



# *Article* **Differentiating Well-Differentiated from Poorly-Differentiated HCC: The Potential and the Limitation of Gd-EOB-DTPA in the Presence of Liver Cirrhosis**

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**Abstract:** This study uses magnetic resonance imaging (MRI) to investigate the potential of the hepatospecific contrast agent gadolinium ethoxybenzyl-diethylenetriaminepentaacetic acid (Gd-EOB-DTPA) in distinguishing G1- from G2/G3-differentiated hepatocellular carcinoma (HCC). Our approach involved analyzing the dynamic behavior of the contrast agent in different phases of imaging by signal intensity (SI) and lesion contrast (C), to surrounding liver parenchyma, and comparing it across distinct groups of patients differentiated based on the histopathological grading of their HCC lesions and the presence of liver cirrhosis. Our results highlighted a significant contrast between well- and poorlydifferentiated lesions regarding the lesion contrast in the arterial and late arterial phases. Furthermore, the hepatobiliary phase showed limited diagnostic value in cirrhotic liver parenchyma due to altered pharmacokinetics. Ultimately, our findings underscore the potential of Gd-EOB-DTPA-enhanced MRI as a tool for improving preoperative diagnosis and treatment selection for HCC while emphasizing the need for continued research to overcome the diagnostic complexities posed by the disease.

**Keywords:** hepatocellular carcinoma (HCC); gadolinium ethoxybenzyl-diethylenetriaminepentaacetic acid (Gd-EOB-DTPA); magnetic resonance imaging (MRI); liver cirrhosis; tumor differentiation

#### **1. Introduction**

Hepatocellular carcinoma (HCC) presents a substantial global health challenge as the predominant form of primary liver cancer. Particularly prevalent in individuals with pre-existing cirrhosis, it ranks as the fifth most common malignancy in males and ninth in females with a rising incidence  $[1-4]$  $[1-4]$ .

Given its asymptomatic nature in the early stages, HCC often eludes early detection, resulting in a diagnosis at advanced stages when curative therapy becomes infeasible, mainly as prognosis is largely dependent on the tumor stage and metastasis [\[5\]](#page-7-2). Therefore, developing robust and efficient strategies for early detection in at-risk populations is paramount in HCC management. HCC mostly occurs in patients with severe liver fibrosis or cirrhosis mainly due to infections with hepatitis B or hepatic C viruses as well as chronic alcohol abuse [\[6\]](#page-7-3). However, HCC also develops in patients with non-alcoholic fatty liver disease, hemochromatosis, or alpha-1 antitrypsin deficiency [\[4\]](#page-7-1).

Current practices for early detection primarily involve semiannual ultrasound examinations for high-risk individuals. Suspicious findings from these examinations typically



**Citation:** Goetz, A.; Verloh, N.; Utpatel, K.; Fellner, C.; Rennert, J.; Einspieler, I.; Doppler, M.; Luerken, L.; Alizadeh, L.S.; Uller, W.; et al. Differentiating Well-Differentiated from Poorly-Differentiated HCC: The Potential and the Limitation of Gd-EOB-DTPA in the Presence of Liver Cirrhosis. *Diagnostics* **2024**, *14*, 1676. [https://doi.org/10.3390/](https://doi.org/10.3390/diagnostics14151676) [diagnostics14151676](https://doi.org/10.3390/diagnostics14151676)

Academic Editor: Søren Møller

Received: 2 July 2024 Revised: 30 July 2024 Accepted: 31 July 2024 Published: 2 August 2024



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lead to further investigation using computed tomography (CT) or magnetic resonance imaging (MRI) [\[7,](#page-8-0)[8\]](#page-8-1). While these methods have proven somewhat effective, there is ongoing debate regarding their relative efficiencies and potential for optimization.

Regarding the diagnostic pathway for uncertain liver lesions, lesions suspicious for malignancy and smaller than 1 cm should undergo sonographic follow-up in three months. For lesions exceeding 1 cm, further CT or MRI diagnostic imaging is recommended. Several studies have asserted the superiority of contrast-enhanced MRI over dynamic contrastenhanced CT for detecting and differentiating liver lesions [\[9](#page-8-2)[,10\]](#page-8-3). Upon arterial hypervascularization and portal venous washout observation, HCC diagnosis is considered validated [\[11\]](#page-8-4). In unclear cases, a biopsy is recommended [\[12\]](#page-8-5).

Several studies have demonstrated that contrast-enhanced MRI is more sensitive in detecting HCC compared to dynamic contrast-enhanced CT. This increased sensitivity is likely due to the superior contrast delineation between the lesion and liver, as well as a more nuanced presentation of different tissue properties [\[9,](#page-8-2)[10,](#page-8-3)[13\]](#page-8-6).

In clinical practice, liver MRI using the hepatocyte-specific contrast agent Gd-EOB-DTPA has become essential for the evaluation of malignant liver lesions.

By using this hepatocyte-specific contrast agent, diagnostic accuracy could be improved by performing an additional hepatobiliary late phase 20 min after contrast agent application [\[14\]](#page-8-7).

The uptake of Gd-EOB-DTPA relies on functioning hepatocytes through organic anion transporters (OATP1B1/B3) and is excreted via the biliary tract through multidrug resistance-associated protein 2 (MRP2) [\[15](#page-8-8)[,16\]](#page-8-9).

A typical assessment of HCC involves the qualitative evaluation of signal intensity and the relative enhancement ratios, comparing the Gd-EOB-DTPA uptake in liver lesions to that in the surrounding liver parenchyma. However, the uptake of Gd-EOB-DTPA is also influenced by the presence and severity of cirrhosis [\[17\]](#page-8-10), showing less Gd-EOB-DTPA uptake in cirrhotically remodeled liver parenchyma. Additionally, as the degree of differentiation of HCC increases, the number of hepatocytes expressing OATP1B1/B3 decreases and MRP2 increases, therefore leading to a reduced accumulation of Gd-EOB-DTPA compared to normal liver parenchyma [\[16,](#page-8-9)[18\]](#page-8-11). As HCCs commonly occur in cirrhotic livers with impaired Gd-EOB-DTPA uptake, this assessment can be challenging.

Certain studies have also identified distinct enhancement patterns that could potentially correlate with different HCC stages [\[16](#page-8-9)[,19](#page-8-12)[–22\]](#page-8-13). However, there remains considerable ambiguity and controversy regarding whether these patterns reliably vary according to tumor grade.

