma cells, whereas ipilimumab binds to CTLA-4 to support the immune response. The efficacy of immune checkpoint inhibition relies on factors involved in the immune balance with melanocyte survival. PD-L1 expression is emphasized as one key element, intricately interwoven with other molecular and cellular players like MHC expression, and mutational load (112, 118). Regardless of the BRAF mutational status, immunotherapies have shown promising efficacy (119). The human monoclonal IgG1 antibody Ipilimumab received its approval for treating advanced or unresectable MM based on survival benefits for patients with advanced MM (120–123).

In a phase III trial, including 676 patients with pretreated metastatic, unresectable stage III or IV melanoma, ipilimumab,

alone or in combination with glycoprotein 100 peptide vaccine (gp100), demonstrated enhanced overall survival compared to gp100 alone. The median overall survival for ipilimumab alone was 10.1 months, combined with gp100 10.0 months, and for gp100 alone 6.4 months. Patients experienced severe therapy-induced immune-related adverse events, most frequently affecting the skin and gastrointestinal tract (i.e., diarrhea, nausea, pruritus), which can be treatment-limiting (120). Ipilimumab was followed by the approval of nivolumab and pembrolizumab (i.e., PD-1 inhibitors) for the therapy of advanced or metastatic melanoma among other tumors. A combination of ipilimumab and nivolumab has been established for the treatment of patients with BRAF wild-type metastatic or unresectable MM (122, 124).

The application of nivolumab, in combination with ipilimumab or by itself, has been shown to be an effective treatment for patients with advanced melanoma. A phase III clinical trial reported a median overall survival rate of more than 60.0 months for patients treated with nivolumab and ipilimumab, 36.9 for patients treated with only nivolumab, and 19.9 months for patients treated with only ipilimumab. The five-year overall survival for patients treated with both agents was 52%, versus 44% with nivolumab, and 26% with ipilimumab. Additionally, quality of life was not permanently impaired (114). A still ongoing multicenter, multicohort, open-label phase Ib/II trial was designed as a first-in-human study to investigate the safety, tolerability, and efficiency of a combination of bi-specific antibody applications, targeting different T-cell co-inhibitory receptors for patients with advanced or metastatic melanoma prior immune checkpoint inhibitor therapy. XmAb2284, CTLA-4 X LAG3 or bavunalimab, targeting both CTLA-4 and LAG-3, and XmAb23104, also known as PD1 X ICOS or XmAb104, targeting PD-1 and ICOS (CD278), a costimulatory molecule for T-cell proliferation and cytokine secretion, are applied intravenously (125) (Table 1; Figure 2).

4.2.2 Immune checkpoint inhibitors for malignant melanoma

T cell immunoglobulin and mucin domain 3 (TIM-3) and T cell immunoreceptor with Ig and ITIM domains (TIGIT) are under investigation as the next wave of co-inhibitory receptor targets. They showcase distinctive functionalities, particularly in regulating the immune system in tissue sites, while being negative regulators of T-cell function like PD-1 and CTLA-4 at the same time. ICI brought about a significant revolution in the treatment of MM, but novel combination therapies are explored to improve response rates (117, 123, 126).

4.3 Treatment approaches for malignant melanoma - beyond monoclonal antibodies and immune checkpoint inhibitors

In this context, vemurafenib and dabrafenib, selective mutant BRAF inhibitors, were approved for patients with metastatic MM and BRAFV600 mutations (127, 128). Targeted therapy includes BRAF and mitogen-activated protein kinase kinase (MEK) inhibitors like vemurafenib, dabrafenib, and trametinib. BRAF, a serine/threonine protein kinase activating the mitogen-activated protein kinase (MAPK) and downstream MEK and ERK signaling pathways, plays a crucial role in tumorigenesis (129, 130). The MEK pathway is promoted by BRAF mutations and might be responsible for melanoma cell proliferation (128, 129, 131, 132). More than 50% of MM cases contain activating BRAF mutations, with a substitution of valine to glutamine in codon 600 (V600E) being the most common BRAF mutation (128, 133).

Clinical phase I and II trials suggested that vemurafenib, also known as PLX4032, represented an effective treatment yielding response rates of more than 50% in MM patients with BRAF V600E mutations (134-137). Chapman et al. investigated the efficacy of vemurafenib compared to dacarbazine, the only chemotherapeutic drug approved for metastatic MM therapy, in a phase III randomized clinical trial including 675 patients with untreated, metastatic MM carrying the BRAF V600E mutation. Vemurafenib showed enhanced overall survival with 84% at six months compared to 64% at six months for dacarbazine use, and improved PFS (135). Still, resistance to BRAF inhibitors over time was shown, most likely due to MAPK-driven acquired resistance. Combining both MEK and mutant BRAF kinase was proposed to delay resistance mechanisms (128). Trametinib is a selective, allosteric MEK1/ MEK2 inhibitor, also known as GSL1120212, which was recently investigated in a phase II clinical trial designed to determine the response rate for patients with metastatic BRAF mutant melanoma. It showed good patient tolerability and significant clinical tumor reduction in patients who were not treated with BRAF inhibitors before, but poor response rates in patients that had been previously treated with a BRAF inhibitor. Kim et al., therefore, stated that these resistance mechanisms against BRAF inhibitors were likely to pass resistance on to MEK inhibitor monotherapy, warranting further investigations (130, 138, 139).

4.4 From bench to beside – immunotherapy as standard therapy for malignant melanomas

While surgery is still the mainstay of treatment, the advent of immunotherapy, specifically the use of the ICI nivolumab and ipilimumab in advanced MM, has revolutionized the therapy of patients with MM (114, 140). In a neoadjuvant setting, an emerging approach for high-risk melanoma may include immunotherapy and agents like ipilimumab, nivolumab, and pembrolizumab, and targeted therapy for melanomas with BRAF mutations.

Adjuvant therapy, aiming to reduce recurrence, also relies on immunotherapy and targeted therapy. The individual treatment approach depends on various factors, like tumor stage, metastasis, and BRAF mutation status. According to the ASCO Guidelines for melanoma treatment, nivolumab and pembrolizumab should be considered for patients with resected stage IIIA/B/C/D BRAF wildtype cutaneous melanoma. These drugs, or a dabrafenib-trametinib combination should be offered for BRAF-mutant tumor patients. No definitive guidance for neoadjuvant cutaneous melanoma treatment has been provided as of 2023. For unresectable or metastatic cases, BRAF wild-type patients should be administered a combination of ipilimumab and nivolumab, nivolumab alone, or pembrolizumab treatment. BRAF-mutant patients can also receive BRAF/MEK inhibitors (e.g., dabrafenib or trametinib). The same treatment strategies are recommended for patients with mucosal melanoma (141, 142).

5 Sarcoma – prevalence, incidence, and risk factors

Broadly speaking, sarcomas manifest as either soft-tissue sarcomas (STS) or bone sarcomas (BS), while STS occur with a four to five times higher prevalence than BS (143). STS encompass a range of mesenchymal malignancies originating from muscles, adipose tissue, tendons, and vasculature structures. They predominantly occur in the extremities (143-145). Among children and adolescents, rhabdomyosarcoma (RMS) and synovial sarcoma are the most frequent subtypes, whereas liposarcoma (LS) and leiomyosarcoma (LMS) are the most common STS observed in adults. Based on prevalence rates, this review will focus on RMS, LS, and LMS (143, 144). RMS is categorized into distinct clinical subtypes based on histopathological features including embryonal RMS (ERMS), alveolar RMS (ARMS), pleomorphic RMS, and spindle cell/sclerosing RMS (ssRMS). ERMS and ARMS are the most common subtypes, whereas the other subtypes are considered rare. PRMS stands out due to its non-responsiveness to chemotherapy, unlike ERMS and ARMS. It is often treated with radiation therapy and wide excision (146, 147).

For STS, 13,400 new cancer cases, and 5,140 new deaths in 2023 were estimated for the United States of America (39). STS can be attributed to environmental and/or genetic risk factors (148–150). Germline mutations in the tumor suppressor gene TP53 are accountable for LFS and can contribute to the early onset of sarcomas and other tumors by causing genomic instability (149–152). Even though STS show a relatively low incidence, STS represents an aggressive oncological disease leading to high mortality rates (143). Over the past two decades, no substantial change in the overall and one-year mortality rates among various histological subtypes has been observed.

5.1 Standard treatment approach in sarcoma therapy

Surgical resection is the backbone of sarcoma therapy aiming for complete resection with negative margins (i.e., R0), but additional drug therapy is commonly indicated to avoid recurrence and enable resection (144, 153).