The role of liver cirrhosis in influencing the diagnostic accuracy of these imaging techniques remains contentious and underexplored.

In this context, the primary objective of this study is to investigate whether specific enhancement patterns correspond to different grades of HCC. We also seek to examine the impact of liver cirrhosis on the diagnostic accuracy of these imaging strategies. Through this dual-pronged investigation, we hope to clarify some of the existing ambiguities in the field and contribute to the ongoing efforts to refine imaging strategies for early HCC detection.

#### **2. Materials and Methods**

#### *2.1. Data Collection and Inclusion Criteria*

This retrospective study was approved by the local institutional review board, ensuring that all the regulations and guidelines were followed. Our study incorporated patients who had undertaken Gd-EOB-DTPA-enhanced MRI of the liver before liver resection and had a histopathological confirmed, untreated, HCC within three months from the examination.

The initial data set comprised 59 resected HCC lesions derived from 52 patients. Exclusion criteria were respiratory artifacts (*n* = 4), incomplete detection of contrast medium phases  $(n = 3)$ , infiltrative growth of the tumor  $(n = 4)$ , or pre-treatment  $(n = 1)$ . Thus, we inspected 47 lesions from 40 patients for their dynamic contrast behavior. Based on differentiation grade, 18 were classified as G1, 23 as G2, and 6 lesions as G3.

#### *2.2. Imaging*

Liver MRI scans were procured using a clinical whole-body 3T system (MAGNETOM Skyra, Siemens Healthcare, Erlangen, Germany), using a composite of body-spine array coil elements (comprising an 18-channel body matrix coil and a 32-channel spine matrix coil) for signal reception. The hepatospecific contrast agent was Gd-EOB-DTPA (Primovist<sup>®</sup>, Bayer Vital GmbH, Leverkusen, Germany). Each patient was administered a dose of Gd-EOB-DTPA, adapted according to their body weight (0.025 mmol/kg body weight). The administration was performed as a bolus injection at a flow rate of 1 mL/s, followed by a 20 mL NaCL flush. A T1-weighted volume interpolated breath-hold examination (VIBE) sequence with fat suppression was used for the signal intensity measurement. The specifications of this sequence are repetition time (TR) of 3.09 ms, echo time (TE) of 1.16 ms, a flip angle of 10°, parallel imaging factor of 2, 64 slices, a reconstructed voxel size of  $1.3 \times 1.3 \times 3.0$  mm, and a measured voxel size of  $1.7 \times 1.3 \times 4.5$  mm. This sequence spanned the entire liver and was captured in a single breath-hold before contrast injection (plain), during the arterial phase (AP) at 10 s, in the late arterial phase (LAP) at 40 s, in the portal venous phase (PVP) at 75 s, and in the hepatobiliary phase (HBP) at 20 min. The acquisition time for each VIBE sequence was 14 s.

#### *2.3. Signal Intensity Measurement*

Signal intensity (SI) was measured in liver lesions and adjacent liver tissue before and during various contrast medium phases, namely arterial, late arterial, portal venous, and hepatobiliary phase. One region of interest (ROI) was manually placed around the tumor boundary on the slice with the greatest tumor extension. This was done individually for each contrast medium phase.

For measuring the signal intensity of liver tissue, a second circular ROI was manually placed in the same slice in the liver parenchyma while avoiding any additional lesions or vascular structures.

The lesion contrast (C) was calculated for each liver lesion relative to the surrounding liver tissue using the formula:

### $C = (SI_{Lesion} - SI_{Liver \text{ parenchyma}})/SI_{Liver \text{ parenchyma}}$

where during the respective contrast phases, this measure demonstrated the contrast agent uptake ratio of the liver lesions to the surrounding liver tissue. It was correlated with the respective histopathological differentiation grade relative to each contrast medium phase.

#### *2.4. Histopathological Analysis*

For diagnostic purposes, tissue specimens were gathered from standard therapeutic surgeries conducted in a five-year period. The samples were preserved in neutral buffered formalin, followed by paraffin embedding. Tissue sections, 4 µm thick, were prepared following a conventional protocol and stained using hematoxylin and eosin (HE). A senior pathologist carried out histological evaluations. The grading of HCC was ascertained following the criteria laid down by the World Health Organization (WHO 5th edition) [\[23\]](#page-8-14). Based on the severity of malignancy, the HCCs were classified into three groups: G1, signifying well differentiated; G2, moderately differentiated; and G3, poorly differentiated. The fibrosis grade was graded using the Ishak scoring system [\[24\]](#page-8-15). In total, 26 HCC lesions were associated with incomplete or complete cirrhotic liver remodeling (ISHAK scores 5 and 6), whereas 21 lesions were not underpinned by cirrhosis (ISHAK scores 1–4).

#### *2.5. Statistical Analysis*

Statistical analysis was conducted using IBM SPSS Statistics Version 26 (Chicago, IL, USA). The measured signal intensities' mean value and corresponding standard deviations were presented. Liver lesions were compared in terms of their degree of differentiation

concerning contrast uptake using the non-parametric Mann-Whitney U test for independent variables. Statistical significance was set at *p* values < 0.05.

#### **3. Results**

#### *3.1. Patient Demographics and Lesion Classification*

In this study, we analyzed 47 HCC lesions from 40 patients. Well-differentiated HCC, denoted as G1 (*n* = 18), and poorly-differentiated HCC, combining G2 (*n* = 23) and G3 (*n* = 6) grades, were considered for analysis. The average age of the patients at the time of the MRI examination was 68.2 years, ranging from 44 to 83 years. No significant differences in the patients' ages, body weights, heights, or BMIs were identified between the patients with normal liver function and those with liver cirrhosis. The majority of the patients were males (*n* = 35, 87.5%), while females constituted a minor proportion (*n* = 5, 12.5%). 26 HCC lesions (22 patients) were associated with precirrhotic or cirrhotic liver remodeling (ISHAK scores 5 and 6), whereas 21 lesions (18 patients) were not underpinned by cirrhosis (ISHAK scores [1](#page-3-0)–4). Table 1 provides a summary of the patient characteristics.



<span id="page-3-0"></span>**Table 1.** Patient characteristics (NCL, non-cirrhotic liver parenchyma; LC, liver cirrhosis).

#### *3.2. Qualitative Analysis of Signal Intensity*

The signal intensities and contrast of HCC lesions were systematically evaluated against the surrounding liver parenchyma across distinct contrast medium phases. Mean values of the signal intensities for individual contrast medium phases with the corresponding lesion contrast are demonstrated in Tables [2](#page-3-1) and [3.](#page-3-2)

<span id="page-3-1"></span>**Table 2.** Mean signal intensity (SI) by phase and differentiation grade.



<span id="page-3-2"></span>**Table 3.** Lesion contrast (C) by phase and differentiation grade.



In the plain phase, the lesions' average signal intensity (SI) was  $168.85 \pm 52.29$ , with a corresponding C of −0.09. When analyzed by differentiation grade, the G1 lesions had an SI of 169.00 ± 54.18 and an SI of 169.18 lesions recorded and SI of 169.18 and SI of 169.18 and SI of 169.18 and SI of 169.18 and SI SI of 169.00  $\pm$  54.18 and a C of −0.01, while the combined G2/G3 lesions recorded an SI of 468.76 ± 52.05 and a increase in SI was observed. The arterial phase, and in SI was observed. The arterial phase, and in SI was  $168.76 \pm 52.05$  and a C of −0.14. In the arterial phase, an increase in SI was observed. The corrections was  $251.68 \pm 73.86$ , with a C of 0.25. The well-differentiated G1.68. average SI for all lesions was  $251.68 \pm 73.86$ , with a C of 0.25. The well-differentiated G1 lesions showed an SI of 260.11  $\pm$  57.50 and a C of 0.40, whereas the combined G2/G3 lesions presented an SI of  $246.45 \pm 82.95$  and a C of 0.17. In the late arterial phase, the average SI for all lesions was  $266.91 \pm 68.06$ , with a C of  $-0.07$ . The G1 lesions showed an SI of  $283.83 \pm 63.13$  and a C of 0.03, while the G2/G3 lesions showed an SI of 256.41  $\pm$  69.94 and a C of 0.03, while the G2/G3 lesions showed an SI of 256.41  $\pm$  69.94 and  $288.88 \pm 0.018$  and a C of  $-0.02$ . In the portal venous phase, the average SI for all lesions was 265.15 ± 67.63, as with a C of  $-0.09$ . The G1 lesions had an SI of 267.78  $\pm$  61.65 and a C of  $-0.04$ , while the G2/G3 lesions recorded an SI of 263.52  $\pm$  72.10 and a C of −0.13. In the hepatobiliary phase, the average SI for all lesions dropped to 235.34 ± 70.95 with a C of −0.25. The G1 lesions average SI for all lesions dropped to 235.34 ± 70.95 with a C of −0.25. The G1 lesions showed an SI of 246.33  $\pm$  80.21 and a C of −0.19, whereas the G2/G3 lesions exhibited an SI of  $228.69 \pm 65.12$  and a C of −0.29. In the plain phase, the lesions internal internal internal internal internal intensity (SI) was 168.85  $\pm$ The corresponding C of  $\alpha$  is the GNU less of  $\alpha$  and analyzed by differentiation grade, the G1 less of  $\alpha$  less had an analyzed by differentiation grade, the G1 less had an analyzed by differentiation grade, the G1 le

Figure [1a](#page-4-0) depicts the absolute signal intensities of liver lesions in each contrast medium phase and Figure [1b](#page-4-0) displays the C of HCC compared to adjacent liver parenchyma. A significant difference emerges between the well-differentiated and the G2/G3 lesions A significant difference emerges between the well-differentiated and the G2/G3 lesions regarding the C in the arterial ( $p = 0.010$ ) and late arterial phases ( $p = 0.040$ ). However, no significant difference is noted in the subsequent phases. Both groups exhibit the typical HCC contrast behavior, characterized by arterial hypervascularization of the lesions and a washout of the contrast agent in the portal venous and hepatobiliary phases.

<span id="page-4-0"></span>

Figure 1. Contrast response of the liver lesions for the plain, arterial (AP), late atrial (LAP), portal venous (PVP), and hepatobiliary phase (HBP). (a) absolute signal intensities (b) contrast (C).  $*$ ,  $p = 0.040$ ; \*\*,  $p = 0.010$ .

## *3.3. Influence of Liver Cirrhosis 3.3. Influence of Liver Cirrhosis*

The contrasting behavior was further investigated for the presence of liver cirrhosis.<br>
The contrasting behavior was further investigated for the presence of liver cirrhosis. A total of 26 HCC lesions were associated with precirrhotic or cirrhotic liver remodeling  $T_{\text{GUT}}$ (ISHAK scores 5 and 6), whereas 21 lesions were not underpinned by cirrhosis (ISHAK) scores 1–4). For G1, the distribution was as follows: 8 lesions in patients without liver circle and 10 in patients with linear circle and  $\overline{G}$  and  $\overline{G}$  and  $\overline{G}$  and  $\overline{G}$  and  $\overline{G}$  and  $\overline{G}$  and  $\overline$ rhosis and 10 in patients with liver cirrhosis. For combined G2 and G3, the distribution was: 13 lesions in patients without liver cirrhosis and 16 in patients with liver cirrhosis.<br>Liver simbosis had distinct invaliations for Cd FOP DTPA watels into the cumsum cirrhosis and 10 in patients with liver cirrhosis. For combined G2 and G3, the distribution

Liver cirrhosis had distinct implications for Gd-EOB-DTPA uptake into the surrounding liver parenchyma and the respective C of the HCC lesions. Figure [2](#page-5-0) illustrates these impacts, showing that in a cirrhotically altered liver, there is a subtle change in C between the portal venous and hepatobiliary phase. In contrast, a more noticeable reduction in C is observed in healthy liver tissue. A significant difference was observed for G2/G3 tumors between non- cirrhotic and cirrhotic liver parenchyma (*p* = 0.001), whereas no significant difference was observed for G1 tumors.