RMS is commonly treated with a multimodal approach including chemotherapy, surgical resection, and/or radiation therapy. Localized RMS is curable for most patients. Chemotherapy includes drugs like vincristine, actinomycin D, and cyclophosphamide. Surgery aims for complete tumor removal, with radiation therapy used for unresectable tumors, or if there's a high risk of local recurrence (154). Radiation therapy can be employed both in a neoadjuvant and adjuvant approach to impair local recurrence. Anthracycline-based chemotherapy approaches remain the standard initial therapy for LS, both resectable and unresectable. Doxorubicin is applied either as a single agent or in combination with ifosfamide (155, 156). It is important to mention, that treatment option for LS rely on the tumor type and stage. LS can be differentiated into several subtypes, including well-differentiated liposarcoma (WDLPS), dedifferentiated liposarcoma (DDLPS), pleomorphic liposarcoma, and myxoid/round-cell liposarcoma (MRCLS). Those subtypes respond differently to systemic therapies. The most frequent subtypes WDLPS and DDLPS remain challenging to treat due to their low response rate to standard chemotherapy. However, MRCLS, for example, shows higher sensitivity to doxorubicinbased regimens (157).

Surgery is often applied in the treatment of localized LMS, also approaching for R0 resection. For unresectable or metastatic cases, doxorubicin is the most commonly used first-line agent in chemotherapy. For some patients, radiation therapy may be beneficial as an adjuvant therapy strategy, or in palliative cases (158).

However, even with combined (neo-)adjuvant treatment strategies, approximately 50% of treated patients still show relapse episodes. For metastatic and refractory sarcomas, the median survival rate remains merely 12 to 18 months (159, 160). Consequently, novel therapeutic strategies, notably antibodybased therapies are warranted to advance sarcoma treatment and improve patient prognosis (161).

5.2 Monoclonal antibodies in sarcoma therapy – fact or fiction?

Olaratumab, a human mAB targeting PDGFR- α , showed enhanced overall survival in phase II trials, when applied in combination with the chemotherapeutic agent doxorubicin, compared to only doxorubicin, for patients with advanced or R/ M STS (162, 163). Thus, it got approved by the FDA, but recently has been taken off the market due to underwhelming findings in phase III studies. Tap et al. stated no significant benefit in overall survival by adding olaratumab to doxorubicin in a double-blinded, randomized trial, including 509 patients from 110 sites in 25 countries (162, 164, 165).

A murine IgG1 mAB called 8H9 targets a distinctive cell surface tumor antigen found in neuroectodermal, epithelial, and mesenchymal tumors, including RMS. More precisely, 8H9 targets B7-H3 (also referred to as CD276) by binding to a region that is crucial for the immunologic function of this immune checkpoint and immunoregulatory molecule. Of note, B7-H3 displays elevated expression in tumor tissues versus suppressed expression in normal tissues. Thus, it has been discussed to carry untapped potential as a target for cancer immunotherapy (166, 167). Modak et al. reported *in vitro* characterization of radiolabeled 8H9 and its potential application *in vivo* with subcutaneous human RMS, stating that the utilization of radiolabeled 8H9 showed effective targeting of RMS xenografts and holds promising potential for potentiating radioimmunotherapy (168). Yet, further studies are warranted to corroborate its clinical impact (Table 1; Figure 1).

5.3 Immunotherapeutic and epigenetic options for sarcoma patients

5.3.1 Monoclonal antibodies for sarcoma patients

Pembrolizumab, nivolumab, and ipilimumab have been investigated for their application in sarcoma treatment (169). A phase I/II study of pembrolizumab and doxorubicin for the treatment of unresectable R/M sarcoma did not reach its primary endpoint (i.e., objective response rate) but demonstrated a notable improvement in PFS when compared to conventional treatment protocols. Patients who received pembrolizumab and doxorubicin showed a median PFS of 8.1 months compared to single doxorubicin with a median PFS of under five months (170).

Zhou et al. proposed combining ipilimumab and nivolumab as an efficient and properly tolerated treatment option for advanced STS (171). Chen et al. reviewed 150 patients with untreated PD-L1 positive R/M STS, who have been therapied with nivolumab, ipilimumab, or nivolumab alone. An improved survival rate of 12.2 months compared to 9.2 months was reached when patients received a combination of both drugs compared to nivolumab alone. However, the application of both drugs showed less tolerability compared to nivolumab alone (172) (Table 1; Figure 2).

5.3.2 Immune checkpoint inhibitors and epigenetic options for sarcoma patients

Undifferentiated pleomorphic sarcoma often presents with a notably dense immune infiltration. ICI targeting PD-1, PD-L1, and CTLA-4 have shown efficacy, with response rates varying between 20% and 40%. A recent phase II trial of neoadjuvant ICI with concurrent radiotherapy (n=10), exhibited a 90% pathologic response rate for nivolumab/ipilimumab in patients with extremity/truncal undifferentiated pleomorphic sarcoma. Adverse effects observed in this study were in line with the known safety profiles of nivolumab and the combination nivolumab/ipilimumab. Postoperative complications, including one anastomotic leak, anemia, and one wound infection also were consistent with expectations. Stating there is no validated cutoff for pathologic response in STS, they used 30% hyalinization as the optimal cutoff for pathologic response with receiver-operating curves based on landmark analysis of early relapse 52 weeks after surgery. A minimum of 30% hyalinization was found in 90% of patients with undifferentiated pleomorphic sarcoma. Also, overall survival at 24 months was 90% in patients with undifferentiated pleomorphic sarcoma (173, 174).

Carcinogenesis extends beyond genetic alterations and encompasses significant involvement of epigenetic mechanisms, especially histone modifications like acetylation, which is managed by histone deacetylase (HDAC). Inhibition of HDAC has been shown to induce apoptosis, differentiation, and cell cycle arrest while reducing angiogenesis and modulating immune response for cancer cells. HDAC inhibitors are tested in several studies for various cancer types, like lymphoma, or BC (175).

An interaction between HDAC inhibitors and the multi-kinase inhibitor pazopanib, both in vitro and in vivo, has been recently evidenced to induce apoptosis in sarcoma cells and decrease tumor growth (175-177). Pazopanib is a multi-TKI, with targets including VEGFR 1-3, and PDGFR- α , and - β , already approved for the treatment of advanced renal cell carcinoma in the European Union and US (178). Axitinib is another TKI, that has been demonstrated to selectively inhibit VEGFR 1-3 and interact with HDAC inhibitors, ultimately leading to the apoptosis of sarcoma tumor cells in vivo and in vitro (176, 179). Axitinib was examined in combination with pembrolizumab in patients suffering from advanced sarcomas in a phase II trial, stating a 3-month PFS of 65.6% for eligible patients and 72.7% for patients with alveolar soft-part sarcoma. The most frequent grade 3/4 treatment-related adverse events encompassed hypertension and autoimmune toxicities, both in 15% of 33 patients enrolled. Additionally, the authors of the study reported serious treatment-related adverse events such as autoimmune colitis, transaminitis, pneumothorax, and hemoptysis in 21% of patients. Thus, Wilky et al. stated that the combination of axitinib and pembrolizumab showed promising antitumoral activity and manageable toxicity in patients with advanced sarcomas. However, further investigations are warranted to pinpoint this treatment combination's clinical and mechanistic effects (180).

5.4 Immunotherapies in sarcoma patients – the most recent guideline updates

Atezolizumab is recommended for unresectable or metastatic alveolar soft-part sarcoma. Pembrolizumab is not specifically FDAapproved for STS, but has been granted approval for specific patients with advanced sarcoma showing high MSI, DNA mismatch repair deficiency (dMMR), or high TMB. Dostarlimab, targeting the PD-1/PD-L1 pathway is approved for patients with advanced sarcoma exhibiting dMMR. Denosumab, a mAB targeting the RANKL pathway, is approved for certain subgroups of bone cancer patients (181, 182).

6 Discussion

Tumor therapy is a dynamic field and this work shows, it is crucial to work within a multidisciplinary team. Recent advancements have paved the way toward promising treatments that improve complete remission rates and prolong PFS. The knowledge of the strengths and limitations of novel mAB and ICI is paramount to providing effective therapy concepts and leveraging surgical and non-surgical therapies. Thus, reconstructive surgeons should be updated on the latest oncological therapy trends and further explore potential links between mAB/ICI administration and surgical outcomes.

Overall, future clinical trials are warranted to explore further the potential of mAB and ICI in the presented cancer entities. There is a lack of ongoing clinical trials in skin cancer and sarcoma (Table 1). Currently, scrutinized target structures for ICI include PD-1 and its ligand PD-L1, which are both inhibitors of antitumoral T cells. Targets of mAB include EGFR and VEGFR, and HER-2. EGFR and VEGFR are growth-factor receptors, and HER-2 is a membrane tyrosine kinase receptor expressed in breast tissue, an oncogene in BC (13, 14, 17, 80, 81).