<span id="page-5-0"></span>

Figure 2. Contrast of HCC lesions in relation to cirrhotic (red) and non-cirrhotic (black) liver parenchyma for the plain, arterial (AP), late atrial (LAP), portal venous (PVP), and hepatobiliary (HBP). (**a**) G1. (**b**) G2/G3. phase (HBP). (**a**) G1. (**b**) G2/G3. Figure 2. Contrast of the resions in relation to christian (lear) and hon-christian (plack) fiver

# *3.4. Image Examples 3.4. Image Examples 3.4. Image Examples*

difference was observed for G1 tumors. The G1 tumors of G1 tumors of G1 tumors.

cirrhotic liver parenchyma are shown in Figure [3.](#page-5-1) All HCC lesions show the typical arterial hypervascularization with washout in the portal venous phase and hypointensity in the hepatobiliary phase. The uptake of Gd-EOB-DTPA in the surrounding liver parenchyma  $\mu$  hepatobiliary phase. The uptake of Gd-EOB-DTPA in the uptake of God-EOB-DTPA in the surrounding liver parameters  $\mu$ differs between non-cirrhotic and cirrhotic liver parenchyma, thus affecting the contrast of HCC losions Examples of the dynamic contrast behavior of HCC lesions in non-cirrhotic and HCC lesions.

<span id="page-5-1"></span>

**Figure 3.** Comparison of well-differentiated and poorly-differentiated HCCs in the presence of liver cirrhosis (ISHAK Score 6) in T1 weighted VIBE sequences (phases as indicated): (**A**) well-differentiated HCC (G1) in liver fibrosis (ISHAK 2), (**B**) well-differentiated HCC (G1) in liver cirrhosis (ISHAK 6), (**C**) poorly-differentiated HCC (G2) in normal liver parenchyma (ISHAK 0), (**D**) poorly-differentiated HCC (G2) in in liver cirrhosis (ISHAK 6).

### **4. Discussion**

To the best of our knowledge, this is the first study to investigate the impact of liver function on Gd-EOB-DTPA for diagnosing HCC in relation to its grading.

Preoperative determination of the degree of differentiation of HCC is essential for choosing appropriate therapy [\[25–](#page-8-16)[28\]](#page-8-17). Various agents like atorvastatin [\[29\]](#page-8-18), cefazolin [\[30\]](#page-8-19), and chemotherapeutic agents such as methotrexate [\[31\]](#page-8-20), rifampicin [\[32\]](#page-9-0), paclitaxel, and docetaxel [\[33\]](#page-9-1) are also absorbed into cells via OATP 1B1 and 1B3. This suggests that tumor therapy effectiveness may depend on tumor differentiation degree as it correlates with these transporter proteins' expression.

According to current German guidelines, the treatment of HCC in a cirrhotic liver is liver transplantation, which also treats the underlying cirrhosis. However, due to organ shortage, there are strict regulations about the eligibility of liver transplantation such as the Milan criteria, in which patients are considered eligible for liver transplantation with one HCC lesion up to 5 cm or up to 3 HCC lesions between 1–3 cm (Mazzaferro 1996). Because these strict criteria may exclude potential patients who would benefit from liver transplantation, it has been suggested that the degree of differentiation should also play a role in the selection criteria for liver transplantation [\[25,](#page-8-16)[26\]](#page-8-21), as it has been shown that the histopathological degree of differentiation is a prognostic factor for the survival rate [\[27](#page-8-22)[,28\]](#page-8-17).

Due to this prognostic influence, patients with poorly-differentiated HCC are excluded from liver transplantation and treated palliatively in some centers [\[34](#page-9-2)[,35\]](#page-9-3). Therefore, determining the correct degree of differentiation of the presenting HCC plays a significant role.

The histopathological degree of differentiation is determined by biopsy, usually as part of the primary diagnosis, with the most common procedure being punch biopsy, which can be associated with complications such as bleeding, infection, or carryover of tumor tissue [\[36\]](#page-9-4). Pawlik et al. [\[37\]](#page-9-5) show a specificity of 92.5% and a sensitivity of only 34.6% concerning the determination of a low degree of differentiation (G3) so that a false-negative classification can frequently occur here.

Several studies have reported that HCCs' appearance on Gd-EOB-DTPA-enhanced MRI examinations vary based on their grading [\[38](#page-9-6)[–43\]](#page-9-7). However, several other studies contradict these results, asserting that Gd-EOB-DTPA uptake in HCC lesions does not correlate with differentiation degree [\[44](#page-9-8)[–47\]](#page-9-9). The regulation of Gd-EOB-DTPA's uptake and excretion is controlled by OATP 1B1/1B3 and MRP-2 [\[15\]](#page-8-8). OATPs are multispecific transporter proteins, with OATP 1B1 and 1B3 subtypes expressed explicitly in the liver [\[48\]](#page-9-10). Studies have demonstrated that advanced HCC lesions often exhibit reduced OATP 1B1 or 1B3 expression, while MRP2 expression remains stable or increases [\[16](#page-8-9)[,18\]](#page-8-11). These findings corroborate our results, which suggest decreased Gd-EOB-DTPA accumulation in poorlydifferentiated HCC lesions, potentially due to reduced Gd-EOB-DTPA uptake or increased biliary excretion rates. Frericks et al. [\[44\]](#page-9-8) and Schelhorn et al. [\[39\]](#page-9-11) showed no correlation between the difference in grade and the signaling of liver lesions. Still, histopathological grading was performed only on biopsy specimens and not completely resected liver tissue. In addition, the evaluation was based on signaling changes of the lesions compared with the surrounding liver parenchyma, without considering whether there was underlying cirrhosis and thus possibly impaired Gd-EOB- DTPA uptake. Tsuboyama et al. [\[46\]](#page-9-12). demonstrated overexpression of OATP—1B3 at all stages of differentiation, linking high Gd-EOB-DTPA accumulation of lesions to altered expression of MRP-2 and, thus, possibly, decreased excretion. They defined high Gd-EOB-DTPA enrichment as greater enrichment compared with surrounding liver parenchyma, independent of liver function. Since they listed only five lesions with high enhancement in their study, no conclusion could be drawn regarding the correlation between tumor grade and Gd-EOB-DTPA enhancement.

The presence of liver cirrhosis, one of the most critical risk factors for developing HCC, also influences Gd-EOB-DTPA uptake in the pre-damaged liver parenchyma [\[17\]](#page-8-10). Tamada et al. [\[49\]](#page-9-13) showed that the accumulation of Gd-EOB-DTPA in the liver parenchyma is significantly reduced by liver cirrhosis, especially in the Child C stage, most likely due to a reduced number of healthy hepatocytes or impaired contrast agent uptake into liver cells. Therefore, marked liver cirrhosis shows a variable appearance of the liver in contrast-enhanced MRI examinations so that the hepatobiliary phase in liver cirrhosis can sometimes only be assessed to a limited extent because the surrounding liver parenchyma

only accumulates suboptimally [\[50\]](#page-9-14). In MRI diagnostics, the signal behavior is generally considered qualitatively, and lesions are therefore assessed in hyper-, iso-, or hypointense, depending on the surrounding tissue. Similarly, the contrast of liver lesions used in this work is calculated using the signal intensity of the surrounding liver parenchyma. It is thus influenced by the condition of the surrounding liver parenchyma in case of liver fibrosis or cirrhosis.

In the case of impaired liver function, it is therefore not possible to clearly distinguish based on the lesion contrast whether it is due to reduced uptake of Gd-EOB-DTPA into the cirrhotically remodeled liver parenchyma or to washout of the liver lesions.

This study's primary limitation is the small number of HCC lesions included, and the rarity of G3 lesions limited their inclusion to only six [\[51\]](#page-9-15). HCC's high intratumoral heterogeneity also challenges pathologists and radiologists, as different degrees of differentiation may exist within one tumor [\[52\]](#page-9-16).

#### **5. Conclusions**

This study emphasizes the potential of using the hepatospecific contrast agent Gd-EOB-DTPA in differentiating well-differentiated from poorly-differentiated HCC during MRI examinations. By analyzing the dynamic contrast agent behavior, we could distinguish between different stages of HCC differentiation, thereby contributing to the precision of preoperative diagnosis and selection of suitable therapeutic strategies. Further research is needed to confirm the utility of combining radiologic and histopathologic features in grading HCC, which could lead to more accurate therapeutic decisions.

However, our findings also highlight a potentially limited additional diagnostic value of the late hepatobiliary phase in cirrhotically remodeled liver parenchyma due to altered pharmacokinetics affecting the contrast agent behavior of HCC lesions. This illustrates the complexity of HCC diagnosis and underlines the importance of a comprehensive understanding of HCC and its behaviors in various physiological contexts.

**Author Contributions:** Conceptualization, N.V. and J.R.; methodology, C.F.; validation, I.E., M.D., and L.L.; formal analysis, N.V.; investigation, A.G. and K.U.; resources, C.S.; data curation, A.G. and N.V.; writing—original draft preparation, A.G. and N.V.; writing—review and editing, L.S.A., W.U. and M.H.; visualization, N.V.; supervision, M.H.; project administration, M.H. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Institutional Review Board Statement:** The study was conducted in accordance with the Declaration of Helsinki and was approved by the Institutional Review Board (or Ethics Committee) of University Hospital Regensburg (protocol code 18-1268-104 and date of approval: 16 January 2019).

**Informed Consent Statement:** Patient consent was waived due to the retrospective design of the study.

**Data Availability Statement:** The data presented in this study are available on request from the corresponding author.