Beyond those structures, more targets show potential. Anti-RANKL mAB therapy shows potential in BRCA1-mutational driven BC and is approved for some types of bone cancer (183) (181, 182). In BC, CD4/6 inhibitors are explored - looking at the crucial role of the cell cycle in tumor development (184–188). CTLA-4 is one of the main targets in mAB therapy in MM, but more structures are scrutinized as promising targets like LAG-3 (117). For ICI in MM, TIM-3, and TIGIT are currently under investigation as coinhibitory-receptor targets (126). Beyond mAb and ICI, BRAF inhibitors have been investigated in MM treatment (138). mAB for sarcoma therapy focus on PDGFR- α and B7-H3, while ICI involve HDAC-inhibitors. Further, the application of multi-TKIs targeting VEGFR 1-3, and PDGFR- α , and - β , is under current investigation for sarcoma therapy (169).

This review condenses the current body of evidence on mAB and ICI in HNC, SC, BC, and sarcoma therapy. Ultimately, this line of research goes beyond the scalpel and may serve as an evidencebased fundament to optimize the therapeutic efficacy of surgical and non-surgical treatments.

Author contributions

LK: Writing – original draft, Writing – review & editing. LH: Writing – original draft, Writing – review & editing. KaH: Writing – review & editing, Writing – original draft. MA: Writing – review & editing, Writing – original draft. KoH: Writing – review & editing, Writing – original draft. HH: Writing – review & editing, Writing – original draft. SB: Writing – original draft, Writing – review & editing. VS: Writing – original draft, Writing – review & editing. VS: Writing – original draft, Writing – review & editing. MP: Writing – review & editing. AS: Writing – original draft, Writing – review & editing. MP: Writing – original draft, Writing – review & editing. SK: Writing – original draft, Writing – review & editing. BP: Writing – review & editing. MK: Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. LH is receiving an individual research grant by the German Research Foundation (DFG).

Conflict of interest

MP received speaker's honoraria and travel funds from Amgen, Kite, and Takeda.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be constructed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the

References

Roser M, Ritchie H. Cancer. Kennesaw, GA, USA: CANCER Editorial Office. (2015).
Society AC. Cancer Facts & Figures 2022. Atlanta: American Cancer Society (2022).

3. Mejareh ZN, Abdollahi B, Hoseinipalangi Z, Jeze MS, Hosseinifard H, Rafiei S, et al. Global, regional, and national prevalence of depression among cancer patients: A systematic review and meta-analysis. *Indian J Psychiatry*. (2021) 63:527–35. doi: 10.4103/indianjpsychiatry.indianjpsychiatry_77_21

4. Burotto M, Wilkerson J, Stein WD, Bates SE, Fojo T. Adjuvant and neoadjuvant cancer therapies: A historical review and a rational approach to understand outcomes. *Semin Oncol.* (2019) 46:83–99. doi: 10.1053/j.seminoncol.2019.01.002

5. Shien T, Iwata H. Adjuvant and neoadjuvant therapy for breast cancer. Jpn J Clin Oncol. (2020) 50:225–9. doi: 10.1093/jjco/hyz213

6. Klaiber U, Hackert T, Neoptolemos JP. Adjuvant treatment for pancreatic cancer. Transl Gastroenterol Hepatol. (2019) 4:27. doi: 10.21037/tgh

7. Wolpin BM, Meyerhardt JA, Mamon HJ, Mayer RJ. Adjuvant treatment of colorectal cancer. CA Cancer J Clin. (2007) 57:168–85. doi: 10.3322/canjclin.57.3.168

 Emons G, Vordermark D. Adjuvant treatment for endometrial cancer. Curr Opin Oncol. (2019) 31:404–10. doi: 10.1097/CCO.000000000000558

9. Pirker R, Filipits M. Adjuvant therapy in patients with completely resected nonsmall-cell lung cancer: current status and perspectives. *Clin Lung Cancer*. (2019) 20:1– 6. doi: 10.1016/j.cllc.2018.09.016

10. Yahya EB, Alqadhi AM. Recent trends in cancer therapy: A review on the current state of gene delivery. *Life Sci.* (2021) 269:119087. doi: 10.1016/j.lfs.2021.119087

11. Adams GP, Weiner LM. Monoclonal antibody therapy of cancer. *Nat Biotechnol.* (2005) 23:1147–57. doi: 10.1038/nbt1137

12. Darvin P, Toor SM, Sasidharan Nair V, Elkord E. Immune checkpoint inhibitors: recent progress and potential biomarkers. *Exp Mol Med.* (2018) 50:1–11. doi: 10.1038/ s12276-018-0191-1

13. Seidel JA, Otsuka A, Kabashima K. Anti-PD-1 and anti-CTLA-4 therapies in cancer: mechanisms of action, efficacy, and limitations. *Front Oncol.* (2018) 8:86. doi: 10.3389/fonc.2018.00086

14. Fasano M, Corte CMD, Liello RD, Viscardi G, Sparano F, Iacovino ML, et al. Immunotherapy for head and neck cancer: Present and future. *Crit Rev Oncol Hematol.* (2022) 174:103679. doi: 10.1016/j.critrevonc.2022.103679

15. Yamashita K, lwatsuki M, Harada K, Eto K, Hiyoshi Y, Ishimoto T, et al. Prognostic impacts of the combined positive score and the tumor proportion score for programmed death ligand-1 expression by double immunohistochemical staining in patients with advanced gastric cancer. *Gastric Cancer*. (2020) 23:95–104. doi: 10.1007/s10120-019-00999-9

16. Tsao LC, Force J, Hartman ZC. Mechanisms of therapeutic antitumor monoclonal antibodies. *Cancer Res.* (2021) 81:4641-51. doi: 10.1158/0008-5472.CAN-21-1109

17. Bayer V. An overview of monoclonal antibodies. Semin Oncol Nurs. (2019) 35:150927. doi: 10.1016/j.soncn.2019.08.006

18. Weiner LM, Surana R, Wang S. Monoclonal antibodies: versatile platforms for cancer immunotherapy. *Nat Rev Immunol.* (2010) 10:317–27. doi: 10.1038/nri2744

19. Bajwa R, Cheema A, Khan T, Amirpour A, Paul A, Chaughtai S, et al. Adverse effects of immune checkpoint inhibitors (Programmed death-1 inhibitors and cytotoxic T-lymphocyte-associated protein-4 inhibitors): results of a retrospective study. *J Clin Med Res.* (2019) 11:225–36. doi: 10.14740/jocmr3750

20. Weber JS, Postow M, Lao CD, Schadendorf D. Management of adverse events following treatment with anti-programmed death-1 agents. *Oncologist*. (2016) 21:1230–40. doi: 10.1634/theoncologist.2016-0055

21. Hofmann L, Forschner A, Loquai C, Goldinger SM, Zimmer L, Ugurel S, et al. Cutaneous, gastrointestinal, hepatic, endocrine, and renal side-effects of anti-PD-1 therapy. *Eur J Cancer*. (2016) 60:190–209. doi: 10.1016/j.ejca.2016.02.025

reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2024.1276306/ full#supplementary-material

22. Spain L, Diem S, Larkin J. Management of toxicities of immune checkpoint inhibitors. *Cancer Treat Rev.* (2016) 44:51–60. doi: 10.1016/j.ctrv.2016.02.001

23. Hansel TT, Kropshofer H, Singer T, Mitchell JA, George AJ. The safety and side effects of monoclonal antibodies. *Nat Rev Drug Discovery*. (2010) 9:325-38. doi: 10.1038/nrd3003

24. Baldo BA. Adverse events to monoclonal antibodies used for cancer therapy: Focus on hypersensitivity responses. *Oncoimmunology*. (2013) 2:e26333. doi: 10.4161/ onci.26333

25. Ma SJ, Oladeru 0T, Miccio JA, lovoli AJ, Hermann GM, Singh AK. Association of timing of adjuvant therapy with survival in patients with resected stage I to II pancreatic cancer. JAMA Netw Open. (2019) 2:e199126. doi: 10.1001/jamanetworkopen.2019.9126

26. Thomas B, Bigdeli AK, Nolte S, Gazyakan E, Harhaus L, Bischel O, et al. The therapeutic role of plastic and reconstructive surgery in the interdisciplinary treatment of soft-tissue sarcomas in Germany-cross-sectional results of a prospective nationwide observational study (PROSa). *Cancers (Basel)*. (2022) 14. doi: 10.3390/cancers14174312

27. Lese I, Baesu C, Hoyos IA, Pais MA, Klenke F, Kollar A, et al. Flap reconstruction outcome following surgical resection of soft tissue and bone sarcoma in the setting of (Neo)adjuvant therapy: A sarcoma center experience. *Cancers (Basel).* (2023) 15. doi: 10.3390/cancers15092423

 Saldanha IJ, Broyles JM, Adam GP, Cao W, Bhuma MR, Mehta S, et al. Implantbased breast reconstruction after mastectomy for breast cancer: A systematic review and meta-analysis. *Plast Reconstr Surg Glob Open*. (2022) 10:e4179. doi: 10.1097/ GOX.0000000000004179