**Conflicts of Interest:** The authors declare no conflicts of interest.

#### **References**

- <span id="page-7-0"></span>1. Leong, T.Y.; Leong, A.S. Epidemiology and carcinogenesis of hepatocellular carcinoma. *HPB* **2005**, *7*, 5–15. [\[CrossRef\]](https://doi.org/10.1080/13651820410024021)
- 2. El-Serag, H.B. Epidemiology of hepatocellular carcinoma. *Clin. Liver Dis.* **2001**, *5*, 87–107. [\[CrossRef\]](https://doi.org/10.1016/S1089-3261(05)70155-0)
- 3. Fattovich, G.; Stroffolini, T.; Zagni, I.; Donato, F. Hepatocellular carcinoma in cirrhosis: Incidence and risk factors. *Gastroenterology* **2004**, *127*, S35–S50. [\[CrossRef\]](https://doi.org/10.1053/j.gastro.2004.09.014)
- <span id="page-7-1"></span>4. El-Serag, H.B. Hepatocellular Carcinoma. *N. Engl. J. Med.* **2011**, *365*, 1118–1127. [\[CrossRef\]](https://doi.org/10.1056/NEJMra1001683)
- <span id="page-7-2"></span>5. Llovet, J.M.; Zucman-Rossi, J.; Pikarsky, E.; Sangro, B.; Schwartz, M.; Sherman, M.; Gores, G. Hepatocellular carcinoma. *Nat. Rev. Dis. Primers* **2016**, *2*, 16018. [\[CrossRef\]](https://doi.org/10.1038/nrdp.2016.18)
- <span id="page-7-3"></span>6. Tarao, K.; Nozaki, A.; Ikeda, T.; Sato, A.; Komatsu, H.; Komatsu, T.; Taguri, M.; Tanaka, K. Real impact of liver cirrhosis on the development of hepatocellular carcinoma in various liver diseases-meta-analytic assessment. *Cancer Med.* **2019**, *8*, 1054–1065. [\[CrossRef\]](https://doi.org/10.1002/cam4.1998)
- <span id="page-8-0"></span>7. Bruix, J.; Sherman, M. Management of hepatocellular carcinoma. *Hepatology* **2005**, *42*, 1208–1236. [\[CrossRef\]](https://doi.org/10.1002/hep.20933)
- <span id="page-8-1"></span>8. Leitlinienprogramm Onkologie. *Diagnostik und Therapie des hepatozellulären Karzinoms, Langversion 1.0*; AWMF Registrierungsnummer: 032-053OL; AWMF: Frankfurt am Main, Germany, 2013.
- <span id="page-8-2"></span>9. Hammerstingl, R.; Huppertz, A.; Breuer, J.; Balzer, T.; Blakeborough, A.; Carter, R.; Fuste, L.C.; Heinz-Peer, G.; Judmaier, W.; Laniado, M.; et al. Diagnostic efficacy of gadoxetic acid (Primovist)-enhanced MRI and spiral CT for a therapeutic strategy: Comparison with intraoperative and histopathologic findings in focal liver lesions. *Eur. Radiol.* **2008**, *18*, 457–467. [\[CrossRef\]](https://doi.org/10.1007/s00330-007-0716-9)
- <span id="page-8-3"></span>10. Semelka, R.C.; Martin, D.R.; Balci, C.; Lance, T. Focal liver lesions: Comparison of dual-phase CT and multisequence multiplanar MR imaging including dynamic gadolinium enhancement. *J. Magn. Reson. Imaging* **2001**, *13*, 397–401. [\[CrossRef\]](https://doi.org/10.1002/jmri.1057)
- <span id="page-8-4"></span>11. Armengol, C.; Sarrias, M.R.; Sala, M. Hepatocellular carcinoma: Present and future. *Med. Clín.* **2018**, *150*, 390–397. [\[CrossRef\]](https://doi.org/10.1016/j.medcle.2017.08.024)
- <span id="page-8-5"></span>12. Ryder, S.D.; British Society of, G. Guidelines for the diagnosis and treatment of hepatocellular carcinoma (HCC) in adults. *Gut* **2003**, *52* (Suppl. S3), iii1–iii8. [\[CrossRef\]](https://doi.org/10.1136/gut.52.suppl_3.iii1)
- <span id="page-8-6"></span>13. Ichikawa, T.; Saito, K.; Yoshioka, N.; Tanimoto, A.; Gokan, T.; Takehara, Y.; Kamura, T.; Gabata, T.; Murakami, T.; Ito, K.; et al. Detection and characterization of focal liver lesions: A Japanese phase III, multicenter comparison between gadoxetic acid disodium-enhanced magnetic resonance imaging and contrast-enhanced computed tomography predominantly in patients with hepatocellular carcinoma and chronic liver disease. *Investig. Radiol.* **2010**, *45*, 133–141. [\[CrossRef\]](https://doi.org/10.1097/RLI.0b013e3181caea5b)
- <span id="page-8-7"></span>14. Vogl, T.J.; Kummel, S.; Hammerstingl, R.; Schellenbeck, M.; Schumacher, G.; Balzer, T.; Schwarz, W.; Muller, P.K.; Bechstein, W.O.; Mack, M.G.; et al. Liver tumors: Comparison of MR imaging with Gd-EOB-DTPA and Gd-DTPA. *Radiology* **1996**, *200*, 59–67. [\[CrossRef\]](https://doi.org/10.1148/radiology.200.1.8657946)
- <span id="page-8-8"></span>15. Leonhardt, M.; Keiser, M.; Oswald, S.; Kuhn, J.; Jia, J.; Grube, M.; Kroemer, H.K.; Siegmund, W.; Weitschies, W. Hepatic uptake of the magnetic resonance imaging contrast agent Gd-EOB-DTPA: Role of human organic anion transporters. *Drug Metab. Dispos.* **2010**, *38*, 1024–1028. [\[CrossRef\]](https://doi.org/10.1124/dmd.110.032862)
- <span id="page-8-9"></span>16. Nies, A.T.; Konig, J.; Pfannschmidt, M.; Klar, E.; Hofmann, W.J.; Keppler, D. Expression of the multidrug resistance proteins MRP2 and MRP3 in human hepatocellular carcinoma. *Int. J. Cancer* **2001**, *94*, 492–499. [\[CrossRef\]](https://doi.org/10.1002/ijc.1498)
- <span id="page-8-10"></span>17. Verloh, N.; Utpatel, K.; Haimerl, M.; Zeman, F.; Fellner, C.; Fichtner-Feigl, S.; Teufel, A.; Stroszczynski, C.; Evert, M.; Wiggermann, P. Liver fibrosis and Gd-EOB-DTPA-enhanced MRI: A histopathologic correlation. *Sci. Rep.* **2015**, *5*, 15408. [\[CrossRef\]](https://doi.org/10.1038/srep15408)
- <span id="page-8-11"></span>18. Zollner, G.; Wagner, M.; Fickert, P.; Silbert, D.; Fuchsbichler, A.; Zatloukal, K.; Denk, H.; Trauner, M. Hepatobiliary transporter expression in human hepatocellular carcinoma. *Liver Int.* **2005**, *25*, 367–379. [\[CrossRef\]](https://doi.org/10.1111/j.1478-3231.2005.01033.x)
- <span id="page-8-12"></span>19. Di Martino, M.; De Filippis, G.; De Santis, A.; Geiger, D.; Del Monte, M.; Lombardo, C.V.; Rossi, M.; Corradini, S.G.; Mennini, G.; Catalano, C. Hepatocellular carcinoma in cirrhotic patients: Prospective comparison of US, CT and MR imaging. *Eur. Radiol.* **2013**, *23*, 887–896. [\[CrossRef\]](https://doi.org/10.1007/s00330-012-2691-z)