29. Ray E. Head and neck reconstructive surgery. Cancer Treat Res. (2018) 174:123-43.

30. Bhat GR, Hyole RG, Li J. Head and neck cancer: Current challenges and future perspectives. *Adv Cancer Res.* (2021) 152:67–102. doi: 10.1016/bs.acr.2021.05.002

31. Johnson DE, Burtness B, Leemans CR, Lui VWY, Bauman JE, Grandis JR. Head and neck squamous cell carcinoma. *Nat Rev Dis Primers*. (2020) 6:92. doi: 10.1038/s41572-020-00224-3

32. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* (2021) 71:209–49. doi: 10.3322/caac.21660

33. Cancer, I.A.f.R.o. Cancer today(2020). Available online at: https://gco.iarc.fr/today/online-analysis-pie?v=2020&mode=population&mode_population=continents&population=900&key=total&sex=0&cancer=1_2_3_4_5_1_14&type=0&statistic=5&prevalence=0&population_group=0&ages_group%5B&5D=0&ages_group%5B&5D=17&hb_items=7&group_cancer=1&include_nmsc=1&include_nmsc=1&include_nmsc=0&includ

34. Gormley M, Creaney G, Schache A, Ingarfield K, Conway DI. Reviewing the epidemiology of head and neck cancer: definitions, trends and risk factors. *Br Dent J.* (2022) 233:780–6. doi: 10.1038/s41415-022-5166-x

35. Bravi F, Lee YA, Hashibe M, Boffetta P, Conway DI, Ferraroni M, et al. Lessons learned from the INHANCE consortium: An overview of recent results on head and neck cancer. *Oral Dis.* (2021) 27:73–93. doi: 10.1111/odi.13502

36. Chen SW, Li SH, Shi DB, Jiang WM, Song M, Yang AK, et al. Expression of PD-1/PD-L1 in head and neck squamous cell carcinoma and its clinical significance. *Int J Biol Markers*. (2019) 34:398–405. doi: 10.1177/1724600819884722

37. Rampias T, Pectasides E, Prasad M, Sasaki C, Gouveris P, Dimou A, et al. Molecular profile of head and neck squamous cell carcinomas bearing p16 high phenotype. *Ann Oncol.* (2013) 24:2124–31. doi: 10.1093/annonc/mdt013

38. El-Naggar AK, Westra WH. p16 expression as a surrogate marker for HPVrelated oropharyngeal carcinoma: a guide for interpretative relevance and consistency. *Head Neck*. (2012) 34:459–61.

39. Siegel RL, Miller KD, Wagle NS, Jemal A. Cancer statistics, 2023. CA Cancer J Clin. (2023) 73:17–48. doi: 10.3322/caac.21763

40. Hashibe M, Brennan P, Chuang SC, Boccia S, Castellsague X, Chen C, et al. Interaction between tobacco and alcohol use and the risk of head and neck cancer: pooled analysis in the International Head and Neck Cancer Epidemiology Consortium. *Cancer Epidemiol Biomarkers Prev.* (2009) 18:541–50.

41. Velleuer E, Dietrich R. Fanconi anemia: young patients at high risk for squamous cell carcinoma. *Mol Cell Pediatr*. (2014) 1:9. doi: 10.1186/s40348-014-0009-8

42. Anderson G, Ebadi M, Vo K, Novak J, Govindarajan A, Amini A. An updated review on head and neck cancer treatment with radiation therapy. *Cancers (Basel)*. (2021) 13. doi: 10.3390/cancers13194912

43. Pfister DG, Spencer S, Adelstein D, Adkins D, Anzai Y, Brizel DM, et al. Head and neck cancers, version 2.2020, NCCN clinical practice guidelines in oncology. *J Natl Compr Canc Netw.* (2020) 18:873–98.

44. Nandakumar A, Nandakumar A. Survival in head and neck cancers - results of A multi- institution study. *Asian Pac J Cancer Prev.* (2016) 17:1745–54. doi: 10.7314/APJCP.2016.17.4.1745

45. Khadela A, Shah Y, Mistry P, Mansuri M, Sureja D, Bodiwala K. A review of efficacy and safety of cetuximab and bevacizumab-based monoclonal antibodies in head and neck cancer. *Med Oncol.* (2022) 40:66. doi: 10.1007/s12032-022-01939-x

46. Wee P, Wang Z. Epidermal growth factor receptor cell proliferation signaling pathways. *Cancers (Basel)*. (2017) 9. doi: 10.3390/cancers9050052

47. Bonner JA, Harari PM, Gira It J, Azarnia N, Shin DM, Cohen RB, et al. Radiotherapy plus cetuximab for squamous-cell carcinoma of the head and neck. N Engl J Med. (2006) 354:567–78. doi: 10.1056/NEJMoa053422

48. Ang KK, Zhang Q, Rosenthal DI, Nguyen-Tan PF, Sherman EJ, Weber RS, et al. Randomized phase III trial of concurrent accelerated radiation plus cisplatin with or without cetuximab for stage III to IV head and neck carcinoma: RTOG 0522. *J Clin Oncol.* (2014) 32:2940–50. doi: 10.1200/JCO.2013.53.5633

49. Vermorken JB, Mesia R, Rivera F, Remenar E, Kawecki A, Rottey S, et al. Platinum-based chemotherapy plus cetuximab in head and neck cancer. *N Engl J Med.* (2008) 359:1116–27. doi: 10.1056/NEJMoa0802656

50. Oncology, G.S.f.H.a.M. Head and neck squamous cell carcinomas(2022). Available online at: https://www.onkopedia-guidelines.info/en/onkopedia/guidelines/ head-and-neck-squamous-cell-carcinomas/@@guideline/html/index.html.

51. Cohen EEW, Bell RB, Bifulco CB, Burtness B, Gillison ML, Harrington KJ, et al. The Society for Immunotherapy of Cancer consensus statement on immunotherapy for the treatment of squamous cell carcinoma of the head and neck (HNSCC). *J Immunother Cancer.* (2019) 7:184. doi: 10.1186/s40425-019-0662-5

52. Moskovitz J, Moy J, Ferris RL. Immunotherapy for head and neck squamous cell carcinoma. *Curr Oncol Rep.* (2018) 20:22. doi: 10.1007/s11912-018-0654-5

53. Pinto C, Barone CA, Girolomoni G, Russi EG, Merlano MC, Ferrari D, et al. Management of skin toxicity associated with cetuximab treatment in combination with chemotherapy or radiotherapy. *Oncologist.* (2011) 16:228–38. doi: 10.1634/ theoncologist.2010-0298

54. Hyytiainen A, Wahbi W, Vayrynen O, Saarilahti K, Karihtala P, Salo T, et al. Angiogenesis inhibitors for head and neck squamous cell carcinoma treatment: is there still hope? *Front Oncol.* (2021) 11:683570. doi: 10.3389/fonc.2021.683570

55. Micaily I, Johnson J, Argiris A. An update on angiogenesis targeting in head and neck squamous cell carcinoma. *Cancers Head Neck*. (2020) 5:5. doi: 10.1186/s41199-020-00051-9

56. Schutz FA, Jardim DL, Je Y, Choueiri TK. Haematologic toxicities associated with the addition of bevacizumab in cancer patients. *Eur J Cancer*. (2011) 47:1161–74. doi: 10.1016/j.ejca.2011.03.005

57. De Felice F. Docetaxel plus radiotherapy for head and neck cancer. *Oral Oncol.* (2023) 139:106359. doi: 10.1016/j.oraloncology.2023.106359

58. Ismail U, Killeen RB. Taxane Toxicity. Treasure Island (FL: StatPearls (2024).

59. Yao M, Galanopoulos N, Lavertu P, Fu P, Gibson M, Argiris A, et al. Phase II study of bevacizumab in combination with docetaxel and radiation in locally advanced squamous cell carcinoma of the head and neck. *Head Neck*. (2015) 37:1665–71. doi: 10.1002/hed.v37.11

60. Argiris A, Li S, Savvides P, Ohr JP, Gilbert J, Levine MA, et al. Phase III randomized trial of chemotherapy with or without bevacizumab in patients with recurrent or metastatic head and neck cancer. *J Clin Oncol.* (2019) 37:3266–74. doi: 10.1200/JCO.19.00555

61. Seiwert TY, Burtness B, Mehra R, Weiss J, Berger R, Eder JP, et al. Safety and clinical activity of pembrolizumab for treatment of recurrent or metastatic squamous cell carcinoma of the head and neck (KEYNOTE-012): an open-label, multicentre, phase 1b trial. *Lancet Oncol.* (2016) 17:956–65. doi: 10.1016/S1470-2045 (16)30066-3

62. Ferris RL, Blumenschein G, Jr Fayette J, Guigay J, Colevas AD, Licitra L, et al. Nivolumab vs investigator's choice in recurrent or metastatic squamous cell carcinoma of the head and neck: 2-year long-term survival update of CheckMate 141 with analyses by tumor PD-L1 expression. *Oral Oncol.* (2018) 81:45–51. doi: 10.1016/j.oraloncology.2018.04.008

63. Gillison ML, Blumenschein G, Fayette J, Guigay J, Colevas AD, Licitra L, et al. Long-term outcomes with nivolumab as first-line treatment in recurrent or metastatic head and neck cancer: subgroup analysis of checkMate 141. *Oncologist.* (2022) 27:e194–8. doi: 10.1093/oncolo/oyab036

64. Wu Y, Chen W, Xu ZP, Gu W. PD-L1 distribution and perspective for cancer immunotherapy-blockade, knockdown, or inhibition. *Front Immunol.* (2019) 10:2022. doi: 10.3389/fimmu.2019.02022

65. Xue Y, Gao S, Gou J, Yin T, He H, Wang Y, et al. Platinum-based chemotherapy in combination with PD-1/PD-L1 inhibitors: preclinical and clinical studies and mechanism of action. *Expert Opin Drug Delivery*. (2021) 18:187–203. doi: 10.1080/17425247.2021.1825376

66. Lu S, Concha-Benavente F, Shayan G, Srivastava RM, Gibson SP, Wang L, et al. STING activation enhances cetuximab-mediated NK cell activation and DC maturation and correlates with HPV(+) status in head and neck cancer. *Oral Oncol.* (2018) 78:186–93. doi: 10.1016/j.oraloncology.2018.01.019

67. Harrington KJ, Burtness B, Greil R, Soulieres D, Tahara M, de Castro G Jr, et al. Pembrolizumab with or without chemotherapy in recurrent or metastatic head and neck squamous cell carcinoma: updated results of the phase III KEYNOTE-048 study. J Clin Oncol. (2023) 41:790–802. doi: 10.1200/JCO.21.02508

68. Burtness B, Rischin D, Greil R, Soulieres D, Tahara M, de Castro G Jr, et al. Pembrolizumab alone or with chemotherapy for recurrent/metastatic head and neck squamous cell carcinoma in KEYNOTE-048: subgroup analysis by programmed death ligand-1 combined positive score. J Clin Oncol. (2022) 40:2321–32. doi: 10.1200/ JCO.21.02198

69. Mao L, Fang M, Chen Y, Wei X, Cao J, Lin J, et al. Atezolizumab plus bevacizumab in patients with unresectable or metastatic mucosal melanoma: A multicenter, open-label, single-arm phase II study. *Clin Cancer Res.* (2022) 28:4642–8. doi: 10.1158/1078-0432.CCR-22-1528

70. Hartmann JT, Haap M, Kopp HG, Lipp HP. Tyrosine kinase inhibitors - a review on pharmacology, metabolism and side effects. *Curr Drug Metab.* (2009) 10:470–81. doi: 10.2174/138920009788897975

71. Saba NF, Steuer CE, Ekpenyong A, McCook-Veal A, Magliocca K, Patel M, et al. Pembrolizumab and cabozantinib in recurrent metastatic head and neck squamous cell carcinoma: a phase 2 trial. *Nat Med.* (2023) 29:880–7. doi: 10.1038/s41591-023-02275-x

72. Yilmaz E, Ismaila N, Bauman JE, Dabney R, Gan G, Jordan R, et al. Immunotherapy and biomarker testing in recurrent and metastatic head and neck cancers: ASCO guideline. J Clin Oncol. (2023) 41:1132–46. doi: 10.1200/JCO.22.02328

73. Surucu A, Hou T, Kuhar M, Durm G, Mesa H. Comparison of PD-L1 IHC 22C3 pharmDx combined positive score (CPS) in primary versus metastatic nodal squamous cell carcinomas of the head and neck: is there a significant difference? *Appl Immunohistochem Mol Morphol.* (2023). doi: 10.1097/PAI.00000000001140

74. Zhao M, Shao T, Ren Y, Zhou C, Tang W. Identifying optimal PD-1/PD-L1 inhibitors in first-line treatment of patients with advanced squamous non-small cell lung cancer in China: Updated systematic review and network meta-analysis. *Front Pharmacol.* (2022) 13:910656. doi: 10.3389/fphar.2022.910656

75. Yilmaz E, Ismaila N, Dabney R, Saba NF, Mell LK. Immunotherapy and biomarker testing in recurrent and metastatic head and neck cancers: ASCO guideline Q and A. JCO Oncol Pract. (2023) 19:194–6. doi: 10.1200/OP.22.00802

76. Vos JL, Elbers JBW, Krijgsman O, Traets JJH, Qiao X, van der Leun AM, et al. Neoadjuvant immunotherapy with nivolumab and ipilimumab induces major pathological responses in patients with head and neck squamous cell carcinoma. *Nat Commun.* (2021) 12:7348.

77. Giaquinto AN, Sung H, Miller KD, Kramer JL, Newman LA, Minihan A, et al. Breast cancer statistics, 2022. *CA Cancer J Clin.* (2022) 72:524–41. doi: 10.3322/caac.21754

78. Loibl S, Poortmans P, Morrow M, Denkert C, Curigliano G. Breast cancer. Lancet. (2021) 397:1750-69. doi: 10.1016/S0140-6736(20)32381-3

79. Behl A, Wani ZA, Das NN, Parmar VS, Len C, Malhotra S, et al. Monoclonal antibodies in breast cancer: A critical appraisal. *Crit Rev Oncol Hematol.* (2023) 183:103915. doi: 10.1016/j.critrevonc.2023.103915

80. Maximiano S, Magalhaes P, Guerreiro MP, Morgado M. Trastuzumab in the treatment of breast cancer. *BioDrugs*. (2016) 30:75-86. doi: 10.1007/s40259-016-0162-9

81. Krishnamurti U, Silverman JF. HER2 in breast cancer: a review and update. Adv Anat Pathol. (2014) 21:100–7. doi: 10.1097/PAP.000000000000015

82. Kashyap D, Pal D, Sharma R, Garg VK, Goel N, Koundal D, et al. Global increase in breast cancer incidence: risk factors and preventive measures. *BioMed Res Int 2022*. (2022) p:9605439. doi: 10.1155/2022/9605439

83. Riggio AI, Varley KE, Welm AL. The lingering mysteries of metastatic recurrence in breast cancer. *Br J Cancer*. (2021) 124:13–26. doi: 10.1038/s41416-020-01161-4

84. Early Breast Cancer Trialists' Collaborative, G. Effects of chemotherapy and hormonal therapy for early breast cancer on recurrence and 15-year survival: an overview of the randomised trials. *Lancet.* (2005) 365:1687–717. doi: 10.1016/S0140-6736(05)66544-0

85. Lux MP, Fasching PA. Breast cancer and genetic BRCA1/2 testing in routine clinical practice: why, when and for whom? *Geburtshilfe Frauenheilkd*. (2023) 83:310–20. doi: 10.1055/a-1929-2629

86. Wallace AJ. New challenges for BRCA testing: a view from the diagnostic laboratory. *Eur J Hum Genet*. (2016) 24 Suppl 1:S10-8. doi: 10.1038/ejhg.2016.94

87. Harbeck N. Neoadjuvant and adjuvant treatment of patients with HER2-positive early breast cancer. *Breast.* (2022) 62 Suppl 1:S12-6. doi: 10.1016/j.breast.2022.01.006

88. Tamirisa N, Hunt KK. Neoadjuvant chemotherapy, endocrine therapy, and targeted therapy for breast cancer: ASCO guideline. *Ann Surg Oncol.* (2022) 29:1489–92. doi: 10.1245/s10434-021-11223-3

89. van den Ende NS, Nguyen AH, Jager A, Kok M, Debets R, van Deurzen CHM. Triple-negative breast cancer and predictive markers of response to neoadjuvant chemotherapy: A systematic review. *Int J Mol Sci.* (2023) 24. doi: 10.3390/ijms24032969

90. Farrar MC, Jacobs TF. Tamoxifen. Treasure Island (FL: StatPearls (2023).

91. Adams S, Gatti-Mays ME, Kalinsky K, Korde LA, Sharon E, Amiri-Kordestani I, et al. Current landscape of immunotherapy in breast cancer: A review. *JAMA Oncol.* (2019) 5:1205–14. doi: 10.1001/jamaoncol.2018.7147

92. Greenblatt K, Khaddour K. Trastuzumab. Treasure Island (FL: StatPearls (2023).

93. Erinjeri JP, Fong AJ, Kemeny NE, Brown KT, Getrajdman GI, Solomon SB. Timing of administration of bevacizumab chemotherapy affects wound healing after chest wall port placement. *Cancer*. (2011) 117:1296–301. doi: 10.1002/cncr.25573

94. Gerber B, von Minckwitz G, Eidtmann H, Rezai M, Fasching P, Tesch H, et al. Surgical outcome after neoadjuvant chemotherapy and bevacizumab: results from the geparQuinto study (GBG 44). Ann Surg Oncol. (2014) 21:2517–24. doi: 10.1245/ s10434-014-3606-9

95. Fu MR. Breast cancer-related lymphedema: Symptoms, diagnosis, risk reduction, and management. World J Clin Oncol. (2014) 5:241–7. doi: 10.5306/wjco.v5.i3.241

96. Emens LA, Adams S, Cimino-Mathews A, Disis ML, Gatti-Mays ME, Ho AY, et al. Society for Immunotherapy of Cancer (SITC) clinical practice guideline on immunotherapy for the treatment of breast cancer. *J Immunother Cancer*. (2021) 9(8).