- 20. Kudo, M. Multistep human hepatocarcinogenesis: Correlation of imaging with pathology. *J. Gastroenterol.* **2009**, *44* (Suppl. S19), 112–118. [\[CrossRef\]](https://doi.org/10.1007/s00535-008-2274-6)
- 21. Matsui, O.; Kobayashi, S.; Sanada, J.; Kouda, W.; Ryu, Y.; Kozaka, K.; Kitao, A.; Nakamura, K.; Gabata, T. Hepatocelluar nodules in liver cirrhosis: Hemodynamic evaluation (angiography-assisted CT) with special reference to multi-step hepatocarcinogenesis. *Abdom. Imaging* **2011**, *36*, 264–272. [\[CrossRef\]](https://doi.org/10.1007/s00261-011-9685-1)
- <span id="page-8-13"></span>22. Kitao, A.; Matsui, O.; Yoneda, N.; Kozaka, K.; Shinmura, R.; Koda, W.; Kobayashi, S.; Gabata, T.; Zen, Y.; Yamashita, T.; et al. The uptake transporter OATP8 expression decreases during multistep hepatocarcinogenesis: Correlation with gadoxetic acid enhanced MR imaging. *Eur. Radiol.* **2011**, *21*, 2056–2066. [\[CrossRef\]](https://doi.org/10.1007/s00330-011-2165-8)
- <span id="page-8-14"></span>23. Fletcher, C.D.; Unni, K.; Mertens, F. World Health Organization Classification of Tumours. In *Pathology and Genetics of Tumours of Soft Tissue and Bone*; IARC Press: Lyon, France, 2002.
- <span id="page-8-15"></span>24. Ishak, K.; Baptista, A.; Bianchi, L.; Callea, F.; De Groote, J.; Gudat, F.; Denk, H.; Desmet, V.; Korb, G.; MacSween, R.N.; et al. Histological grading and staging of chronic hepatitis. *J. Hepatol.* **1995**, *22*, 696–699. [\[CrossRef\]](https://doi.org/10.1016/0168-8278(95)80226-6)
- <span id="page-8-16"></span>25. De Carlis, L.; Giacomoni, A.; Lauterio, A.; Slim, A.; Sammartino, C.; Pirotta, V.; Colella, G.; Forti, D. Liver transplantation for hepatocellular cancer: Should the current indication criteria be changed? *Transpl. Int.* **2003**, *16*, 115–122. [\[CrossRef\]](https://doi.org/10.1111/j.1432-2277.2003.tb00272.x)
- <span id="page-8-21"></span>26. Durand, F.; Belghiti, J.; Paradis, V. Liver transplantation for hepatocellular carcinoma: Role of biopsy. *Liver Transpl.* **2007**, *13*, S17–S23. [\[CrossRef\]](https://doi.org/10.1002/lt.21326)
- <span id="page-8-22"></span>27. Jonas, S.; Bechstein, W.O.; Steinmuller, T.; Herrmann, M.; Radke, C.; Berg, T.; Settmacher, U.; Neuhaus, P. Vascular invasion and histopathologic grading determine outcome after liver transplantation for hepatocellular carcinoma in cirrhosis. *Hepatology* **2001**, *33*, 1080–1086. [\[CrossRef\]](https://doi.org/10.1053/jhep.2001.23561)
- <span id="page-8-17"></span>28. Tamura, S.; Kato, T.; Berho, M.; Misiakos, E.P.; O'Brien, C.; Reddy, K.R.; Nery, J.R.; Burke, G.W.; Schiff, E.R.; Miller, J.; et al. Impact of histological grade of hepatocellular carcinoma on the outcome of liver transplantation. *Arch. Surg.* **2001**, *136*, 25–30; discussion 31. [\[CrossRef\]](https://doi.org/10.1001/archsurg.136.1.25)
- <span id="page-8-18"></span>29. Lau, Y.Y.; Huang, Y.; Frassetto, L.; Benet, L.Z. Effect of OATP1B transporter inhibition on the pharmacokinetics of atorvastatin in healthy volunteers. *Clin. Pharmacol. Ther.* **2007**, *81*, 194–204. [\[CrossRef\]](https://doi.org/10.1038/sj.clpt.6100038)
- <span id="page-8-19"></span>30. Nakakariya, M.; Shimada, T.; Irokawa, M.; Maeda, T.; Tamai, I. Identification and species similarity of OATP transporters responsible for hepatic uptake of beta-lactam antibiotics. *Drug Metab. Pharmacokinet.* **2008**, *23*, 347–355. [\[CrossRef\]](https://doi.org/10.2133/dmpk.23.347)
- <span id="page-8-20"></span>31. Abe, T.; Unno, M.; Onogawa, T.; Tokui, T.; Kondo, T.N.; Nakagomi, R.; Adachi, H.; Fujiwara, K.; Okabe, M.; Suzuki, T.; et al. LST-2, a human liver-specific organic anion transporter, determines methotrexate sensitivity in gastrointestinal cancers. *Gastroenterology* **2001**, *120*, 1689–1699. [\[CrossRef\]](https://doi.org/10.1053/gast.2001.24804)
- <span id="page-9-0"></span>32. Tirona, R.G.; Leake, B.F.; Wolkoff, A.W.; Kim, R.B. Human organic anion transporting polypeptide-C (SLC21A6) is a major determinant of rifampin-mediated pregnane X receptor activation. *J. Pharmacol. Exp. Ther.* **2003**, *304*, 223–228. [\[CrossRef\]](https://doi.org/10.1124/jpet.102.043026)
- <span id="page-9-1"></span>33. Smith, N.F.; Acharya, M.R.; Desai, N.; Figg, W.D.; Sparreboom, A. Identification of OATP1B3 as a high-affinity hepatocellular transporter of paclitaxel. *Cancer Biol. Ther.* **2005**, *4*, 815–818. [\[CrossRef\]](https://doi.org/10.4161/cbt.4.8.1867)
- <span id="page-9-2"></span>34. Cillo, U.; Vitale, A.; Bassanello, M.; Boccagni, P.; Brolese, A.; Zanus, G.; Burra, P.; Fagiuoli, S.; Farinati, F.; Rugge, M.; et al. Liver transplantation for the treatment of moderately or well-differentiated hepatocellular carcinoma. *Ann. Surg.* **2004**, *239*, 150–159. [\[CrossRef\]](https://doi.org/10.1097/01.sla.0000109146.72827.76)
- <span id="page-9-3"></span>35. DuBay, D.; Sandroussi, C.; Sandhu, L.; Cleary, S.; Guba, M.; Cattral, M.S.; McGilvray, I.; Ghanekar, A.; Selzner, M.; Greig, P.D.; et al. Liver transplantation for advanced hepatocellular carcinoma using poor tumor differentiation on biopsy as an exclusion criterion. *Ann. Surg.* **2011**, *253*, 166–172. [\[CrossRef\]](https://doi.org/10.1097/SLA.0b013e31820508f1)
- <span id="page-9-4"></span>36. Takamori, R.; Wong, L.L.; Dang, C.; Wong, L. Needle-tract implantation from hepatocellular cancer: Is needle biopsy of the liver always necessary? *Liver Transpl.* **2000**, *6*, 67–72. [\[CrossRef\]](https://doi.org/10.1002/lt.500060103)
- <span id="page-9-5"></span>37. Pawlik, T.M.; Gleisner, A.L.; Anders, R.A.; Assumpcao, L.; Maley, W.; Choti, M.A. Preoperative assessment of hepatocellular carcinoma tumor grade using needle biopsy: Implications for transplant eligibility. *Ann. Surg.* **2007**, *245*, 435–442. [\[CrossRef\]](https://doi.org/10.1097/01.sla.0000250420.73854.ad)
- <span id="page-9-6"></span>38. Huppertz, A.; Haraida, S.; Kraus, A.; Zech, C.J.; Scheidler, J.; Breuer, J.; Helmberger, T.K.; Reiser, M.F. Enhancement of focal liver lesions at gadoxetic acid-enhanced MR imaging: Correlation with histopathologic findings and spiral CT--initial observations. *Radiology* **2005**, *234*, 468–478. [\[CrossRef\]](https://doi.org/10.1148/radiol.2342040278)
- <span id="page-9-11"></span>39. Schelhorn, J.; Best, J.; Dechene, A.; Gobel, T.; Bertram, S.; Lauenstein, T.; Kinner, S. Evaluation of combined Gd-EOB-DTPA and gadobutrol magnetic resonance imaging for the prediction of hepatocellular carcinoma grading. *Acta Radiol.* **2016**, *57*, 932–938. [\[CrossRef\]](https://doi.org/10.1177/0284185115616293)
- 40. Fujita, M.; Yamamoto, R.; Fritz-Zieroth, B.; Yamanaka, T.; Takahashi, M.; Miyazawa, T.; Tatsuta, M.; Terada, N.; Hosomi, N.; Inoue, E.; et al. Contrast enhancement with Gd-EOB-DTPA in MR imaging of hepatocellular carcinoma in mice: A comparison with superparamagnetic iron oxide. *J. Magn. Reson. Imaging* **1996**, *6*, 472–477. [\[CrossRef\]](https://doi.org/10.1002/jmri.1880060310)
- 41. Kim, S.H.; Kim, S.H.; Lee, J.; Kim, M.J.; Jeon, Y.H.; Park, Y.; Choi, D.; Lee, W.J.; Lim, H.K. Gadoxetic acid-enhanced MRI versus triple-phase MDCT for the preoperative detection of hepatocellular carcinoma. *AJR Am. J. Roentgenol.* **2009**, *192*, 1675–1681. [\[CrossRef\]](https://doi.org/10.2214/AJR.08.1262)
- 42. Chang, W.C.; Chen, R.C.; Chou, C.T.; Lin, C.Y.; Yu, C.Y.; Liu, C.H.; Chou, J.M.; Hsu, H.H.; Huang, G.S. Histological grade of hepatocellular carcinoma correlates with arterial enhancement on gadoxetic acid-enhanced and diffusion-weighted MR images. *Abdom. Imaging* **2014**, *39*, 1202–1212. [\[CrossRef\]](https://doi.org/10.1007/s00261-014-0168-z)
- <span id="page-9-7"></span>43. Choi, J.W.; Lee, J.M.; Kim, S.J.; Yoon, J.H.; Baek, J.H.; Han, J.K.; Choi, B.I. Hepatocellular carcinoma: Imaging patterns on gadoxetic acid-enhanced MR Images and their value as an imaging biomarker. *Radiology* **2013**, *267*, 776–786. [\[CrossRef\]](https://doi.org/10.1148/radiol.13120775)
- <span id="page-9-8"></span>44. Frericks, B.B.; Loddenkemper, C.; Huppertz, A.; Valdeig, S.; Stroux, A.; Seja, M.; Wolf, K.J.; Albrecht, T. Qualitative and quantitative evaluation of hepatocellular carcinoma and cirrhotic liver enhancement using Gd-EOB-DTPA. *AJR Am. J. Roentgenol.* **2009**, *193*, 1053–1060. [\[CrossRef\]](https://doi.org/10.2214/AJR.08.1946)
- 45. Huang, K.; Dong, Z.; Cai, H.; Huang, M.; Peng, Z.; Xu, L.; Jia, Y.; Song, C.; Li, Z.P.; Feng, S.T. Imaging biomarkers for well and moderate hepatocellular carcinoma: Preoperative magnetic resonance image and histopathological correlation. *BMC Cancer* **2019**, *19*, 364. [\[CrossRef\]](https://doi.org/10.1186/s12885-019-5574-8)
- <span id="page-9-12"></span>46. Tsuboyama, T.; Onishi, H.; Kim, T.; Akita, H.; Hori, M.; Tatsumi, M.; Nakamoto, A.; Nagano, H.; Matsuura, N.; Wakasa, K. Hepatocellular carcinoma: Hepatocyte-selective enhancement at gadoxetic acid–enhanced MR imaging—Correlation with expression of sinusoidal and canalicular transporters and bile accumulation. *Radiology* **2010**, *255*, 824–833. [\[CrossRef\]](https://doi.org/10.1148/radiol.10091557)
- <span id="page-9-9"></span>47. Narita, M.; Hatano, E.; Arizono, S.; Miyagawa-Hayashino, A.; Isoda, H.; Kitamura, K.; Taura, K.; Yasuchika, K.; Nitta, T.; Ikai, I.; et al. Expression of OATP1B3 determines uptake of Gd-EOB-DTPA in hepatocellular carcinoma. *J. Gastroenterol.* **2009**, *44*, 793–798. [\[CrossRef\]](https://doi.org/10.1007/s00535-009-0056-4)
- <span id="page-9-10"></span>48. Abe, T.; Kakyo, M.; Tokui, T.; Nakagomi, R.; Nishio, T.; Nakai, D.; Nomura, H.; Unno, M.; Suzuki, M.; Naitoh, T.; et al. Identification of a novel gene family encoding human liver-specific organic anion transporter LST-1. *J. Biol. Chem.* **1999**, *274*, 17159–17163. [\[CrossRef\]](https://doi.org/10.1074/jbc.274.24.17159)
- <span id="page-9-13"></span>49. Tamada, T.; Ito, K.; Higaki, A.; Yoshida, K.; Kanki, A.; Sato, T.; Higashi, H.; Sone, T. Gd-EOB-DTPA-enhanced MR imaging: Evaluation of hepatic enhancement effects in normal and cirrhotic livers. *Eur. J. Radiol.* **2011**, *80*, e311–e316. [\[CrossRef\]](https://doi.org/10.1016/j.ejrad.2011.01.020)
- <span id="page-9-14"></span>50. Chernyak, V.; Fowler, K.J.; Kamaya, A.; Kielar, A.Z.; Elsayes, K.M.; Bashir, M.R.; Kono, Y.; Do, R.K.; Mitchell, D.G.; Singal, A.G.; et al. Liver Imaging Reporting and Data System (LI-RADS) Version 2018: Imaging of Hepatocellular Carcinoma in At-Risk Patients. *Radiology* **2018**, *289*, 816–830. [\[CrossRef\]](https://doi.org/10.1148/radiol.2018181494)
- <span id="page-9-15"></span>51. Hamilton, S.R. World Health Organization classification of tumours. In *Pathology and Genetics of Tumours of the Digestive System*; IARC Press: Lyon, France, 2000.
- <span id="page-9-16"></span>52. Kenmochi, K.; Sugihara, S.; Kojiro, M. Relationship of histologic grade of hepatocellular carcinoma (HCC) to tumor size, and demonstration of tumor cells of multiple different grades in single small HCC. *Liver* **1987**, *7*, 18–26. [\[CrossRef\]](https://doi.org/10.1111/j.1600-0676.1987.tb00310.x)

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