97. Heistein JB, Acharya U, Mukkamalla SKR. Malignant Melanoma. Treasure Island (FL: StatPearls Publishing (2023).

98. Linares MA, Zakaria A, Nizran P. Skin cancer. Prim Care. (2015) 42:645-59. doi: 10.1016/j.pop.2015.07.006

99. Shalhout SZ, Emerick KS, Kaufman HL, Miller DM. Immunotherapy for nonmelanoma skin cancer. *Curr Oncol Rep.* (2021) 23:125. doi: 10.1007/s11912-021-01120-z

100. Gruber P, Zito PM. Skin Cancer. Treasure Island (FL: StatPearls (2023).

101. Glazer AM, Winkelmann RR, Farberg AS, Rigel DS. Analysis of trends in US melanoma incidence and mortality. *JAMA Dermatol.* (2017) 153:225–6. doi: 10.1001/jamadermatol.2016.4512

102. Rigel DS. Trends in dermatology: melanoma incidence. Arch Dermatol. (2010) 146:318. doi: 10.1001/archdermatol.2009.379

103. Gordon R. Skin cancer: an overview of epidemiology and risk factors. Semin Oncol Nurs. (2013) 29:160–9. doi: 10.1016/j.soncn.2013.06.002

104. D'Orazio J, Jarrett S, Amaro-Ortiz A, Scott T. UV radiation and the skin. Int J Mol Sci. (2013) 14:12222–48. doi: 10.3390/ijms140612222

105. Berger MF, Levin JZ, Vijayendran K, Sivachenko A, Adiconis X, Maguire J, et al. Integrative analysis of the melanoma transcriptome. *Genome Res.* (2010) 20:413–27. doi: 10.1101/gr.103697.109

106. Hodis E, Watson , Kryukov GV, Arold ST, Imielinski M, Theurillat JP, et al. A landscape of driver mutations in melanoma. *Cell.* (2012) 150:251-63. doi: 10.1016/j.cell.2012.06.024

107. Conforti C, Zalaudek I. Epidemiology and risk factors of melanoma: A review. Dermatol Pract Concept. (2021) 11:e2021161S. doi: 10.5826/dpc.11S1a161S

108. Toussi A, Mans N, Welborn J, Kiuru M. Germline mutations predisposing to melanoma. *J Cutan Pathol.* (2020) 47:606–16. doi: 10.1111/cup.13689

109. Soura E, Eliades PJ, Shannon K, Stratigos AJ, Tsao H. Hereditary melanoma: Update on syndromes and management: Genetics of familial atypical multiple mole melanoma syndrome. *J Am Acad Dermatol.* (2016) 74:395–407. doi: 10.1016/ j.jaad.2015.08.038

110. Reddy BY, Miller DM, Tsao H. Somatic driver mutations in melanoma. *Cancer*. (2017) 123:2104–17. doi: 10.1002/cncr.30593

111. Ticha I, Hejny J, Michalkova R, Kodet O, Krkavcova E, Hajkova N, et al. A comprehensive evaluation of pathogenic mutations in primary cutaneous melanomas, including the identification of novel loss-of-function variants. *Sci Rep.* (2019) 9:17050. doi: 10.1038/s41598-019-53636-x

112. Davis LE, Shalin SC, Tackett AJ. Current state of melanoma diagnosis and treatment. *Cancer Biol Ther.* (2019) 20:1366–79. doi: 10.1080/15384047.2019.1640032

113. Sandru A, Voinea S, Panaitescu E, Blidaru A. Survival rates of patients with metastatic Malignant melanoma. J Med Life. (2014) 7:572–6.

114. Larkin J, Chiarion-Sileni V, Gonzalez R, Grob JJ, Rutkowski P, Lao CD, et al. Five-year survival with combined nivolumab and ipilimumab in advanced melanoma. *N Engl J Med.* (2019) 381:1535–46. doi: 10.1056/NEJMoa1910836

115. Tawbi HA, Schadendorf D, Lipson EJ, Ascierto PA, Matamala L, Castillo Gutierrez E, et al. Relatlimab and nivolumab versus nivolumab in untreated advanced melanoma. *N Engl J Med.* (2022) 386:24–34. doi: 10.1056/NEJMoa2109970

116. Triebel F, Jitsukawa S, Baixeras E, Roman-Roman S, Genevee C, Viegas-Pequignot E, et al. LAG-3, a novel lymphocyte activation gene closely related to CD4. *J Exp Med.* (1990) 171:1393–405. doi: 10.1084/jem.171.5.1393

117. Huuhtanen J, Kasanen H, Peltola K, Lonnberg T, Glumoff V, Bruck O, et al. Single-cell characterization of anti-LAG-3 and anti-PD-1 combination treatment in patients with melanoma. *J Clin Invest.* (2023) 133. doi: 10.1172/JCI164809

118. Byrne EH, Fisher DE. Immune and molecular correlates in melanoma treated with immune checkpoint blockade. *Cancer*. (2017) 123:2143–53. doi: 10.1002/ cncr.30444

119. Ascierto PA, Simeone E, Sileni VC, Pigozzo J, Maio M, Altomonte M, et al. Clinical experience with ipilimumab 3 mg/kg: real-world efficacy and safety data from an expanded access programme cohort. *J Transl Med.* (2014) 12:116. doi: 10.1186/1479-5876-12-116

120. Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, et al. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med.* (2010) 363:711–23. doi: 10.1056/NEJMx100063

121. Weber JS, Kahler KC, Hauschild A. Management of immune-related adverse events and kinetics of response with ipilimumab. *J Clin Oncol.* (2012) 30:2691–7. doi: 10.1200/JCO.2012.41.6750

122. Graziani G, Tentori L, Navarra P. Ipilimumab: a novel immunostimulatory monoclonal antibody for the treatment of cancer. *Pharmacol Res.* (2012) 65:9–22. doi: 10.1016/j.phrs.2011.09.002

123. Bhandaru M, Rotte A. Monoclonal antibodies for the treatment of melanoma: present and future strategies. *Methods Mol Biol 2019*. (1904) p:83–108.

124. Buchbinder EI, Desai A. CTLA-4 and PD-1 pathways: similarities, differences, and implications of their inhibition. *Am J Clin Oncol.* (2016) 39:98–106. doi: 10.1097/COC.00000000000239

125. McGee MC, Zhang T, Magazine N, Islam R, Carossino M, Huang W. PD-1 and ICOS counter-regulate tissue resident regulatory T cell development and IL-10 production during flu. *Front Immunol.* (2022) 13:984476. doi: 10.3389/fimmu.2022.984476

126. Anderson AC, Joller N, Kuchroo VK. Lag-3, tim-3, and TIGIT: co-inhibitory receptors with specialized functions in immune regulation. *Immunity*. (2016) 44:989–1004. doi: 10.1016/j.immuni.2016.05.001

127. Ascierto PA, Kirkwood JM, Grob JJ, Simeone E, Grimaldi AM, Maio M, et al. The role of BRAF V600 mutation in melanoma. *J Transl Med.* (2012) 10:85. doi: 10.1186/1479-5876-10-85

128. Eroglu Z, Ribas A. Combination therapy with BRAF and MEK inhibitors for melanoma: latest evidence and place in therapy. *Ther Adv Med Oncol.* (2016) 8:48–56. doi: 10.1177/1758834015616934

129. Guo YJ, Pan WW, Liu SB, Shen ZF, Xu Y, Hu LL. ERK/MAPK signalling pathway and tumorigenesis. *Exp Ther Med.* (2020) 19:1997–2007. doi: 10.3892/etm

130. Wagle N, Van Allen EM, Treacy DJ, Frederick DT, Cooper ZA, Taylor-Weiner A, et al. MAP kinase pathway alterations in BRAF-mutant melanoma patients with acquired resistance to combined RAF/MEK inhibition. *Cancer Discovery*. (2014) 4:61–8. doi: 10.1158/2159-8290.CD-13-0631

131. Lorentzen HF. Targeted therapy for Malignant melanoma. Curr Opin Pharmacol. (2019) 46:116-21. doi: 10.1016/j.coph.2019.05.010

132. Gonzalez-Del Pino GL, Li K, Park E, Schmoker AM, Ha BH, Eck MJ. Allosteric MEK inhibitors act on BRAF/MEK complexes to block MEK activation. *Proc Natl Acad Sci U.S.A.* (2021) 118. doi: 10.1073/pnas.2107207118

133. Davies H, Bignell GR, Cox C, Stephens P, Edkins S, Clegg S, et al. Mutations of the BRAF gene in human cancer. *Nature*. (2002) 417:949–54.

134. Young K, Minchom A, Larkin J. BRIM-1, -2 and -3 trials: improved survival with vemurafenib in metastatic melanoma patients with a BRAF(V600E) mutation. *Future Oncol.* (2012) 8:499–507. doi: 10.2217/fon.12.43

135. Chapman PB, Hauschild A, Robert C, Haanen JB, Ascierto P, Larkin J, et al. Improved survival with vemurafenib in melanoma with BRAF V600E mutation. *N Engl J Med.* (2011) 364:2507–16. doi: 10.1056/NEJMoa1103782

136. Flaherty KT, Puzanov I, Kim KB, Ribas A, McArthur GA, Sosman JA, et al. Inhibition of mutated, activated BRAF in metastatic melanoma. *N Engl J Med.* (2010) 363:809–19. doi: 10.1056/NEJMoa1002011

137. Sosman JA, Kim KB, Schuchter L, Gonzalez R, Pavlick AC, Weber JS, et al. Survival in BRAF V600-mutant advanced melanoma treated with vemurafenib. *N Engl J Med.* (2012) 366:707–14.

138. Dossett LA, Kudchadkar RR, Zager JS. BRAF and MEK inhibition in melanoma. Expert Opin Drug Saf. (2015) 14:559–70. doi: 10.1517/14740338.2015.1011618

139. Kim KB, Kefford R, Pavlick AC, Infante JR, Ribas A, Sosman JA, et al. Phase II study of the MEK1/MEK2 inhibitor Trametinib in patients with metastatic BRAFmutant cutaneous melanoma previously treated with or without a BRAF inhibitor. *J Clin Oncol.* (2013) 31:482–9. doi: 10.1200/JCO.2012.43.5966

140. Pavri SN, Clune J, Ariyan S, Narayan D. Malignant melanoma: beyond the basics. *Plast Reconstr Surg.* (2016) 138:330e–40e. doi: 10.1097/PRS.000000000020367

141. Pawlik L, Morgenroth S, Dummer R. Recent progress in the diagnosis and treatment of melanoma and other skin cancers. *Cancers (Basel)*. (2023) 15. doi: 10.3390/cancers15061824

142. Seth R, Messersmith H, Funchain PSystemic Therapy for Melanoma Guideline Expert. Systemic therapy for melanoma: ASCO guideline rapid recommendation update. *J Clin Oncol.* (2022) 40:2375-7. doi: 10.1200/JCO.22.00944

143. Voltan K, Baptista AM, Etchebehere M. Extremities soft tissue sarcomas, more common and as dangerous as bone sarcomas. *Rev Bras Ortop (Sao Paulo).* (2021) 56:419–24.

144. Popovich JR, Kashyap S, Gasalberti DP, Cassaro S. Sarcoma. Treasure Island (FL: StatPearls (2023).

145. Singhi EK, Moore DC, Muslimani A. Metastatic soft tissue sarcomas: A review of treatment and new pharmacotherapies. PT. (2018) 43:410–29.

146. Chen C, Dorado Garcia H, Scheer M, Henssen AG. Current and future treatment strategies for rhabdomyosarcoma. *Front Oncol.* (2019) 9:1458. doi: 10.3389/fonc.2019.01458

147. Zarrabi A, Perrin D, Kavoosi M, Sommer M, Sezen S, Mehrbod P, et al. Rhabdomyosarcoma: current therapy, challenges, and future approaches to treatment strategies. *Cancers (Basel).* (2023) 15. doi: 10.3390/cancers15215269

148. Chung YH, Jang JW, Cho JY. Soft tissue sarcoma in neurofibromatosis type 1: A rare case of Malignant peripheral nerve sheath tumor of the skin. *Arch Plast Surg.* (2020) 47:92–6. doi: 10.5999/aps.01529

149. Li FP, Fraumeni JF Jr. Soft-tissue sarcomas, breast cancer, and other neoplasms. A familial syndrome? *Ann Intern Med.* (1969) 71:747–52. doi: 10.7326/0003-4819-71-4-747

150. Burningham Z, Hashibe M, Spector L, Schiffman JD. The epidemiology of sarcoma. Clin Sarcoma Res. (2012) 2:14. doi: 10.1186/2045-3329-2-14

151. Malkin D, Li FP, Strong LC, Fraumeni JF Jr, Nelson CE, Kim DH, et al. Germ line p53 mutations in a familial syndrome of breast cancer, sarcomas, and other neoplasms. *Science.* (1990) 250:1233–8. doi: 10.1126/science.1978757

152. Lahat G, Lazar A, Lev D. Sarcoma epidemiology and etiology: potential environmental and genetic factors. *Surg Clin North Am.* (2008) 88:451-81. doi: 10.1016/j.suc.2008.03.006

153. Gamboa AC, Gronchi A, Cardona K. Soft-tissue sarcoma in adults: An update on the current state of histiotype-specific management in an era of personalized medicine. *CA Cancer J Clin.* (2020) 70:200–29. doi: 10.3322/caac.21605

154. Miwa S, Yamamoto N, Hayashi K, Takeuchi A, Igarashi K, Tsuchiya H. Recent advances and challenges in the treatment of rhabdomyosarcoma. *Cancers (Basel).* (2020) 12. doi: 10.3390/cancers12071758

155. Suarez-Kelly LP, Baldi GG, Gronchi A. Pharmacotherapy for liposarcoma: current state of the art and emerging systemic treatments. *Expert Opin Pharmacother*. (2019) 20:1503–15. doi: 10.1080/14656566.2019.1618271

156. Crago AM, Dickson MA. Liposarcoma: multimodality management and future targeted therapies. Surg Oncol Clin N Am. (2016) 25:761–73. doi: 10.1016/j.soc.2016.05.007

157. Thirasastr P, Somaiah N. Overview of systemic therapy options in liposarcoma, with a focus on the activity of selinexor, a selective inhibitor of nuclear export in dedifferentiated liposarcoma. *Ther Adv Med Oncol.* (2022) 14:17588359221081073. doi: 10.1177/17588359221081073

158. Mangla A, Yadav U. Leiomyosarcoma. Treasure Island (FL: StatPearls (2023).

159. Florou V, Nascimento AG, Gulia A, de Lima Lopes G Jr. Global health perspective in sarcomas and other rare cancers. *Am Soc Clin Oncol Educ Book*. (2018) 38:916–24. doi: 10.1200/EDBK_200589

160. Italiano A, Mathoulin-Pelissier S, Cesne AL, Terrier P, Bonvalot S, Collin F, et al. Trends in survival for patients with metastatic soft-tissue sarcoma. *Cancer*. (2011) 117:1049–54. doi: 10.1002/cncr.25538

161. Polito L, Calafato G, Bortolotti M, Chiarelli Olivari C, Maiello S, et al. Antibody conjugates for sarcoma therapy: how far along are we? *Biomedicines*. (2021) 9. doi: 10.3390/biomedicines9080978

162. Okuno SH, Maran A, Robinson SI. Olaratumab for the treatment of advanced soft tissue sarcoma. *Expert Rev Anticancer Ther.* (2017) 17:883–7. doi: 10.1080/14737140.2017.1374857

163. Johnson-Arbor K, Dubey R. Doxorubicin. Treasure Island (FL: StatPearls (2023).

164. Bou Zerdan M, Bidikian AH, Alameh I, Nakib CE, Assi HI. Olaratumab's failure in soft tissue sarcoma. *Rare Tumors*. (2021) 13:20363613211034115. doi: 10.1177/20363613211034115

165. Tap WD, Wagner AJ, Schoffski P, Martin-Broto J, Krarup-Hansen A, Ganjoo KN, et al. Effect of doxorubicin plus olaratumab vs doxorubicin plus placebo on survival in patients with advanced soft tissue sarcomas: the ANNOUNCE randomized clinical trial. *JAMA*. (2020) 323:1266–76. doi: 10.1001/jama.2020.1707

166. Ahmed M, Cheng M, Zhao Q, Goldgur Y, Cheal SM, Guo HF, et al. Humanized affinity-matured monoclonal antibody 8H9 has potent antitumor activity and binds to FG loop of tumor antigen B7-H3. *J Biol Chem.* (2015) 290:30018–29. doi: 10.1074/ jbc.M115.679852

167. Zhou WT, Jin WL. B7-H3/CD276: an emerging cancer immunotherapy. Front Immunol. (2021) 12:701006.

168. Modak S, Guo HF, Humm JL, Smith-Jones PM, La rson SM, Cheung NK. Radioimmunotargeting of human rhabdomyosarcoma using monoclonal antibody 8H9. *Cancer Biother Radiopharm.* (2005) 20:534–46. doi: 10.1089/cbr.2005.20.534

169. Birdi HK, Jirovec A, Cortes-Kaplan S, Werier J, Nessim C, Diallo JS, et al. Immunotherapy for sarcomas: new frontiers and unveiled opportunities. *J Immunother Cancer*. (2021) 9. doi: 10.1136/jitc-2020-001580

170. Pollack SM, Redman MW, Baker KK, Wagner MJ, Schroeder BA, Loggers ET, et al. Assessment of doxorubicin and pembrolizumab in patients with advanced anthracycline-naive sarcoma: A phase 1/2 nonrandomized clinical trial. *JAMA Oncol.* (2020) 6:1778–82. doi: 10.1001/jamaoncol.2020.3689

171. Zhou M, Bui N, Bolleddu S, Lohman M, Becker HC, Ganjoo K. Nivolumab plus ipilimumab for soft tissue sarcoma: a single institution retrospective review. *Immunotherapy.* (2020) 12:1303–12.

172. Chen Y, Liu X, Liu J, Liang D, Zhao M, Yu W, et al. Nivolumab plus ipilimumab versus nivolumab in individuals with treatment-naive programmed death-ligand 1 positive metastatic soft tissue sarcomas: a multicentre retrospective study. *BMC Cancer*. (2021) 21:108. doi: 10.1186/s12885-021-07843-3

173. Lazcano R, Barreto CM, Salazar R, Carapeto F, Traweek RS, Leung CH, et al. The immune landscape of undifferentiated pleomorphic sarcoma. *Front Oncol.* (2022) 12:1008484.

174. Roland CL, Nassif Haddad EF, Keung EZ, Wang WL, Lazar AJ, Lin H, et al. A randomized, non-comparative phase 2 study of neoadjuvant immune-checkpoint blockade in retroperitoneal dedifferentiated liposarcoma and extremity/truncal undifferentiated pleomorphic sarcoma. *Nat Cancer.* (2024).

175. Eckschlager T, Pich J, Stiborova M, Hrabeta J. Histone deacetylase inhibitors as anticancer drugs. Int J Mol Sci. (2017) 18. doi: 10.3390/ijms18071414

176. Roberts JL, Booth L, Poklepovic A, Dent P. Axitinib and HDAC inhibitors interact to kill sarcoma cells. *Front Oncol.* (2021) 11:723966. doi: 10.3389/ fonc.2021.723966

177. Tavallai S, Hamed HA, Grant S, Poklepovic A, Dent P. Pazopanib and HDAC inhibitors interact to kill sarcoma cells. *Cancer Biol Ther.* (2014) 15:578–85. doi: 10.4161/cbt.28163

178. McCormack PL. Pazopanib: a review of its use in the management of advanced renal cell carcinoma. *Drugs.* (2014) 74:1111-25. doi: 10.1007/s40265-014-0243-3

179. Gross-Goupil M, Francois L, Quivy A, Ravaud A. Axitinib: a review of its safety and efficacy in the treatment of adults with advanced renal cell carcinoma. *Clin Med Insights Oncol.* (2013) 7:269–77. doi: 10.4137/CMO.S10594

180. Wilky BA, Trucco MM, Subhawong TK, Florou V, Park W, Kwon D, et al. Axitinib plus pembrolizumab in patients with advanced sarcomas including alveolar soft-part sarcoma: a single-centre, single-arm, phase 2 trial. *Lancet Oncol.* (2019) 20:837–48. doi: 10.1016/S1470-2045(19)30153-6

181. Society AC. Immunotherapy for soft tissue sarcoma(2022). Available at: https://www.cancer.org/cancer/types/soft-tissue-sarcoma/treating/immunotherapy.html.

182. Institute CR. Immunotherapy for sarcoma. Available online at: https://www. cancerresearch.org/cancer-types/sarcoma.

183. Sigl V, Jones LP, Penninger JM. RANKL/RANK: from bone loss to the prevention of breast cancer. *Open Biol.* (2016) 6. doi: 10.1098/rsob.160230

184. Loibl S, Furlanetto J. Integrating CDK4/6 inhibitors in the treatment of patients with early breast cancer. *Breast.* (2022) 62 Suppl 1:S70-9. doi: 10.1016/j.breast.2021.12.008

185. Slamon DJ, Neven P, Chia S, Fasching PA, De Laurentiis M, Im SA, et al. Overall survival with ribociclib plus fulvestrant in advanced breast cancer. *N Engl J Med.* (2020) 382:514–24. doi: 10.1056/NEJMoa1911149

186. Sledge GW Jr., Toi M, Neven P, Sohn J, Inoue K, Pivot X, et al. MONARCH 2: abemaciclib in combination with fulvestrant in women with HR+/HER2- advanced breast cancer who had progressed while receiving endocrine therapy. *J Clin Oncol.* (2017) 35:2875–84. doi: 10.1200/JCO.2017.73.7585

187. Finn RS, Boer K, Bondarenko I, Patel R, Pinter T, Schmidt M, et al. Overall survival results from the randomized phase 2 study of palbociclib in combination with letrozole versus letrozole alone for first-line treatment of ER+/HER2- advanced breast cancer (PALOMA-1, TRIO-18). *Breast Cancer Res Treat.* (2020) 183:419–28. doi: 10.1007/s10549-020-05755-7

188. Finn RS, Cristofanilli M, Ettl J, Gelmon KA, Colleoni M, Giorgetti C, et al. Treatment effect of palbociclib plus endocrine therapy by prognostic and intrinsic subtype and biomarker analysis in patients with bone-only disease: a joint analysis of PALOMA-2 and PALOMA-3 clinical trials. *Breast Cancer Res Treat.* (2020) 184 (1):23–35.

Glossary

5-FU	(5-)fluorouracil	
ARMS	alveolar rhabdomyosarcoma	
BAP1	BRAC1 associated protein	
BAP1- TPDS	BAP1 tumor predispositions syndrome	
BC	breast cancer	
BCC	basal cell carcinoma	
BRCA1/2	breast cancer gene 1/2	
BRAF	v-raf murine sarcoma viral oncogene homolog B1	
BS	bone sarcomas	
CDK	cyclin-dependent kinase	
CDKN2A	cyclin-dependent kinase inhibitor	
CPS	Combined Positive Score	
CTLA-4	cytotoxic T-lymphocyte antigen 4	
CXCR4	CXC chemokine receptor type 4	
DDLPS	dedifferentiated liposarcoma	
dMMR	DNA mismatch repair deficiency	
EBV	epstein-barr virus, human herpesvirus 4	
EGFR	epidermal growth factor receptor	
ER	estrogen receptor	
ERK	extracellular signal-regulated kinase	
ERMS	embryonal rhabdomyosarcoma	
FCS	five-year cumulative survival	
FDA	United States Food and Drug Administration	
FGFR	fibroblast growth factor receptor	
HDAC	histon deacetylase	
HER-2	human epidermal growth factor receptor 2	
HNC	head and neck cancer	
HNSCC	head and neck squamous cell carcinoma	
HPV	human papilloma virus	
ICI	immune checkpoint inhibitors	
IgG4	humanized mononuclear immunoglobulin G4	
LAG-3	lymphocyte activation gene 3	
LFS	Li-Fraumeni syndrome	
LMS	Leiomyosarcoma	
LS	Liposarcoma	
mAB	monoclonal antibodies	
МАРК	mitogen-activated protein kinase	
MEK	mitogen-activated protein kinase kinase	

Continued MHC-I major histocompatibility complex I MITF microphthalmia-associated transcription factor malignant melanoma MM MRCLS myxoid/round-cell liposarcoma MSI microsatellite instability mTOR mammalian target of rapamycin NF1 neurofibromatosis type 1 NK cells natural killer cells PALB2 partner and localizer of BRCA2 gene poly-ADP-ribose polymerase PARP PD-1 programmed cell death protein 1 PD-L1 programmed cell death protein ligand 1 PDGFR platelet-derived growth factor receptor PFS progession-free survival PI3K phosphatidylinositol-3-kinase PI3KCA phosphatidylinositol-4,5-biphosphaste 3-kinase catalytic subunit alpha PR progesterone receptor PTEN phosphate and tensin homolog gene R/M recurrent or metastatic RAC1 ras-related C3 botulinum toxin substrate 1 RANK receptor activator of nuclear factor kappa-B RANKL RANK ligand RMS Rhabdomyosarcoma SC skin cancer SCC squamous cell carcinoma ssRMS spinde cell/sclerosing rhabdomyosarcoma STS soft tissue sarcomas TIGIT T cell immunoreceptor with Ig and ITIM domains TIM-3 T cell immunoglobulin and mucin domain 3 TKI tyrosine kinase inhibitor TMB tumor mutational burden TNBC triple negative breast cancer TNFα tumor necrosis factor alpha $\mathrm{T}_{\mathrm{regs}}$ regulatory T cells TP53 tumor protein p53 gene VEGF vascular endothelial growth factor VEGFR vascular endothelial growth factor receptor WDLPS well-differentiated liposarcoma

(Continued)